

US EPA ARCHIVE DOCUMENT

## DATA EVALUATION RECORD

§ 72-3(B) -- ACUTE EC<sub>50</sub> TEST WITH AN ESTUARINE/MARINE MOLLUSK SHELL DEPOSITION STUDY

1. CHEMICAL: 2Chloro-4,6-bis(isopropylamino)-s-trizine  
PC Code No.: 080808

2. TEST MATERIAL: Propazine Purity: 98%

3. CITATION Authors: R. L. Boero; P. L. Kowalski; T. J. Ward  
Title: Flow through shell deposition test with propazine and the eastern oyster  
Study Completion Date: 8/4/95  
Laboratory: T.R. Wilbury Laboratories, Inc./ABC Laboratories, Inc.  
Sponsor: Griffin Corporation  
Laboratory Report ID: ABC Laboratories #41956  
MRID No.: 442873-06  
DP Barcode: D237791

4. REVIEWED BY: Thomas M. Steeger, Ph.D., Fishery Biologist, EFED, ERB IV, U.S. EPA

Signature: *Thomas M Steeger*

Date: 10/14/97

5. APPROVED BY: Ann Stavola, Aquatic Biologist, EFED, ERB IV, U.S. EPA

Signature: *Ann Stavola*

Date: 10/15/98

6. STUDY PARAMETERS

Scientific Name of Test Organism: *Crassostrea virginica*  
Age or Size of Test Organism: juvenile  
Definitive Test Duration: 96 hours  
Study Method: Flow-through  
Type of Concentrations: Mean measured/Nominal

7. CONCLUSIONS: This study was not scientifically sound and does not fulfill guideline requirements. Precipitant was observed in the two highest treatment groups, 3.0 and 5.0 mg a.i./L. Recovery of Propazine ranged between 74 - 79% and underscores that solubility may have been problematic in all treatment groups. Thus the conclusion that the EC<sub>50</sub> was greater than 3.72 ppm is not warranted as the oysters may not have been exposed to these levels due to the reduced solubility of Propazine in salt water.



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**Results Synopsis**

EC<sub>50</sub>: >3.72 ppm ai

95% C.I.: \_\_\_\_\_ ppm ai

NOEL: >3.72 ppm ai

Probit Slope: \_\_\_\_\_

**8. ADEQUACY OF THE STUDY**

**A. Classification: INVALID**

**B. Rationale:** Conventional centrifugation is required for all test media where undissolved test material or precipitate is observed in the test chambers or where the solubility and hence bioavailability are in question. Filtration may be used instead of centrifugation if the analytical method is validated over a range of acceptable concentrations. For aquatic toxicity tests, the Environmental Fate and Effects Division defines solubility as the extractable chemical in the centrifuged or filtered supernate (EPA 1994).

Additionally, unfiltered sea water was used however data on complete water analysis is not provided. Total organic carbon is not reported.

**C. Repairability: none**

**9. BACKGROUND**

**10. GUIDELINE DEVIATIONS**

1. Salinity of dilution on Day 0 of acclimation was lower than the test salinity.

2. Samples at 0 hrs were stored in refrigerator until end of Definitive test rather than being shipped immediately

**11. SUBMISSION PURPOSE:** To determine the effect of Propazine to oysters by determining the median effective concentration of Propazine causing a 50% decrease in shell deposition (compared to a control) after a specified exposure period and to examine the sublethal/behavioral responses during the course of the study.

12. MATERIALS AND METHODS

A. Test Organisms

Guideline Criteria	Reported Information
<u>Species</u> Preferred species are the Pacific oyster ( <i>Crassostrea gigas</i> ) and the Eastern oyster ( <i>Crassostrea virginica</i> )	<i>Crassostrea virginica</i>
<u>Mean valve height</u> 25 - 50 mm along the long axis	25 - 50 mm
<u>Supplier</u>	P. Cummins Oyster Co.
Are all oysters from same source?	Yes
Are all oysters from the same year class?	Not reported

B. Source/Acclimation

Guideline Criteria	Reported Information
<u>Acclimation Period</u> Minimum 10 days	10 days
Wild caught organisms were quarantined for 7 days?	N/A
Were there signs of disease or injury?	No
If treated for disease, was there no sign of the disease remaining during the 48 hours prior to testing?	N/A
<u>Amount of peripheral shell growth removed prior to testing</u>	3-5 mm

Guideline Criteria	Reported Information
<b><u>Feeding during the acclimation</u></b> Must be fed to avoid stress.	Yes
<b><u>Pretest Mortality</u></b> <3% mortality 48 hours prior to testing	< 3% mortality prior to testing.

## C. Test System

Guideline Criteria	Reported Information
<b><u>Source of dilution water</u></b> Natural unfiltered seawater from an uncontaminated source.	Unfiltered natural sea water
<b><u>Does water support test animals without observable signs of stress?</u></b>	Yes
<b><u>Salinity</u></b> 30-34 ‰ (parts per thousand) salinity, weekly range < 6 ‰	33‰
<b><u>Water Temperature</u></b> 15°-30° C, consistent in all test vessels	18.1 - 19.3°C
<b><u>pH</u></b>	7.7 - 8.5
<b><u>Dissolved Oxygen</u></b> ≥ 60% throughout	Range: 7.2 - 8.0 mg/L
<b><u>Total Organic Carbon</u></b>	not reported
<b><u>Test Aquaria</u></b> Should be constructed of glass or stainless steel.	Glass
<b><u>Type of Dilution System</u></b> Must provide reproducible supply of toxicant	proportional diluter delivering 0.3 ml stock into 3,000 ml of dilution water
<b><u>Flow rate</u></b> Consistent flow rate	8.3 vol/24 hours

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Guideline Criteria	Reported Information
Was the loading of organism such that each individual sits on the bottom with water flowing freely around it?	Yes
<u>Photoperiod</u> 16 hours light, 8 hours dark	16 hours light, 8 hours dark
<u>Solvents</u> Not to exceed 0.5 ml/L	Solvent: Dimethylformamide Maximum conc.: 0.1 ml/L

D. Test Design

Guideline Criteria	Reported Information
<u>Range Finding Test</u> If EC <sub>50</sub> >100 mg/L with 30 or more oysters, then no definitive test is required.	Not performed; relied on historical data
<u>Nominal Concentrations of Definitive Test</u> Control & 5 treatment levels; each conc. should be 60% of the next highest conc.; concentrations should be in a geometric series	0, 0.75, 1.3, 2.0, 3.0, 5.0 mg ai/L
<u>Number of Test Organisms</u> Minimum 20 individual per test level and in each control	20
Test organisms randomly or impartially assigned to test vessels?	Yes
Biological observations made every 24 hours?	Yes

Guideline Criteria	Reported Information
<p><b>Water Parameter Measurements</b></p> <p>1. <u>Temperature</u> Measured hourly in at least one chamber.</p> <p>2. <u>DO and pH</u> Measured at beginning of test and every 48 h in the high, medium, and low doses and in the control</p>	<p>1. Temperature recorded daily</p> <p>2. DO and pH recorded daily</p>
<p>Was chemical analysis performed to determine the concentration of the test material at the beginning and end of the test? (Optional)</p>	<p>Yes</p>

13. REPORTED RESULTS

A. General Results

Guideline Criteria	Reported Information
<p>Quality assurance and GLP compliance statements were included in the report?</p>	<p>Yes</p>
<p><u>Control Mortality</u> Not more than 10% of control organisms may die or show abnormal behavior.</p>	<p>0%</p>
<p><u>Control Shell Deposition</u> Must be at least 2 mm.</p>	<p>2.7 mm</p>
<p><u>Recovery of Chemical</u></p>	<p>88 - 104%</p>
<p>Raw data included?</p>	<p>Yes</p>
<p>Signs of toxicity (if any) were described?</p>	<p>N/A</p>

Shell Growth

Concentration (ppm)		Number Per Level	Number Dead	Mean Shell Deposition (mm)	Mean Percent Reduction
Nominal	Mean Measured				
Control	<0.167	20	0	2.7 ± 0.8	--
Solvent Control	<0.167	20	0	2.9 ± 1.0	--
0.75	0.584	20	0	2.9 ± 0.7	ns
1.33	1.00	20	0	2.6 ± 0.6	ns
2.0	1.58	20	0	2.9 ± 0.9	ns
3.0	2.33	20	0	2.4 ± 0.8	ns
5.0	3.72	20	0	2.7 ± 0.9	ns

**B. Statistical Results**

Method:

96-hr EC<sub>50</sub>: >3.72 ppm ai                      95% C.I.: \_\_\_ - \_\_\_ ppm ai

Probit Slope: \_\_\_                                      NOEC: 3.72 ppm ai

**14. VERIFICATION OF STATISTICAL RESULTS**

Parameter	Result
Statistical Method for EC <sub>50</sub>	computerized EC <sub>50</sub> program (Stephan 1983)
EC <sub>50</sub> (95% C.I.)	ppm ai
Probit Slope	N/A
Statistical Method for NOEC	
NOEC	ppm ai

**15. REVIEWER'S COMMENTS:**

Measured concentrations of propazine ranged from 74 - 79% of nominal. It is not clear whether the differences between test and nominal concentrations was a result of solubility. Propazine solubility reported to be 8.6 mg/L; in two of the treatments, i.e., 3.0 and 5.0 mg/L, there was a precipitate. It is interesting to note that the difference between nominal and test



concentrations was roughly 25% lower than in treatments involving bluegill.

Conventional centrifugation is required for all test media where undissolved test material or precipitate is observed in the test chambers or where the solubility and hence bioavailability are in question. Filtration may be used instead of centrifugation if the analytical method is validated over a range of acceptable concentrations. For aquatic toxicity tests, the Environmental Fate and Effects Division defines solubility as the extractable chemical in the centrifuged or filtered supernate (EPA 1994).

Unfiltered sea water was used for the exposure system and Table 1 provides a characterization of the dilution water; however, total organic carbon analysis is not included. Propazine is capable of adsorbing to organic matter thus rendering the herbicide less bioavailable; however, the compound is more likely to adsorb in its protonated form. At the pH of the test system (~ 8.0), Propazine would be predominately in its unionized form and less like to adsorb. The information on total organic carbon should be provided though.

#### References

EPA 1994. Pesticide reregistration rejection rate analysis: ecological effects. Office of Prevention, Pesticides, and Toxic Substances Publication EPA 738-R-94-035, Washington D. C.