US ERA ARCHIVE DOCUMENT

DATA EVALUATION RECORD

PXTS (Polymeric xylenol tetrasulfide) MRID 46062617

Bacterial Reverse Mutation Test OPPTS 870.5100

Prepared for

Antimicrobial Division Office of Pesticide Programs U.S. Environmental Protection Agency 1921 Jefferson Davis Highway Arlington, VA 22202

Prepared by

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Under Subcontract to

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EPA WAM:

Killian Swift, Ph.D.

This review may have been altered by EPA subsequent to the contractors' signatures above.

EPA Reviewer: <u>Tim McMahon</u> Senior Toxicologist, Antimicrobial Division (7510C) Signature Date 711/04

DATA EVALUATION RECORD

STUDY TYPE: In vitro Bacterial Gene Mutation (Bacterial system, Salmonella typhimurium; E. coli)/ mammalian activation gene mutation assay; OPPTS 870.5100 [§84-2]; OECD 471 (formerly OECD 471 & 472).

PC CODE: 006929

DP BARCODE: D299112

TEST MATERIAL (PURITY): Polymeric xylenol tetrasulfide (100% a.i.)

SYNONYMS: PXTS

CITATION: Stankowski Jr., L.F. (2002) Salmonella-Escherichia coli/Mammalian-Microsome

Reverse Mutation Assay with a Confirmatory Assay with PXTS. Covance

Laboratories Inc., Vienna, Virginia. Study Number 23134-0-409OECD. May 13,

2002. MRID 46062617, Unpublished.

SPONSOR: Akzo Nobel Functional Chemicals Inc.

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EXECUTIVE SUMMARY: In a reverse gene mutation assay in bacteria using the plate-incorporation protocol (MRID 46062617), strains TA98, TA100, TA1535, TA1537, and WP2uvrA were exposed to PXTS (100% a.i., Batch# 6, bottle no. 1 and lot 1685-11-1). Concentrations in the main reversion assay were determined by a preliminary toxicity test with TA100 and WP2uvrA (6.67-5000 μg/plate, ±S9). S. typhimurium strains were incubated with concentrations of 3.33, 10.0, 33.3, 100, 333, and 500 μg/plate in the presence of mammalian metabolic activation (from Sprague-Dawley rats induced with Aroclor 1254; +S9) and 1.00, 3.33, 10.0, 33.3, 50.0, and 66.7 μg/plate in the absence of mammalian metabolic activation (-S9). WP2uvrA was exposed to 33.3, 100, 333, 667, 1000, and 2000 μg/plate, +S9, and 10.0, 33.3, 100, 333, 667, and 1000 μg/plate, -S9. Because of a slight increase in revertants at 500 μg/plate in TA100, +S9, the dose range was shifted up in the confirmatory assay for the Salmonella strains (10-1000 μg/plate). For the -S9 confirmatory assay, the dose range was shifted down to a range of 0.333-50 μg/plate due to cytotoxicity observed at 50 μg/plate and higher in TA100 in the first assay. The confirmatory assay in WP2uvrA, +S9, used a dose range of 100-3330 μg/plate, while that for the -S9 condition used a range of 10-667 μg/plate.

PXTS was tested up to cytotoxic concentrations. Severe decreases in spontaneous revertants were noted in TA100 at ≥33.3 μg/plate and in WP2μνγA at ≥667 μg/plate, both -S9. In the initial



PXTS was tested up to cytotoxic concentrations. Severe decreases in spontaneous revertants were noted in TA100 at ≥ 33.3 µg/plate and in WP2uvrA at ≥ 667 µg/plate, both -S9. In the initial mutagenicity assay, background lawn inhibition was observed in strains TA98, TA100, and TA1537 at concentrations ≥ 333 µg/plate (+S9), in TA100 at ≥ 10.0 µg/plate (-S9), in all Salmonellu strains at doses ≥ 33.3 µg/plate (-S9), and in WP2uvrA at concentrations of 2000 µg/plate (+S9) and ≥ 100 µg/plate (-S9). In the confirmatory assay, background growth inhibition was observed at doses of 333 µg/plate, +S9 in TA100 and TA1537, in all Salmonella strains at ≥ 500 µg/plate, and at ≥ 1000 µg/plate in WP2uvrA. Reduced or absent background lawns were noted at ≥ 3.33 µg/plate (TA100), ≥ 33.3 µg/plate (all Salmonella strains), and ≥ 100 µg/plate (WP2uvrA), -S9. Precipitation of the test article was generally noted at doses of 333 µg/plate or higher in all strains, ±S9. The positive controls induced appropriate responses in each strain. There was no evidence of induced mutant colonies over background or a concentration-related positive response. Therefore, the test article is considered negative in the bacterial reversion assay.

This study is classified as ACCEPTABLE (GUIDELINE) and satisfies the guideline requirement for Test Guideline OPPTS 870.5100; OECD 471 for in vitro mutagenicity (bacterial reverse gene mutation) data.

<u>COMPLIANCE</u>: Signed and dated GLP, Quality Assurance, and Data Confidentiality statements were provided. The homogeneity, stability, and concentrations of the test formulations were not verified.

I. MATERIALS AND METHODS

A. MATERIALS:

1. Test Material:

PXTS

Description:

Black solid

Lot/Batch #:

Lot No. 1685-11-1/ Batch No. 6, Bottle no. 1

Purity:

100 % a.i.

CAS # of TGAI:

Not provided Structure not available

Solvent Used:

Ethanol

2. Control Materials:

Negative:

Tester strain and S9 were plated alone as a check of sterility

Solvent (final conc'n):

Ethanol (100 µL)

Positive:

Nonactivation:

Sodium azide 2.0 µg/plate TA100, TA1535

2-Nitrofluorene 1.0 µg/plate TA98

Other (list):

ICR-191 2.0 µg/plate TA1537

4-Nitroquinoline-N-oxide 1.0 µg/plate WP2nvrA

Activation:

2-Aminoanthracene (2-anthramine) 2.5 µg/plate all S.

typhimurium strains

Other (list):

2-Aminoanthracene (2-anthramine) 25.0 µg/plate WP2

urrA

Benzo[a]pyrene 2.5 µg/plate TA98

3. Activation: S9 derived from

ACU X	induced	X	Aroclor 1254 (200 mg/mL corn oil)	Rat (male Sprague- Dawley)	X	Liver
-	pon-induced		Phenobarbitol	Mouse		Lung
			None	Hamster		Other
1			Other	Other		

S9 mix composition: The S9 homogenate was purchased from Molecular Toxicology, Inc. Two different lots were utilized: Lot 1324 (35.0 mg/mL protein) and Lot 1350 (39.2 mg/mL protein). Fresh S9 mix was prepared prior to each use. The S9 homogenate was mixed with 4 μmol NADP*, 5 μmol glucose-6-phosphate, 3.3 μmol KCl, 8 μmol MgCl₂, 100 μmol sodium phosphate buffer (pH 7.4), and water. The final S9 mix contained 10% S9 concentrate.

4. Test organisms: S. typhimurium strains

		The second secon						Acceptance and the second and the se	
ſ		TA97	X	TA98	Х	TA100		TA102	TAI04
ľ	X	TA1535	X	TA1537		TA1538	Х	list any others WP2uvrA	

Properly maintained?

X Yes

No



[Polymeric Ayleno] Tetrasulfic	
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X Yes

5. Test compound concentrations used:

Preliminary Cytotoxicity Test

Non-activated conditions: 6.67, 10.0, 33.3, 66.7, 100, 333, 667, 1000, 3330, and 5000 µg/plate

TA100 and WP2uvrA

Activated conditions: 6.67, 10.0, 33.3, 66.7, 100, 333, 667, 1000, 3330, and 5000 µg/plate

TA100 and WP2invrA

Main Assay (S. typhimurium strains)

Non-activated conditions: 1.00, 3.33, 10.0, 33.3, 50.0, and 66.7 µg/plate Activated conditions: 3.33, 10.0, 33.3, 100, 333, and 500 µg/plate

Main Assav (WP2uvrA)

Non-activated conditions: 33.3, 100, 333, 667, 1000, and 2000 µg/plate Activated conditions: 10.0, 33.3, 100, 333, 667, and 1000 µg/plate

Confirmatory Assay (S. typhimurium strains)

Non-activated conditions: 0.333, 1.00, 3.33, 10, 33.3, and 50.0 µg/plate Activated conditions: 10.0, 33.3, 100, 333, 500, and 1000 µg/plate

Confirmatory Assay (WP2uvrA)

Non-activated conditions: 10.0, 33.3, 100, 333, 500, and 667 µg/plate Activated conditions: 100, 333, 667, 1000, 2000, and 3330 µg/plate

One plate was prepared for each concentration, strain, and condition for the preliminary toxicity test. Three replicates for each strain, concentration, and condition were counted for the number of revertant colonies in the mutagenicity test. The solvent and positive control plates were assayed in triplicate as well.

B. TEST PERFORMANCE

1. Type of Salmonella assay:

X standard plate test

_ pre-incubation (_ minutes)

- "Prival" modification (i.e. azo-reduction method)
- _ spot test
- other
- 2. Protocol: Dose selection was based on the results of a preliminary toxicity test conducted for the test article in which ten doses were tested (6.7, 10.0, 33.3, 67.7, 100, 333, 667, 1000, 3330, and 5000 µg/plate). In the main assay, all the bacterial tester strains were tested in the absence and presence of metabolic activators for all test concentrations. The plate incorporation method of analysis was used in both the preliminary toxicity test and mutagenicity tests. An aliquot (100



μL) of the bacterial suspension of an overnight culture (0.5 × 10° cells/mL), 100 μL of the solvent or test article or 50 μL of the positive control, and 500 μL of S9 or 500 μL of water was added to 2.0 mL of the selective molten (45±2°C) top agar (0.7% [w/v] agar and 0.5% [w/v] NaCl supplemented with 0.5 mM histidine/biotin (Salmonella) or 0.5 mM tryptophan (E. coli). This mixture was vortexed and poured onto a petri dish containing 25 mL of minimal agar (Vogel-Bonner medium E supplemented with 1.5 w/v agar and 0.2% glucose) and was allowed to solidify. The cultures were inverted and incubated for 52±4 hours at 37 ± 2°C. Revertant colonies were counted by hand or by automatic colony counter from three replicate plates per strain, concentration, and condition. Plates that were not counted immediately after incubation were stored at 0.1-10°C. Both precipitate and background lawn were evaluated by eye and microscopically.

- 3. <u>Statistical Analysis</u>: Mean numbers of revertant colonies and their standard deviations were calculated for each strain, concentration, and condition. No statistical analyses were performed. The treatment of the data was considered to be appropriate.
- 4. Evaluation Criteria: A positive response in TA98, TA100, and WP2uvrA was defined as a dose-dependent, ≥2-fold increase in the mean number of revertants per plate relative to the control plates for at least one bacterial strain. A positive response in TA1535 and TA1537 was defined as a dose-dependent, ≥3-fold increase in mean revertants over the control values for at least one bacterial strain.

For the assay to be considered valid, the following criteria must be met: 1) strains TA98, TA100, TA1535, and TA1537 must display a sensitivity to crystal violet; 2) strains TA98 and TA100 must grow in culture broth supplemented with ampicillin (thereby demonstrating pKM101 plasmid presence); 3) at least 3 doses must be evaluated; 4) cell densities $\geq 0.5 \times 10^9$ bacteria/mL must be plated; 5) the positive control (\pm S9) must produce a at least a 3-fold increase in revertants over the corresponding vehicle control; and 6) all vehicle control cultures must produce a characteristic number of spontaneous revertants per plate within the historical control range (listed below).

Bacterial Strain	ı Rev	ertant Range
TA98		8-60
TA100		60-240
TA1535		4-45
TA1537		2-25
WP2uvrA		5-40

II. REPORTED RESULTS

A. PRELIMINARY CYTOTOXICITY ASSAY: Bacterial strains TA100 and WP2uvrA were exposed to 10 doses ranging from 6.67 to 5000 μ g/plate, \pm S9. Thinning or disappearance of the background lawn was observed in TA100 at concentrations \geq 33.3 (-S9) and \geq 333 μ g/plate (+S9). The background lawn was reduced in WP2uvrA strain at doses \geq 66.7, -S9, and \geq 667



μg/plate, +S9. Precipitation was noted at doses ≥333 (+S9) and ≥100 (-S9) μg/plate in the TA100 plates and 2667 µg/plate (±S9) in the WP2uvrA plates.

B. MUTAGENICITY ASSAY: The results of the initial mutation assay are presented in Table 1 below. Background lawn inhibition was observed in strains TA98, TA100, and TA1537 at concentrations ≥ 333 μg/plate (+S9), in TA100 at 10.0 μg/plate (-S9), in all Salmonella strains at doses ≥ 33.3 µg/plate (-S9), and in WP2uvrA at concentrations ≥ 100 µg/plate (-S9) and 2000 μg/plate (+S9). Concentrations ≥333 μg/plate (+S9) in TA98 and TA100, ≥333 μg/plate (+S9) in TA1535 and TA1537, and ≥667 µg/plate (±S9) in the WP2uvrA strain produced precipitation. The test article did not induce a significant increase in revertants in any strain, ±S9. A dosedependent increase (approximately 1.6-fold over the control values) was observed in TA100, +S9. This increase was considered by the study author to be a "statistical aberration due to random fluctuation of the spontaneous revertant frequency" and unrelated to the test article. The positive control compounds displayed an appropriate increase in revertants compared to the controls, demonstrating the sensitivity of the assay.

Table 1. Summary of Initial Mutagenicity Assay

T.	TA98	TA100	TA1535	TA1537	WP2 uvr	1
			+89			
	79±4	101≟4	19±8	11±2	Vehicle Control	17±2
Vehicle Control	37±6	106±12	23±3	12±2	33.3 µg/plate	23±3
3,33 µg/plate	38±6	104±9	19±3	9±3	100 μg/plate	21±2
10.0 μg/plate		114±5	19:4	9±4	333 µg/plate	30 ±6
33.3 µg/plate	41±6	115±15	19±2	13±3	667 μg/plate	24±2°
. 100 µg/plate	36±1 31±8 ^{b, c}	124±14 ^{b. 6}	13±3 [€]	10±6 ^{b,c}	1000 μg/plate	37 ±6 €
333 µg/plate		162±17 ^{b, c}	16±4°	7±6 ^{b.s}	2000 µg/plate	18±5 h ,1
500 µg/plate Positive Control	35±6 ^{5,6} 384±70	1093±184	146±17	113±13	Positive Control	380±19
			-S9			
Vehicle Control	21±5	91±17	16±2	6±2	Vehicle Control	16±3
	25±6	88±15	12±3	8±1	10.0 μg/plate	15±4
1.00 µg/plate	23±6	75±7	10±2	5#2	33.3 μg/plate	15±4
3.33 µg/plate	20±4	 	12=4	7±5	100 μg/plate	17±3
10.0 µg/plate		3±4"	5±0 ^h	5=20	333 µg/plate	18±8
33.3 µg/plate 50.0 µg/plate	14±5 ^b	0∓0, >=-	4±3'	2±26	667 μg/plate	1±2 ^b .



Olymeric Aylenet (et			TA1535	TA1537	WP2 m	rA .
	TA98	TA100			1000 µg/plate	0±0h.c
66.7 µg/plate	8±3 ^b	0±0"	2±3"	l±1		220±28
Positive Control	234±29	1131±63	826±18	781±268	Positive Control	

Data obtained from study Tables 4-5 (pp. 24-25).

The results of the confirmatory assay are presented in Table 2 below. Severe decreases in spontaneous revertants, +S9, were noted at \geq 333 µg/plate in TA1537, and at the top dose used in all strains, +S9. No revertants were observed in TA100 and TA1537 at 50 µg/plate, -S9, and severe reductions in spontaneous revertants were noted in WP2 uvrA at ≥333 μg/plate, -S9. Background lawn inhibition was observed at 333 μg/plate (TA100 and TA1537), 500 μg/plate (TA100, TA1535, and TA1537), 1000 μg/plate (all Salmonella strains), and ≥ 1000 μg/plate (WP2 wrA), +S9. Concentrations ≥3.33µg/plate (TA100), ≥33.3 µg/plate (all Salmonella strains) and 2100 µg/plate (WP2 uvrA), -S9, induced either a reduced or absent background lawn. Precipitation was observed at doses ≥ 500 µg/plate, +S9, in all Salmonella strains and at 3330 µg/plate, +S9, or 500 µg/plate, -S9, in WP2 uvrA. The test article did not induce a significant increase in revertants in any strain, ±S9.

Both the negative and positive cultures produced expected responses, consistent with historical control ranges in both experiments.

Table 2. Summary of Confirmatory Mutagenicity Assay

Ī	TA98	TA100	TA1535	TA1537	WP2 uvrA	
			+89			
. T	40±3	101±6	38±11	8±3	Vehicle Control	20±2
Vehicle Control		99±11	34±2	11±4	100 µg/plate	27±1
10.0 µg/plate	38±5		35±7	825	333 µg/plate	31±2
33.3 µg/plate	33=2	99±18	29±4	13±3	667 µg/plate	28±10
100 μg/plate	39±12	95±5		1±2	1000 μg/plate	25±9¢
333 µg/plate	39±9	73±13 ^b	25±7	070p.e	2000 μg/plate	19±11 ^b
500 μg/plate	38±7°	79±64 ^{b. s}	20±4 ^{b, €}			0+0p.c
1000 µg/plate	28±16 ^{b. c}	0±0c, q	8±7 ^{b, c}	0±0°-4	3330 µg/plate	
Positive Control	389±37	397±35	122±5	93±8	Positive Control	489±94
			-S9			
Vehicle Control	26+7	93±8	21±7	8≢2	Vehicle Control	15±8

Reduced background lawn

Precipitate

ymeric Nylenol Tetra				TA1537	WP2 uvrA		
	TA98	TA100	TA1535		10.0 µg/plate	14±3	
0.333 μg/plate	19±4	102=5	24±4	6.1.4		18±1	
	24±3	87±10	16±3	8±1	33.3 µg/plate	19±4 ^b	
1.00 µg/plate		99±5 ^b	18±4	8±1	100 µg/plate		
3.33 µg/plate	21±7		23±8	8±5	333 µg/plate	3±3 ^h	
10.0 μg/plate	22±4	93±11 ^b		7-2	500 μg/plate	0±1 ^{h, c} .s	
33.3 µg/plate	14±3 ^b	66±34 ^b	20±8 ^b		667 µg/plate	0+1p, c.s	
50.0 μg/plate	11±7 ^b	0±0 ^b	5±8 ^b	0±0p		181±25	
Positive Control	250±26	1049±70	792±94	1235±103	Positive Control		

^a Data obtained from study Tables 7-8 (pp. 27-28).

III. DISCUSSION and CONCLUSIONS

A. INVESTIGATORS' CONCLUSIONS: Based on the results of the initial and confirmatory mutagenicity assays, PXTS was considered not mutagenic in the absence and presence of metabolic activation.

B. REVIEWER COMMENTS: In the first reversion assay, a slight dose-dependent increase in mutant colonies was observed in TA100, +S9 at a dose of 500 µg/plate. This response did not meet the stated criteria for a positive response and was not observed in a confirmatory assay using a slightly higher dose range (10.0-1000 vs. 3.33-500 µg/plate). Based on these findings, the observed slight dose-dependent increase in revertants was appropriately concluded by the authors to not be a treatment-related response. Overall, the study was conducted appropriately and reported clearly, therefore, the study sufficiently evaluated the mutagenicity of PXTS.

C. STUDY DEFICIENCIES:

The homogeneity, stability, and concentration of the test formulations were not analyzed.

The protocol deviation was minor and would not adversely affect the outcome and interpretation of the study results.

D. STUDY CLASSIFICATION: This study is classified as ACCEPTABLE (GUIDELINE) and satisfies the guideline requirement for Test Guideline OPPTS 870.5100; OECD 471 for in vitro mutagenicity (bacterial reverse gene mutation) data.



h Reduced background lawn

e Precipitate

^{*}Actual mean and standard deviation were 0.33±0.578. The discrepancy in data reporting was due to rounding.