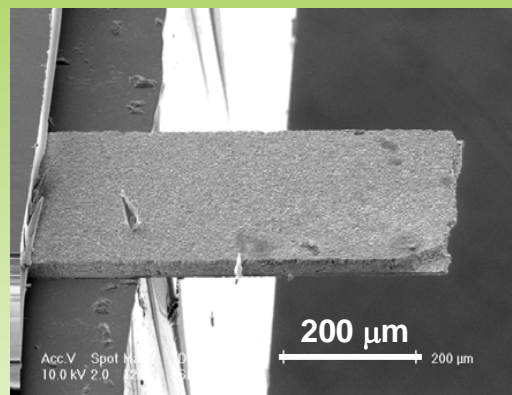
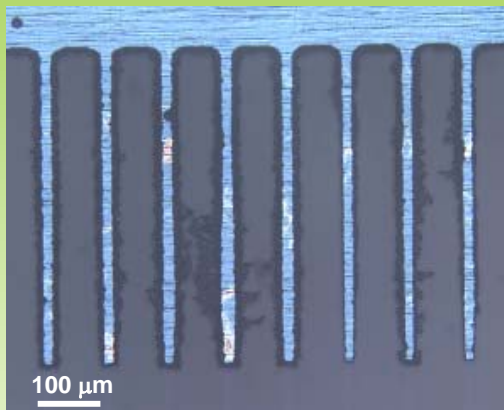


# Ultrasensitive Pathogen Quantification in Drinking Water Using High Piezoelectric PMN-PT Microcantilevers



Wan Y Shih and Wei-Heng Shih

Dept Materials Science and Engineering

Drexel University

August 18-20, 2004



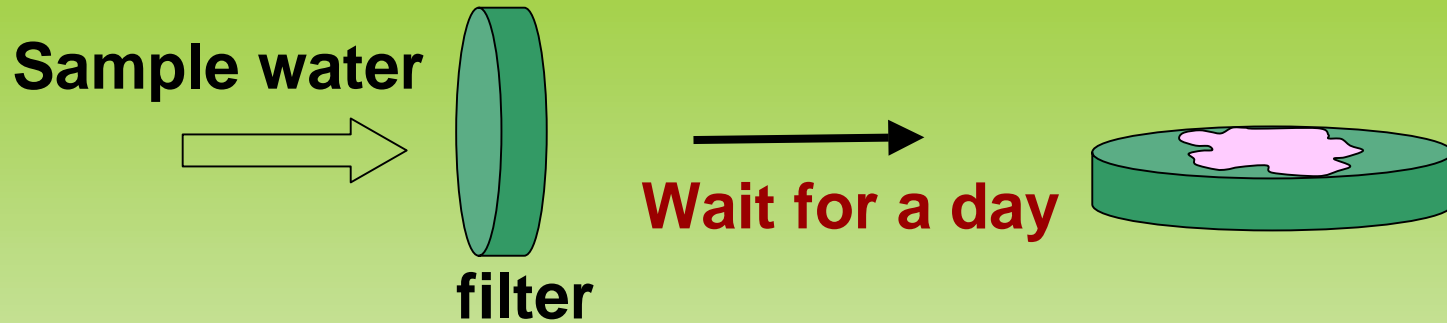
# E.coli 0157:H7



- E.coli 0157:H7 is the main cause of :
  - Foodborne illness
    - Contaminated meat – improperly cooked beef
    - Lettuce
    - Unpasteurized milk and juice
    - Salami
    - Sprouts
  - Waterborne illness
    - Creeks, rivers, streams, lakes, ground water, portable water and swimming pools

**E coli alone killed some 240 people in 2001!**

# Current pathogen detection technology



## Disadvantages:

- Slow
- Not real-time, in-situ
- Not quantitative

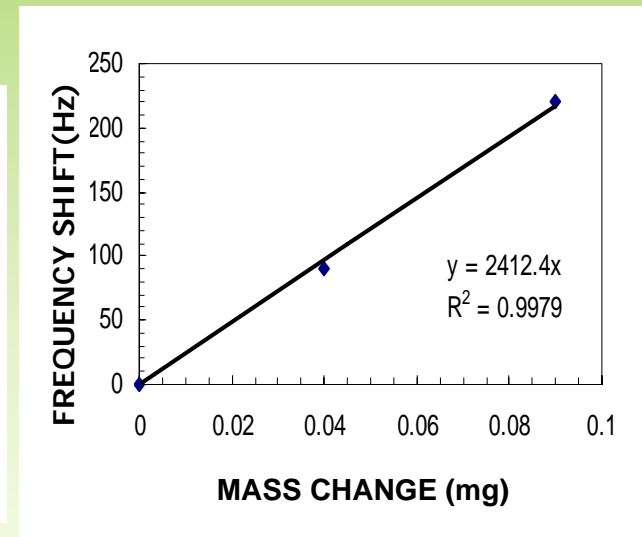
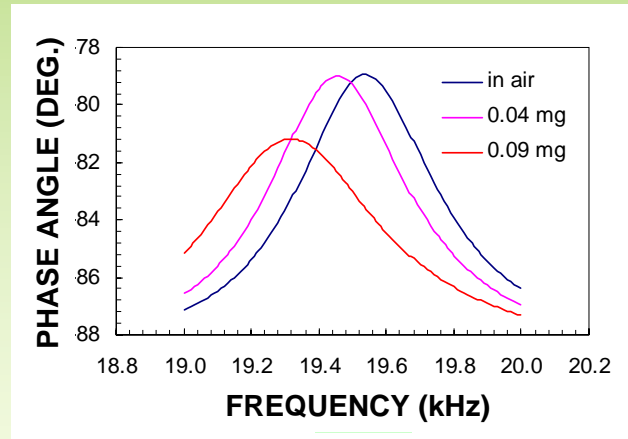
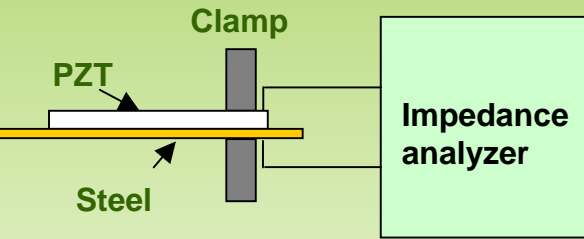
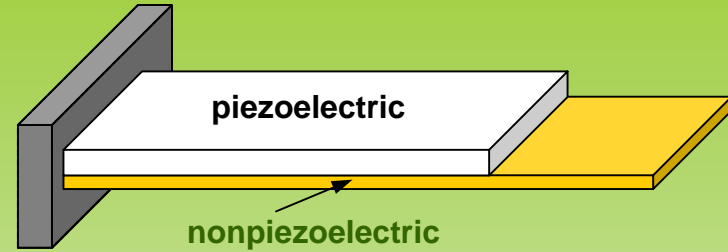
**Need:** a sensor that is

*direct, sensitive, rapid, real-time, and in-situ*

# Dynamic Applications: Biosensors

## DESIGN

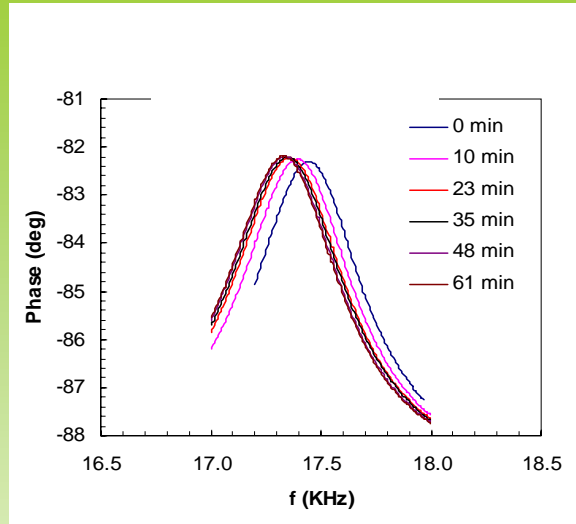
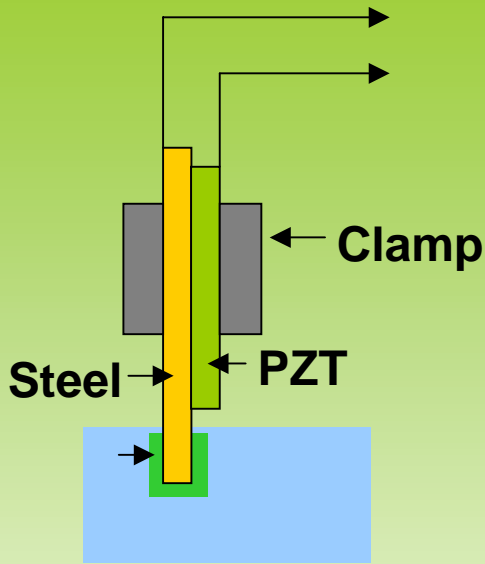
Detection: **DETECT** MASS CHANGE  
BY RESONANCE FREQUENCY SHIFT  
Measurement: ELECTRICAL



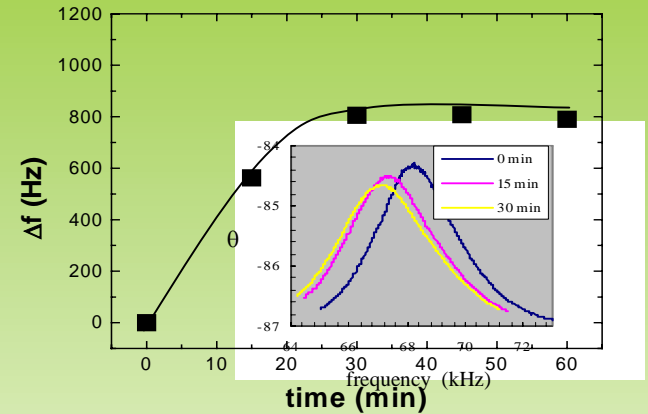
## Advantages:

- ❑ Label free
- ❑ Portable
- ❑ Rapid real-time detection
- ❑ Airborne and in-liquid

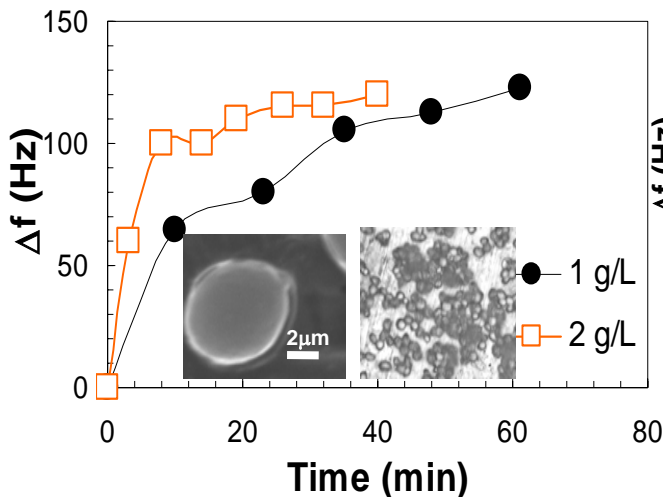
# Piezoelectric Microcantilever Biosensing



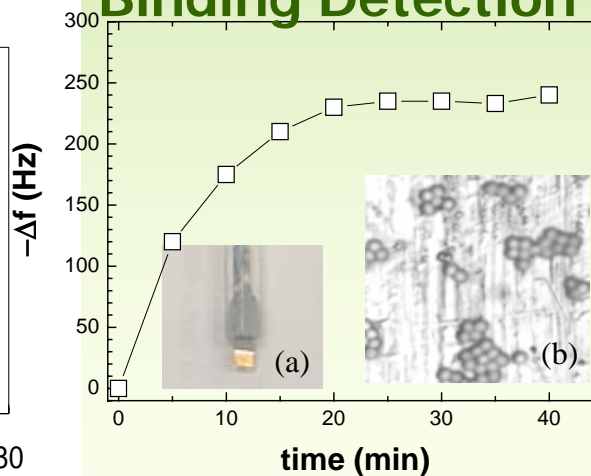
## Protein Detection: Avidin



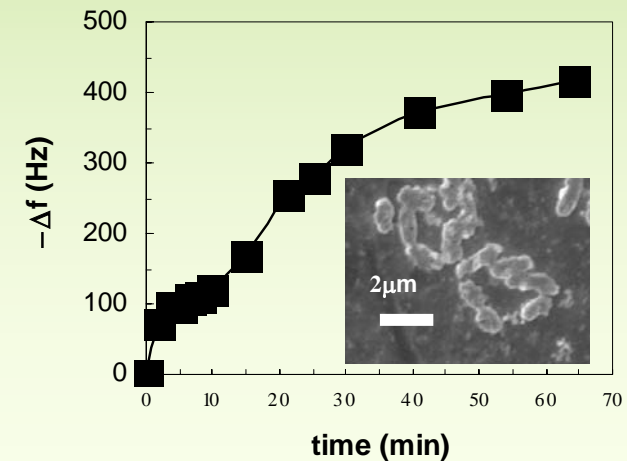
## Yeast Quantification



## Biotin-Avidin Binding Detection

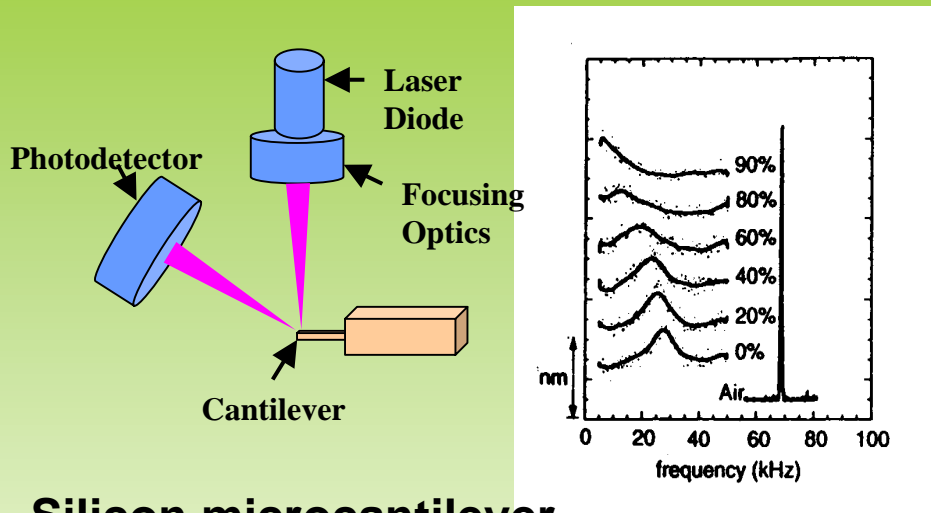


## E coli Detection



# Comparisons

## Silicon microcantilever



## Silicon microcantilever

### Disadvantages:

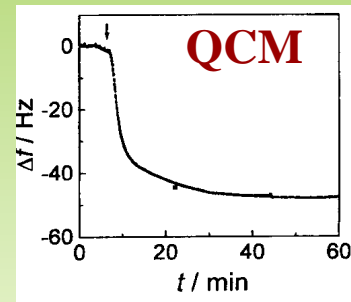
- ❑ need a driver for actuation
- ❑ Need optical system for detection
- ❑ Q factor  $\approx 1$  in water,
- ❑ in-water detection difficult
- ❑ Difficult for multiplexing

T. Thundat, E. A. Wachter, S. L. Sharp, and R. J. Warmack, *Appl. Phys. Lett.*, **66**, 1695 (1995).

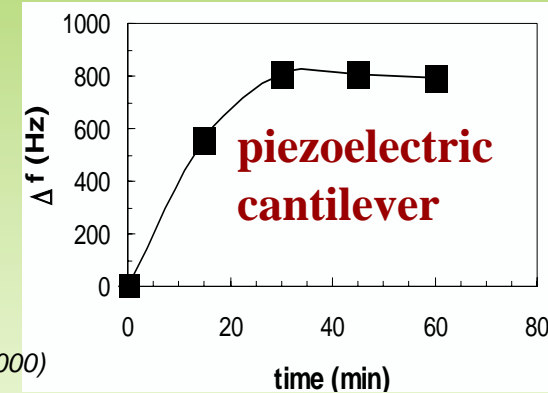
B. Ilic, D. Czaplewski, H. G. Craighead, P. Neuzil, C. Campagnolo, and C. Batt, *Appl. Phys. Lett.*, **77**, 450 (2000).

## Quartz Crystal Microbalance (QCM)

### Avidin binding on 3-mercaptopropionic acid



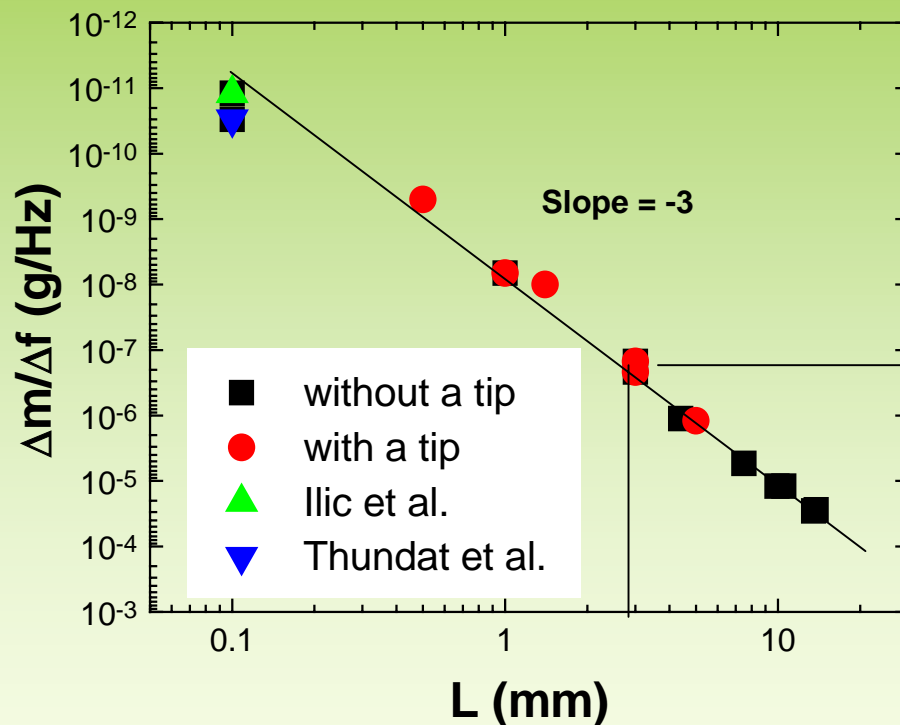
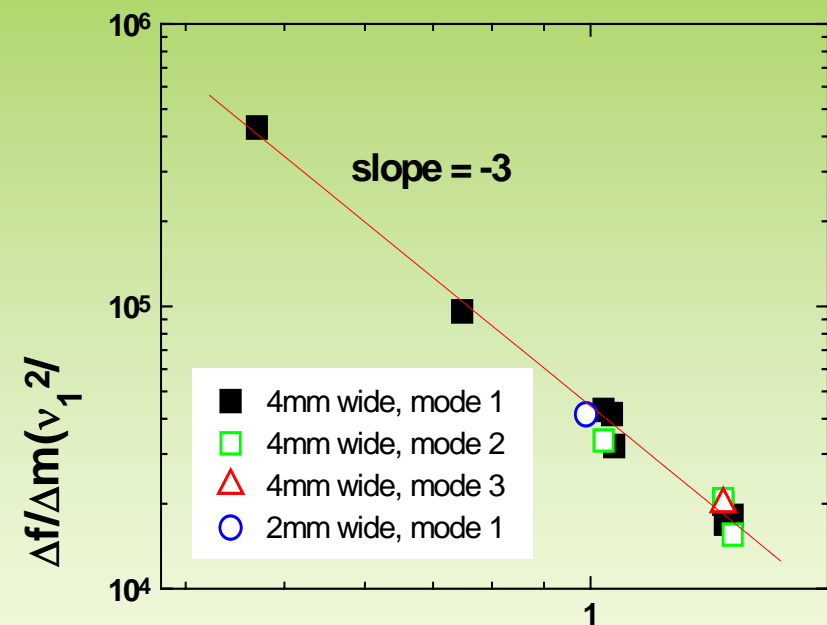
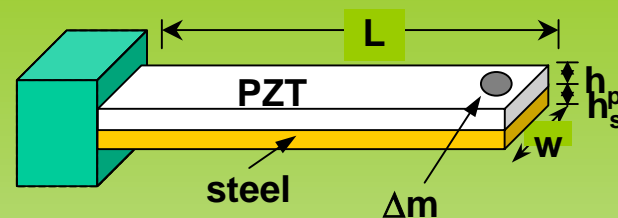
*Biophysical Journal*, **78**, 487 (2000)



- ❑ Cantilever of 0.7 mm long PZT with a 2.5 mm tip is already **16 times** more sensitive in  $\Delta f$  than the most common 5MHz QCM
- ❑ Also difficult for multiplexing

# Mass Detection sensitivity

$$\frac{\Delta f}{\Delta m} \cong \frac{1}{2} \frac{f}{M} = \frac{v^2}{4\pi} \frac{1}{L^3 w} \left( \frac{1}{0.236 \sqrt{12} \tilde{\rho}} \sqrt{\frac{\tilde{E}}{\tilde{\rho}}} \right)$$



**At 100 $\mu$ m length,  $\Delta m/\Delta f$  will approach  $10^{-16}$  g/Hz !**

# Objectives

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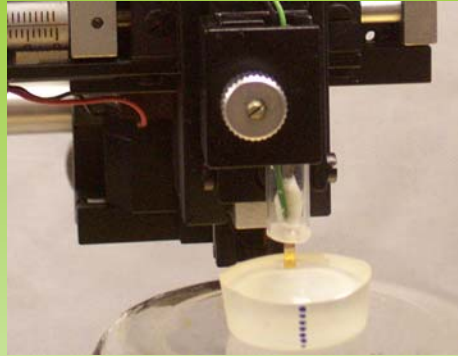
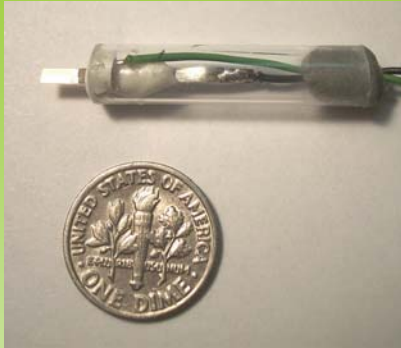
To develop high piezoelectric microcantilever sensors for real-time, in situ ultrasensitive pathogen pathogen detection in water

## Tasks:

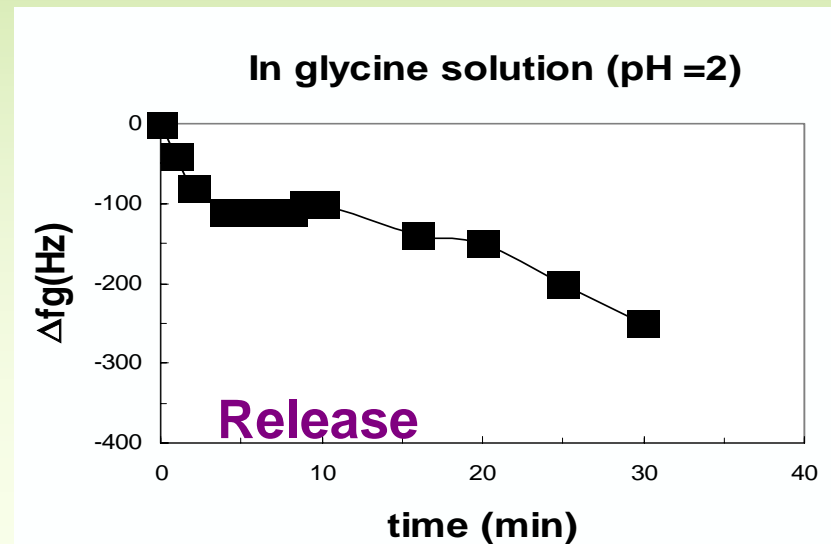
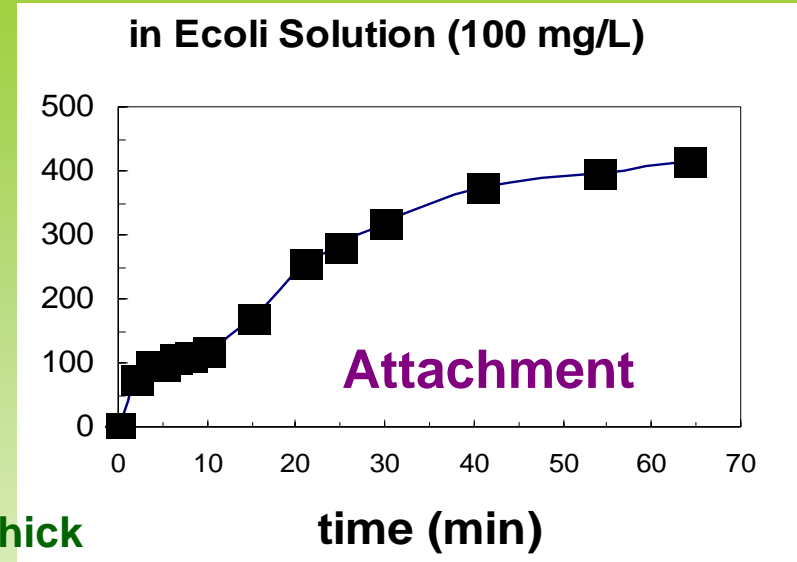
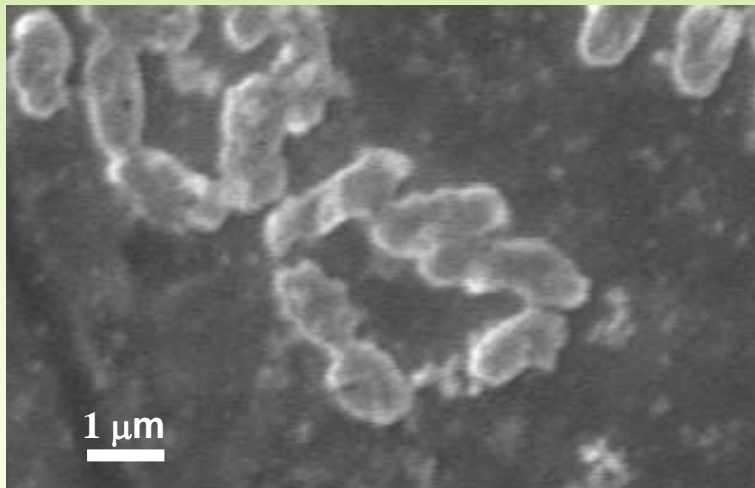
- To develop highly piezoelectric lead magnesium niobate-lead titanate (PMN-PT) films for microcantilever miniaturization
- To fabricate and characterize highly PMN-PT microcantilever sensors
- To use sub-millimeter PZT cantilevers for immediate pathogen detection development



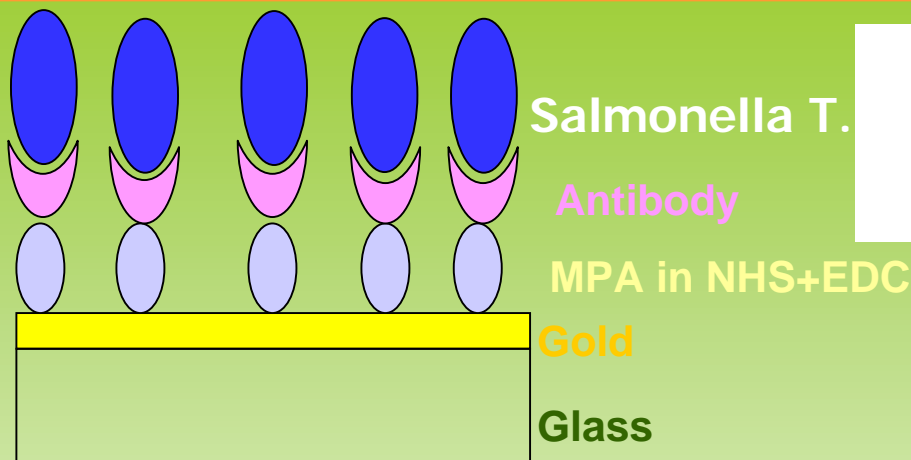
# Ecoli 057:H7 Detection and Release



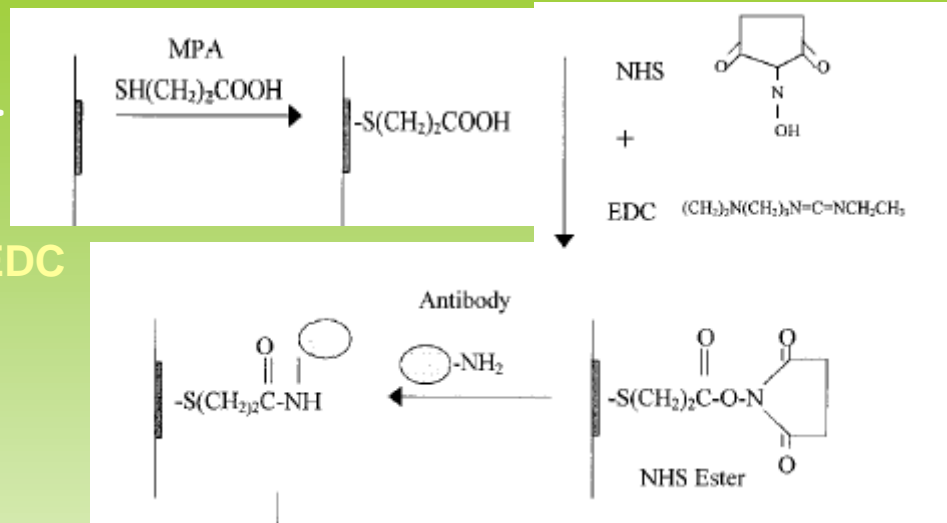
PZT: 0.6 mm long, 2 mm wide, 127  $\mu\text{m}$  thick  
 stainless steel tip: 4 mm long, 2 mm wide, 50  $\mu\text{m}$  thick



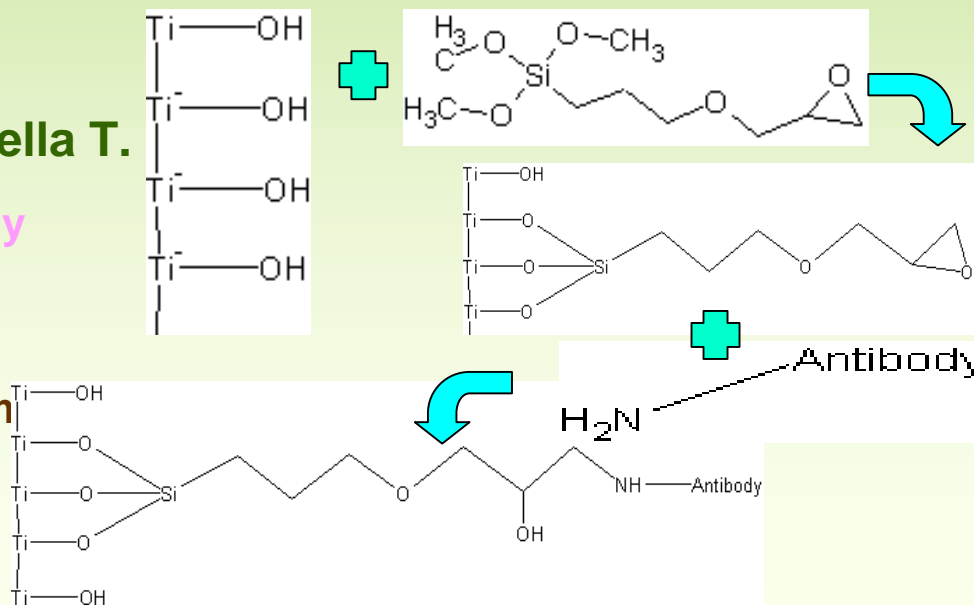
# Immobilization on glass, Ti, and Au



**PZT/Au-coated glass cantilever**

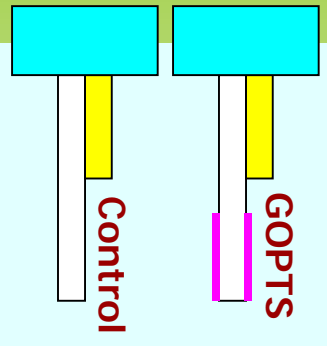


**PZT/Ti cantilever**

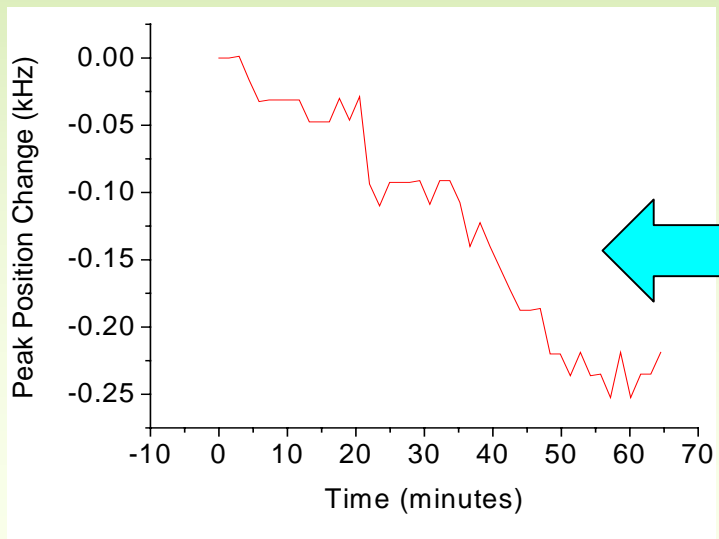
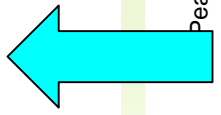
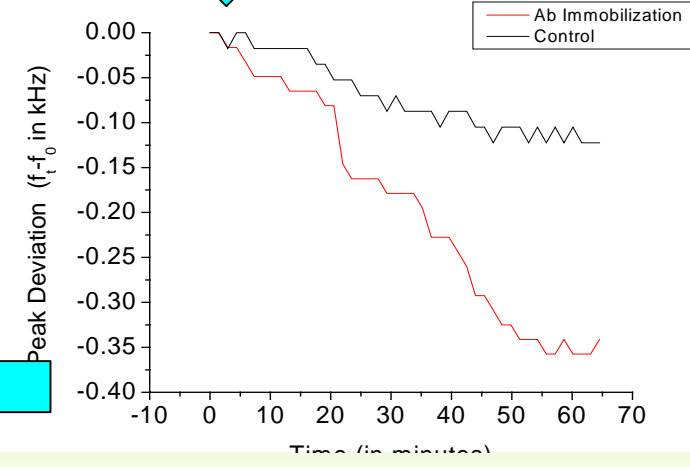
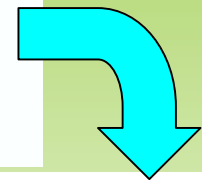
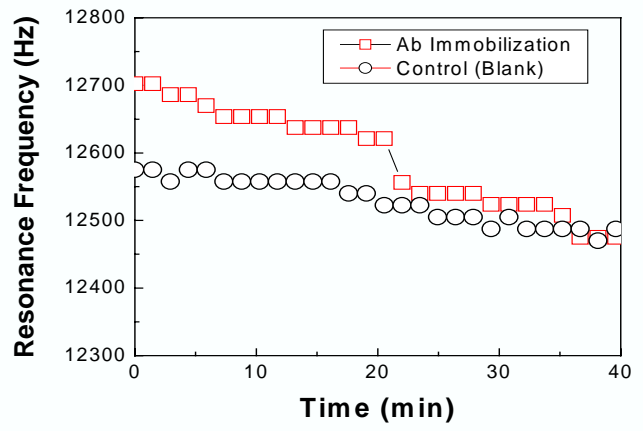


# Multiplexing

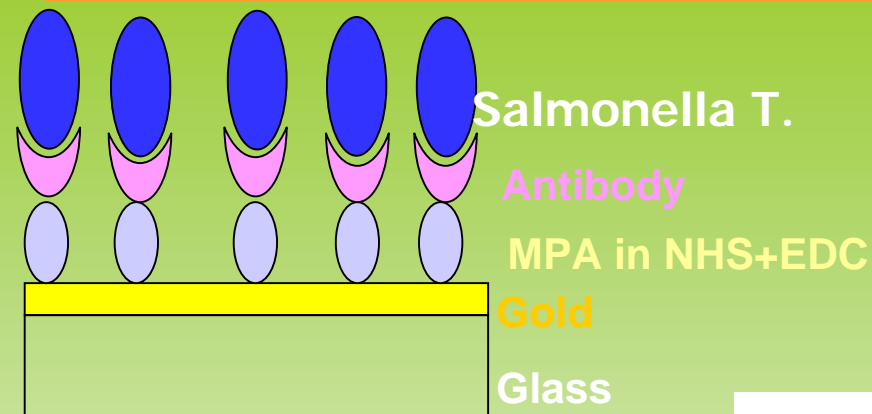
Detection with a Control



Antibody solution

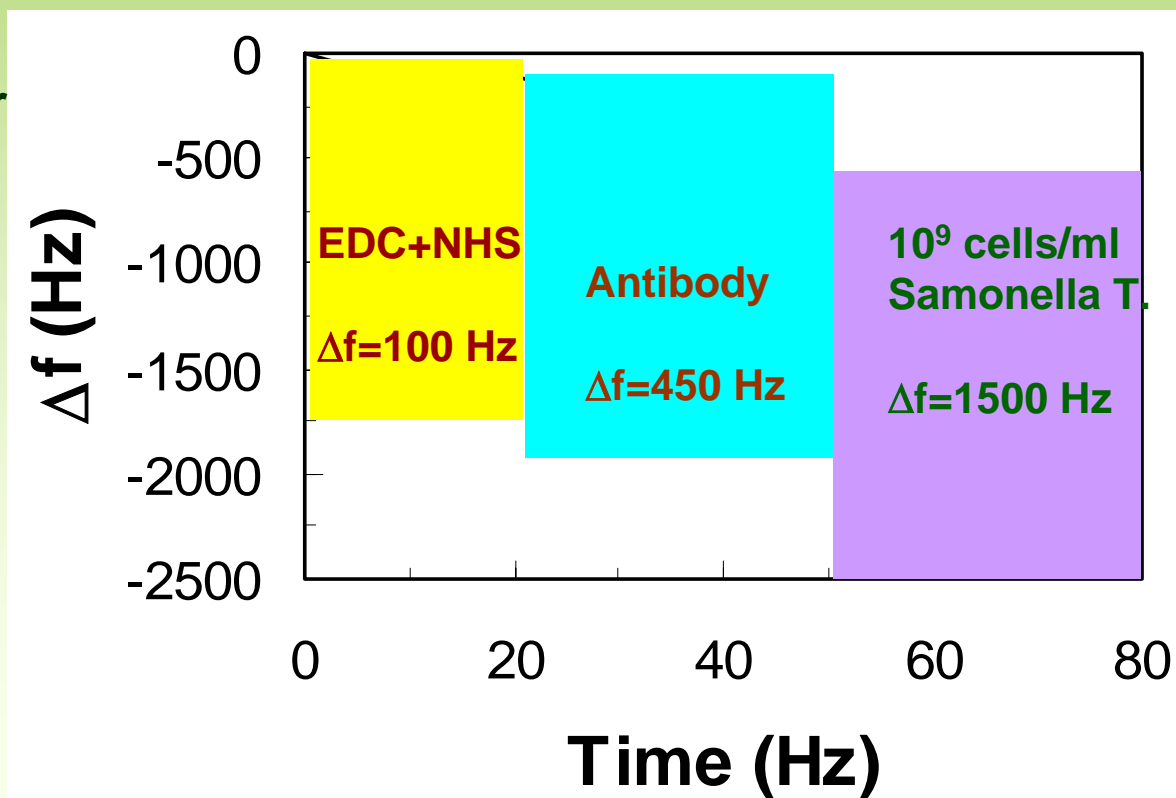
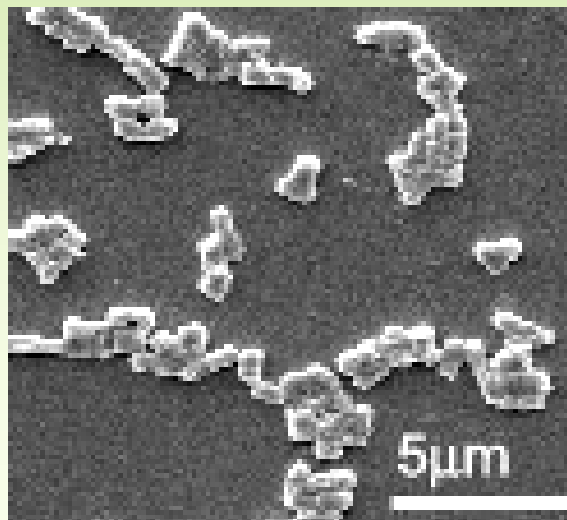


# Salmonella detection on gold coated PZT/glass cantilever

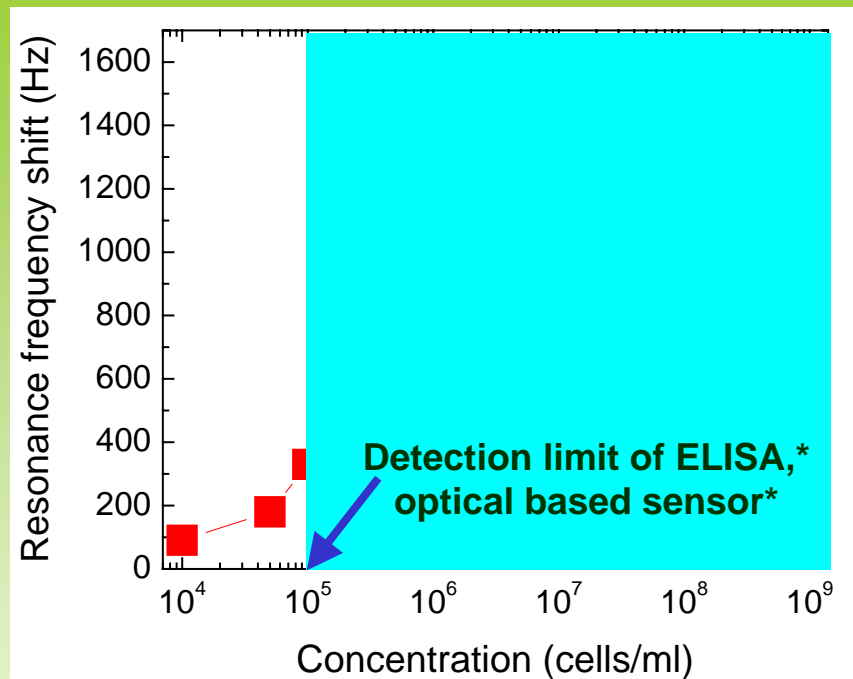
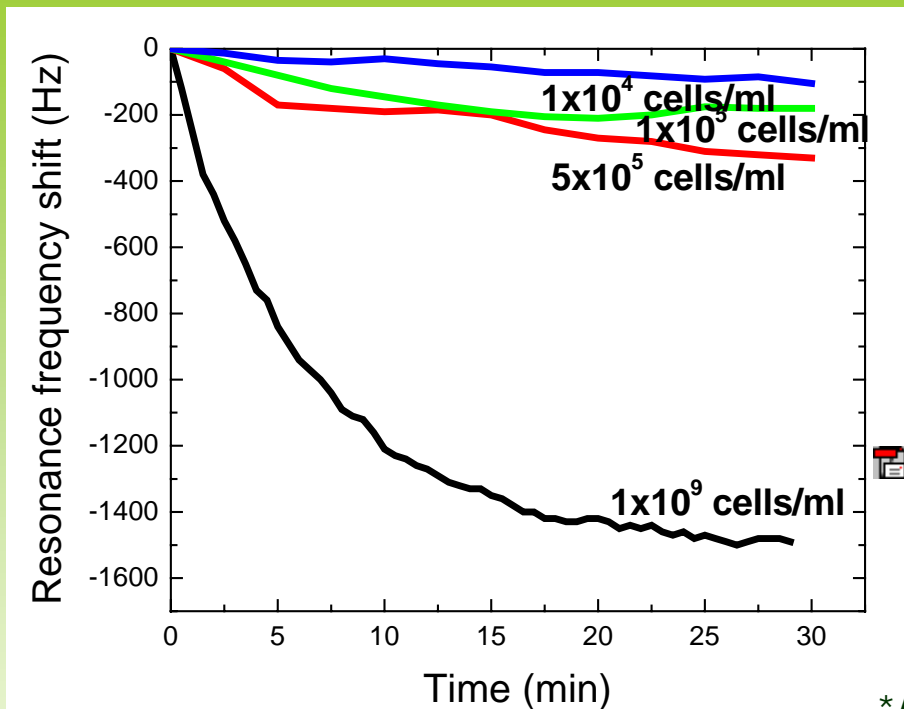


PZT/Au-coated glass cantilever

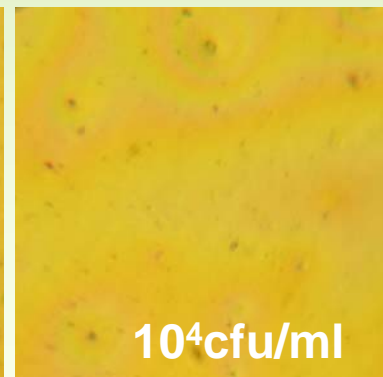
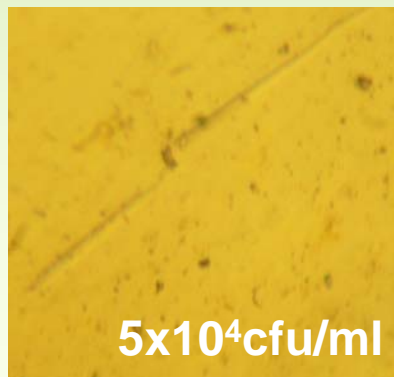
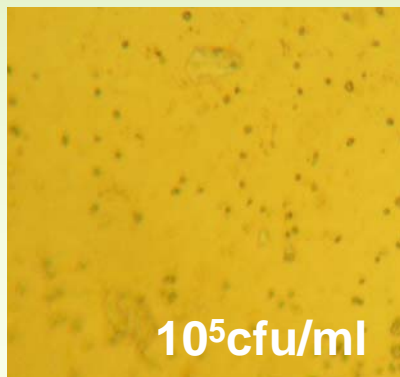
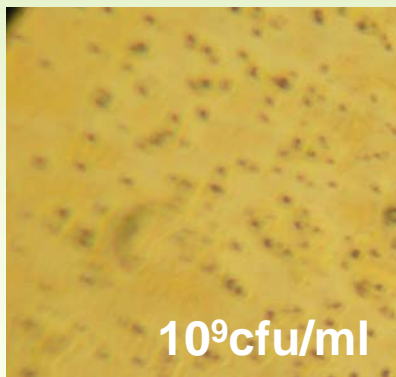
Salmonella T



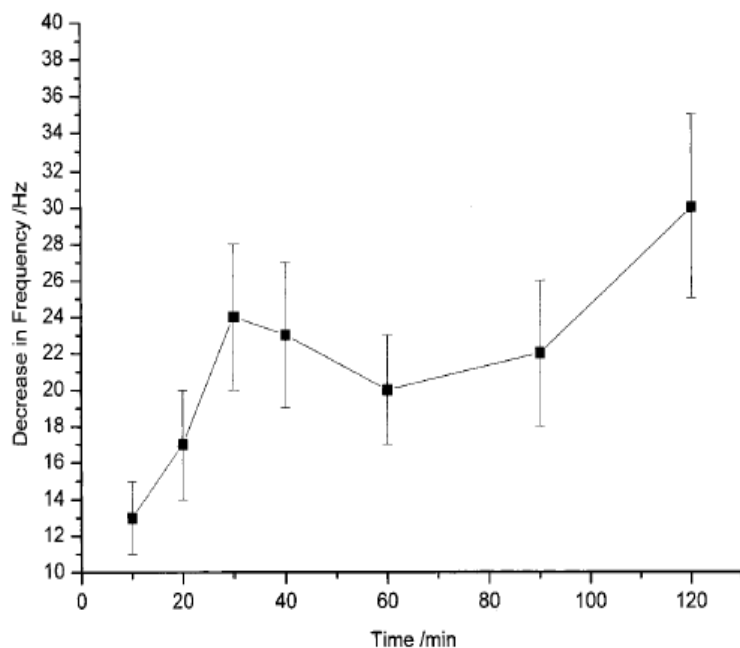
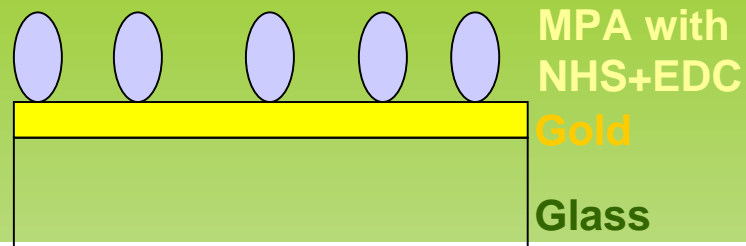
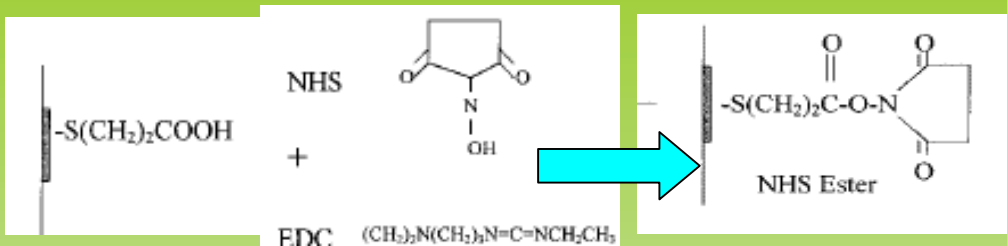
# Salmonella detection on gold coated PZT/glass cantilever



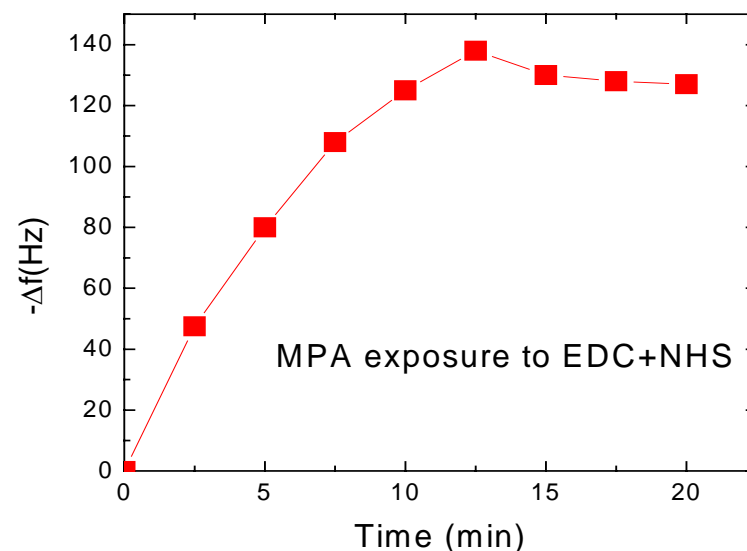
\*Anal. Chem. 71, 3846 (1999), Anal Chem. 75, 5293 (2003)



# MPA exposure to EDC + NHS

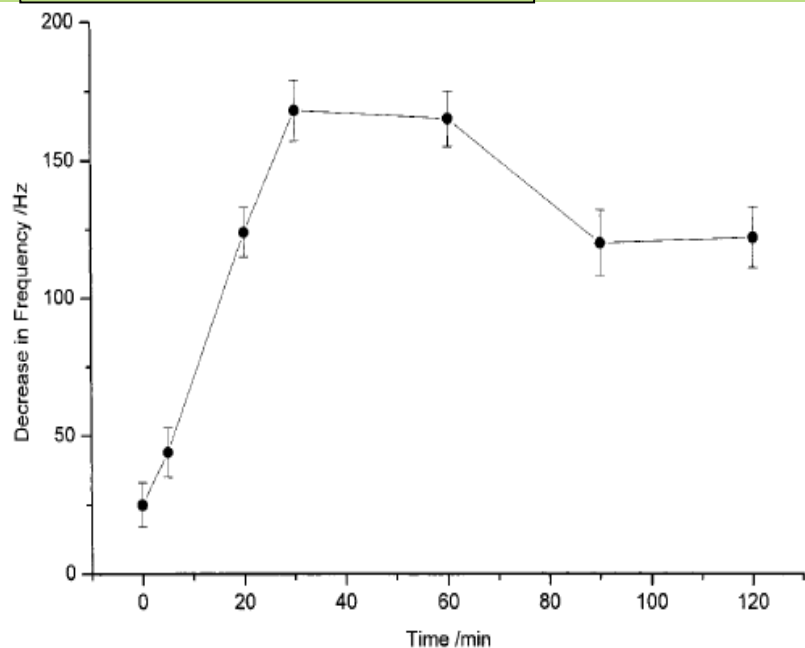
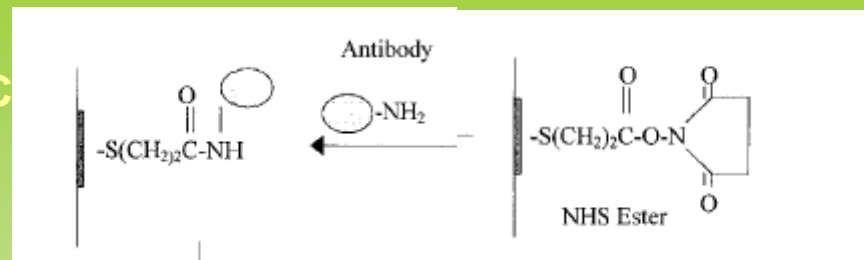
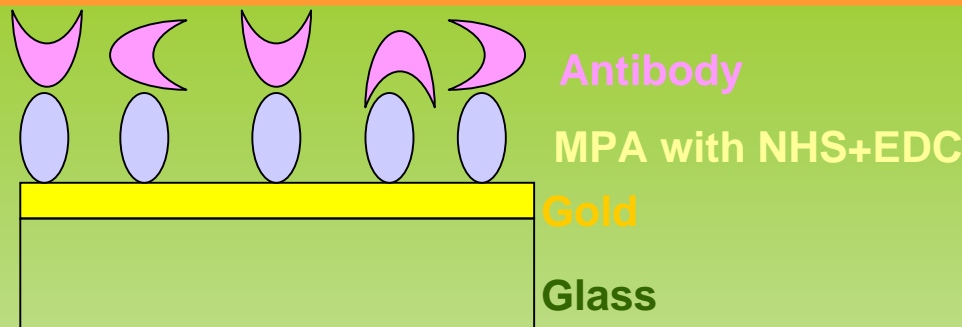


**10 MHz QCM\***  
 **$\Delta f = 30$  Hz**

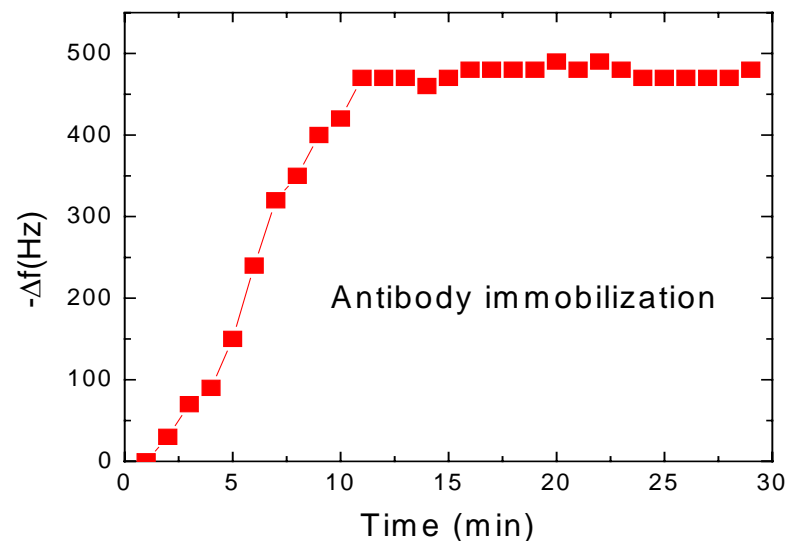


**PZT/glass cantilever**  
 **$\Delta f = 100$  Hz**

# Antibody immobilization detection



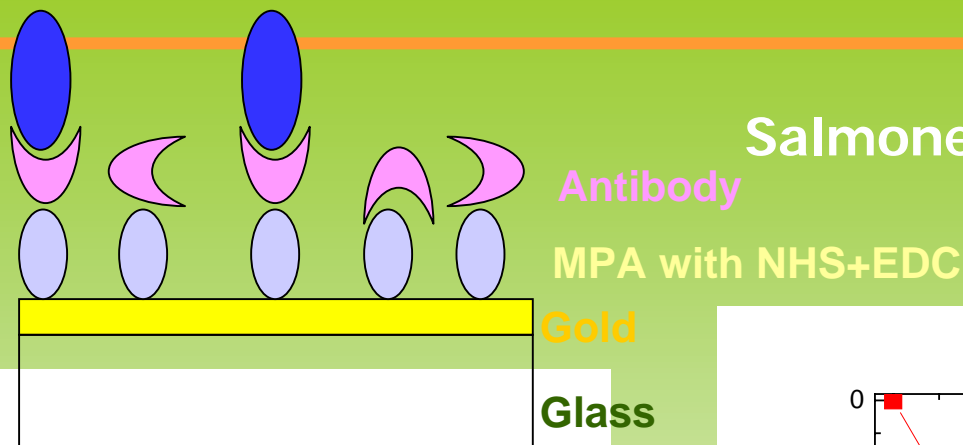
**10 MHz QCM\***  
 $\Delta f = 150 \text{ Hz}$



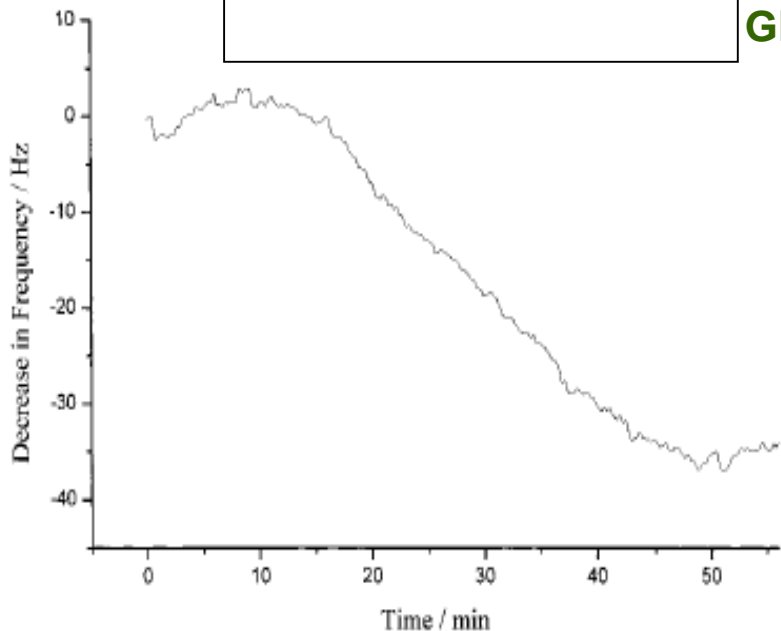
**PZT/glass cantilever**  
 $\Delta f = 450 \text{ Hz}$

\*Y. S. Fung\* and Y. Y. Wong, Anal. Chem. 2001, 73, 5302-5309

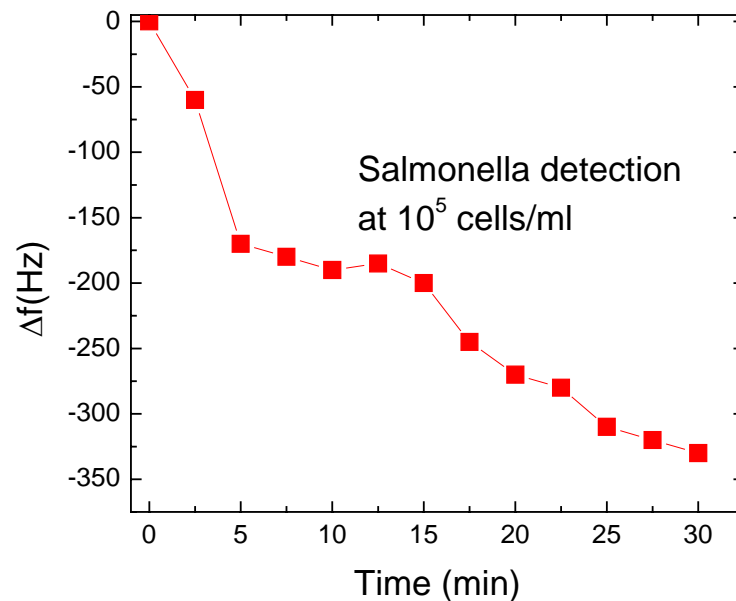
# Salmonella detection at $10^5$ cells/ml



Salmonella T. at  $10^5$  cfu/ml



**10 MHz QCM\***  
 **$\Delta f = 40$  Hz**



**PZT/glass cantilever**  
 **$\Delta f = 300$  Hz**

\*Y. S. Fung\* and Y. Y. Wong, Anal. Chem. **2001**, 73, 5302-5309

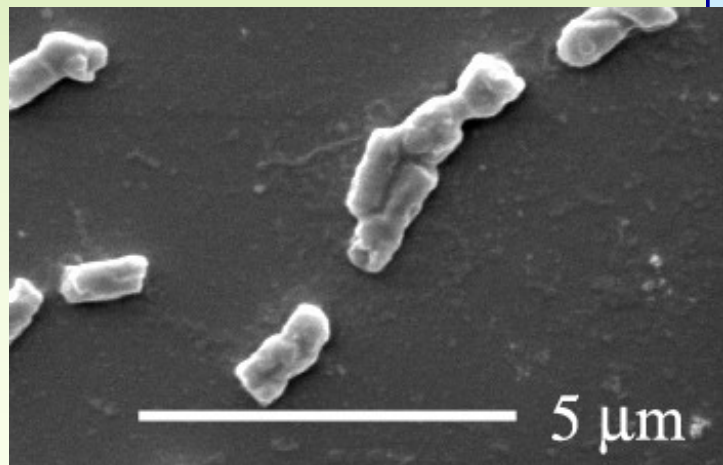
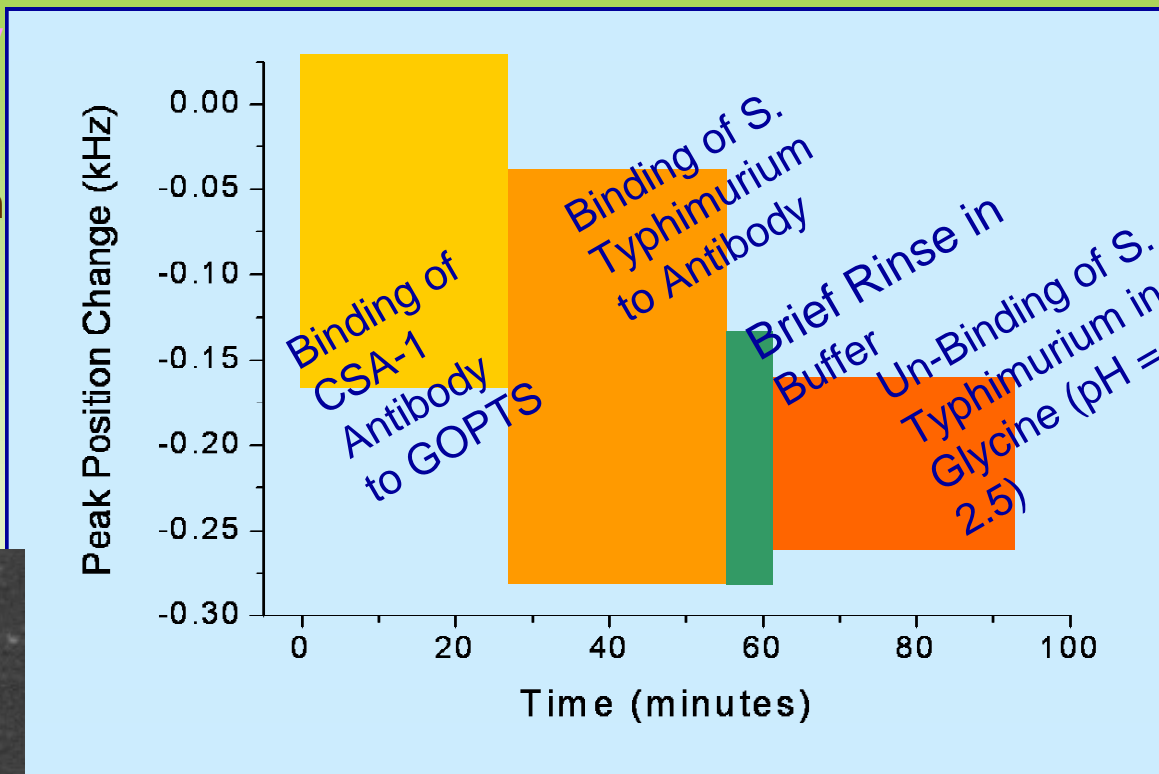
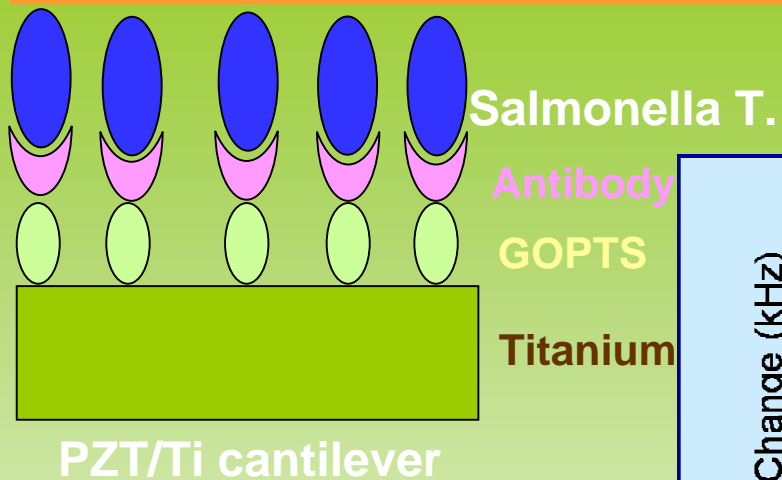


# Summary for 0.4 mm long PZT/glass cantilever

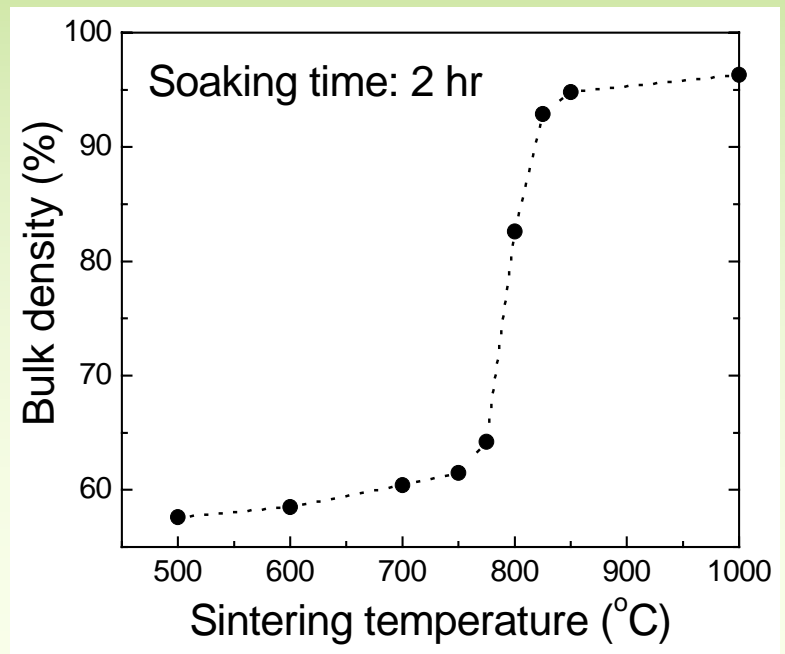
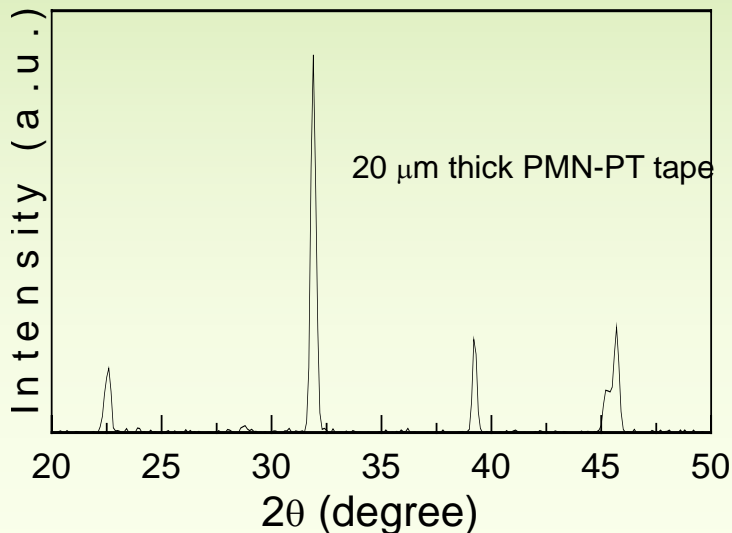
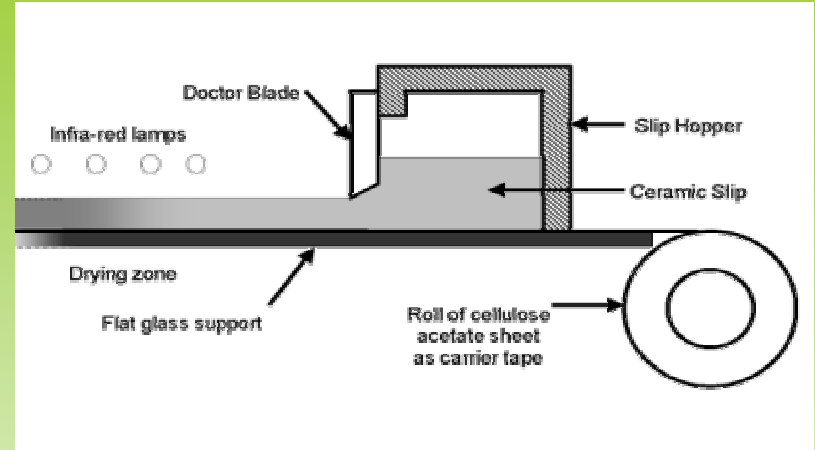
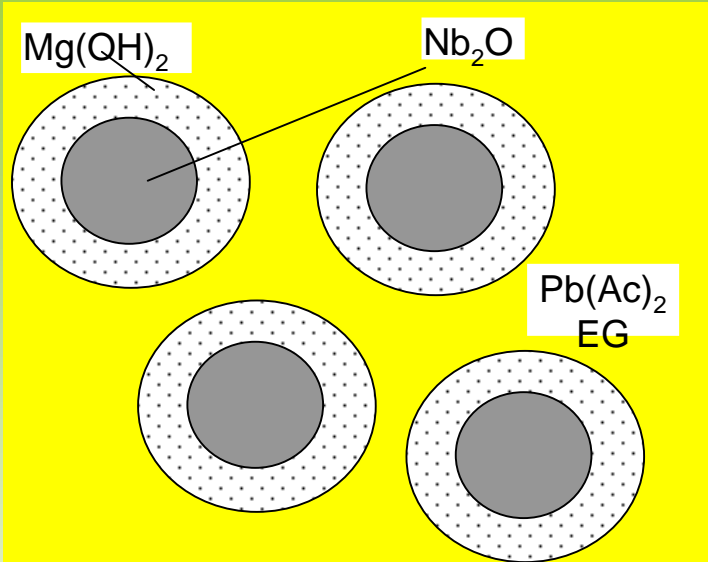
Detection sensitivity:  $3 \times 10^{-11}$  g/Hz

<b>C (cells/ml)</b>	<b><math>1 \times 10^9</math></b>	<b><math>1 \times 10^5</math></b>	<b><math>5 \times 10^4</math></b>	<b><math>1 \times 10^4</math></b>
<b><math>\Delta f</math></b>	<b>1400 Hz</b>	<b>300 Hz</b>	<b>180 Hz</b>	<b>90 Hz</b>
<b><math>\Delta m</math></b>	<b>42 ng</b>	<b>10 ng</b>	<b>6 ng</b>	<b>3 ng</b>
<b>Cells needed</b>	<b><math>3 \times 10^5</math></b>	<b>4000</b>	<b>3000</b>	<b>1500</b>
<b>Vol. needed</b>	<b>4 <math>\mu</math>l</b>	<b>50 <math>\mu</math>l</b>	<b>70 <math>\mu</math>l</b>	<b>170 <math>\mu</math>l</b>
<b>Diffusion type</b>	<b>Linear</b>	<b>3-d</b>	<b>3-d</b>	<b>3-d</b>

# 1.5 mm long PZT/Ti cantilever

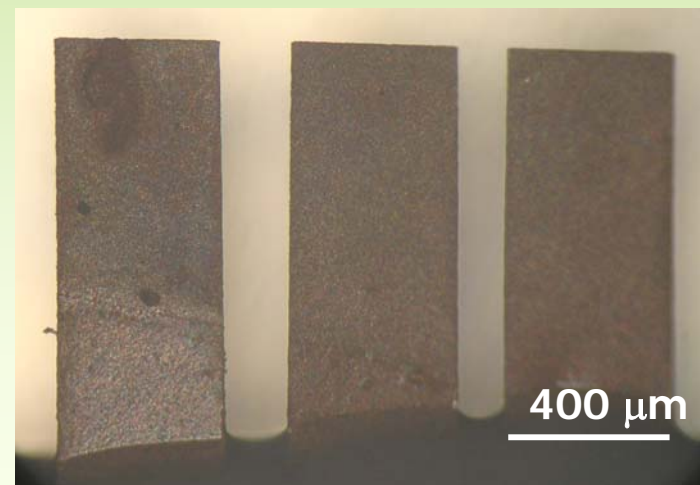
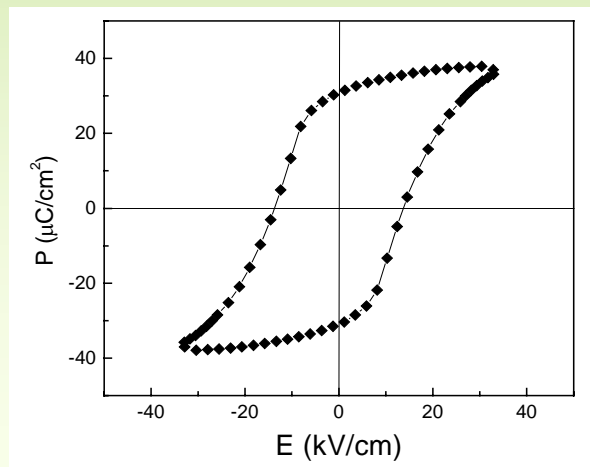
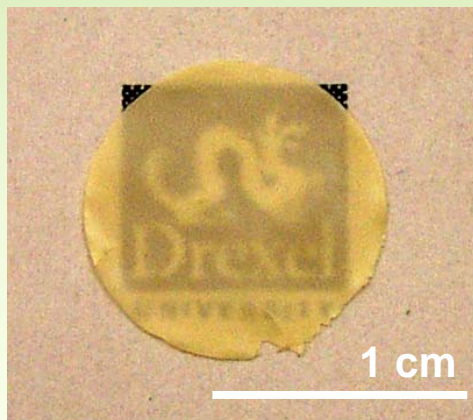
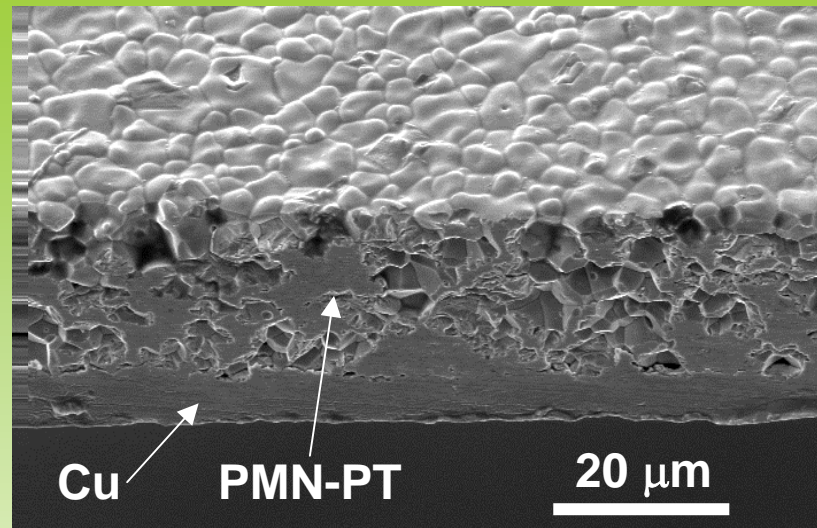
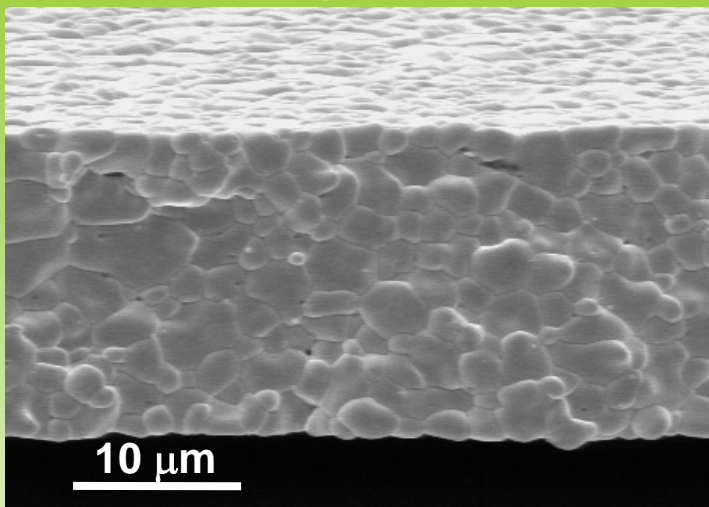


# Novel Coating Process for PMN-PT

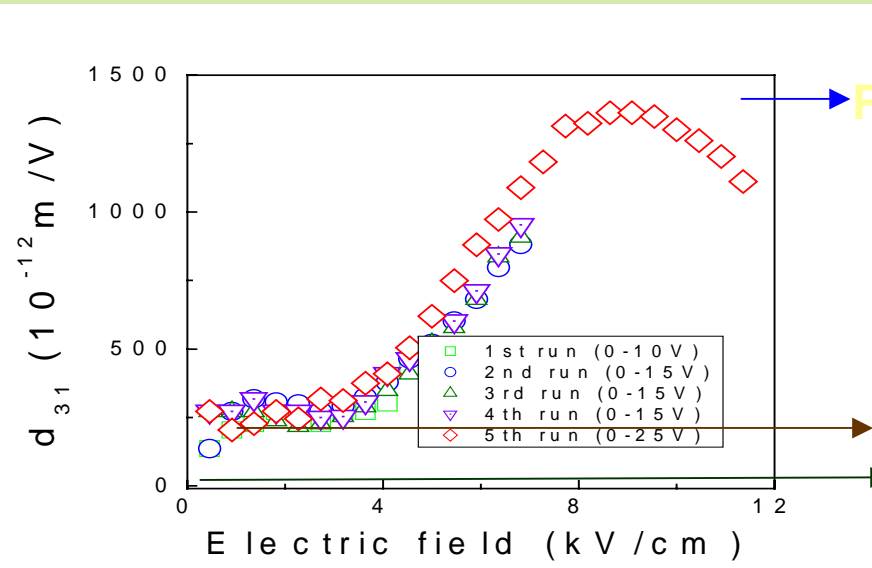
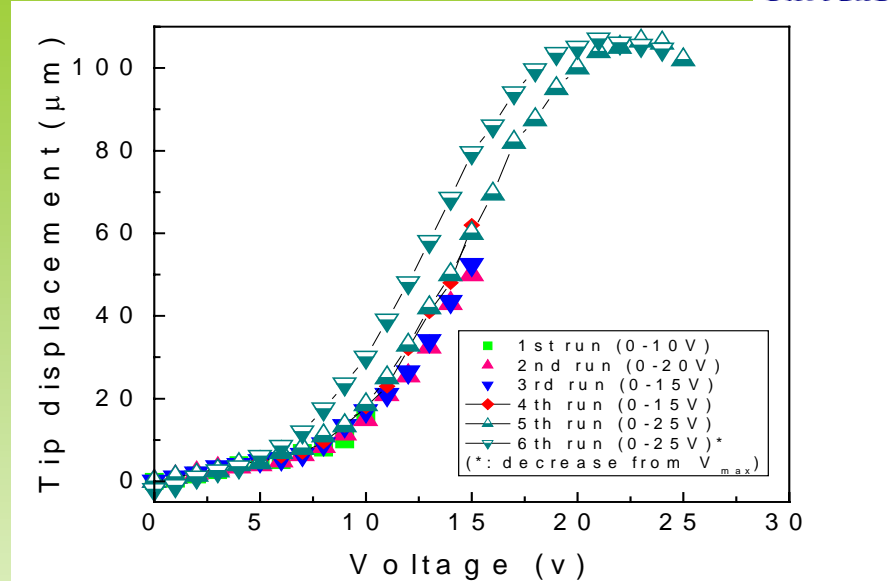
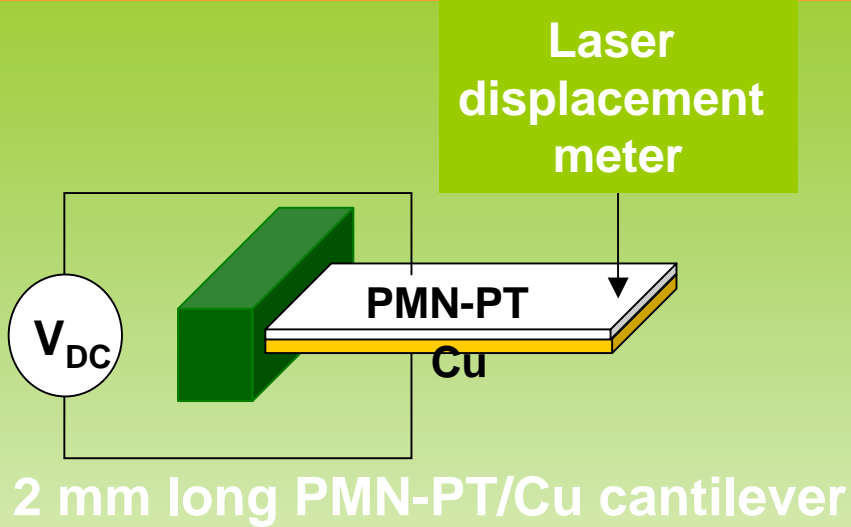


# PMN-PT/Cu Microcantilevers

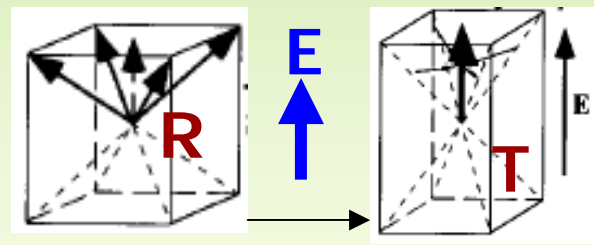
## Freestanding PMN-PT film



# PMN-PT/Cu Microcantilevers



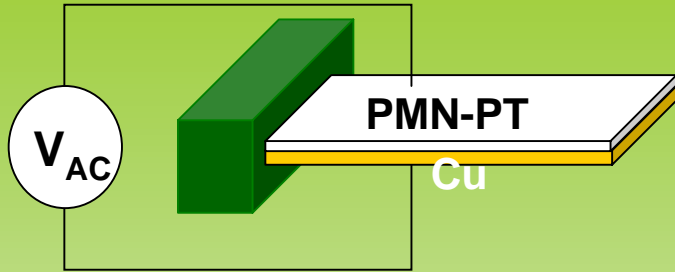
PMN-PT single crystal cut in (100)  $d_{31} \sim 1500$



Bulk PMN-PT  $d_{31} \sim 250$   
 ZnO<sub>2</sub>  $d_{31} < 10$

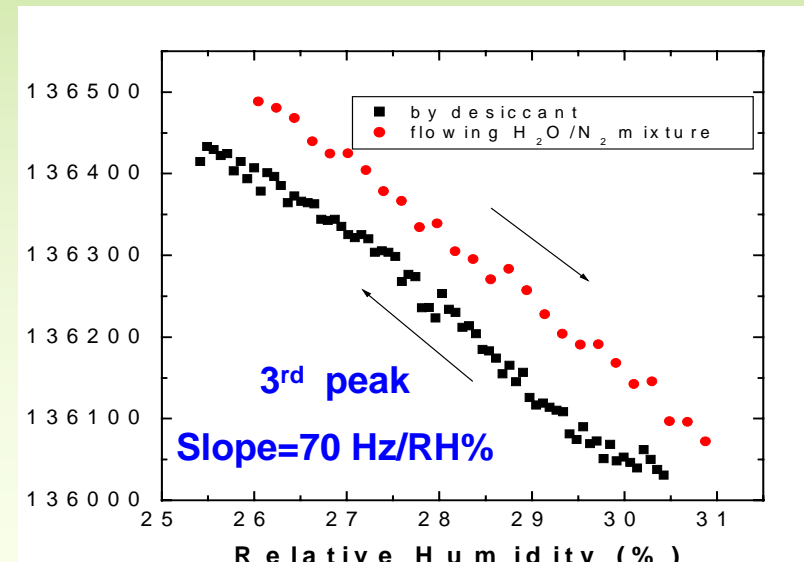
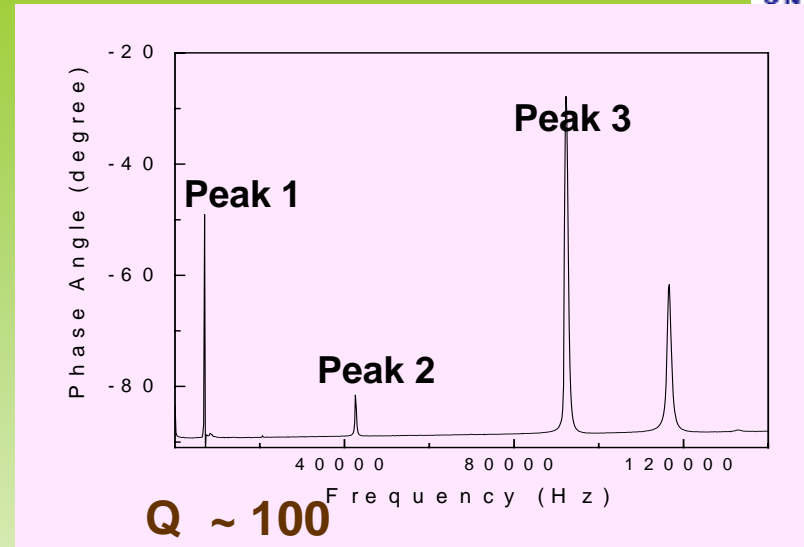
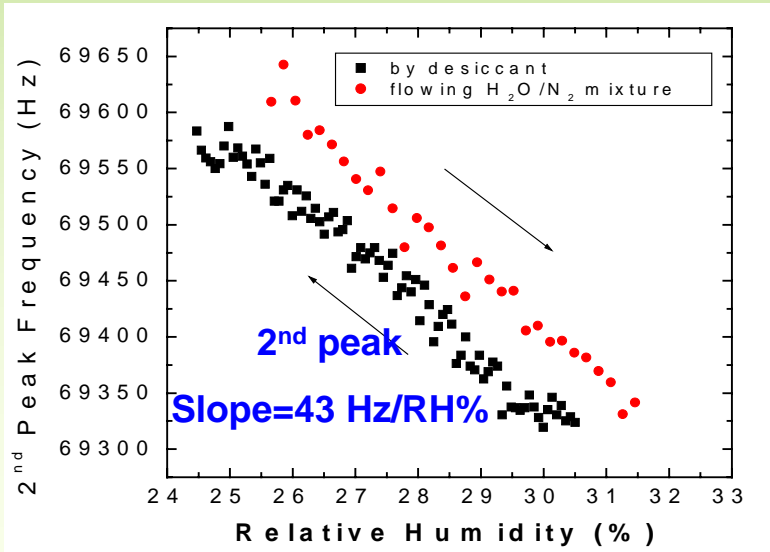
$$d_{31} = \frac{h}{3VL^2} \cdot \frac{E_1^2 t_1^4 + E_2^2 t_2^4 + 2E_1 t_1 E_2 t_2 (2t_1^2 + 2t_2^2 + 3t_1 t_2)}{E_1 E_2 t_1 (t_1 + t_2)}$$

# PMN-PT/Cu Microcantilevers



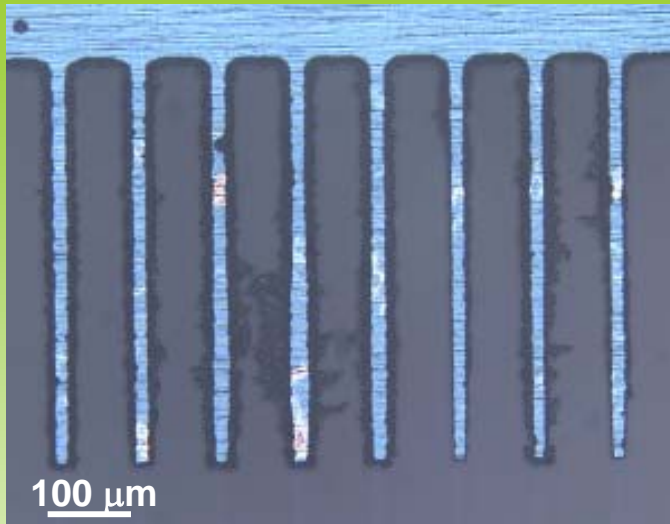
0.6 mm long PMN-PT/Cu cantilever  
PMN-PT 20  $\mu\text{m}$  thick, Cu 5  $\mu\text{m}$  thick

## Humidity Sensing

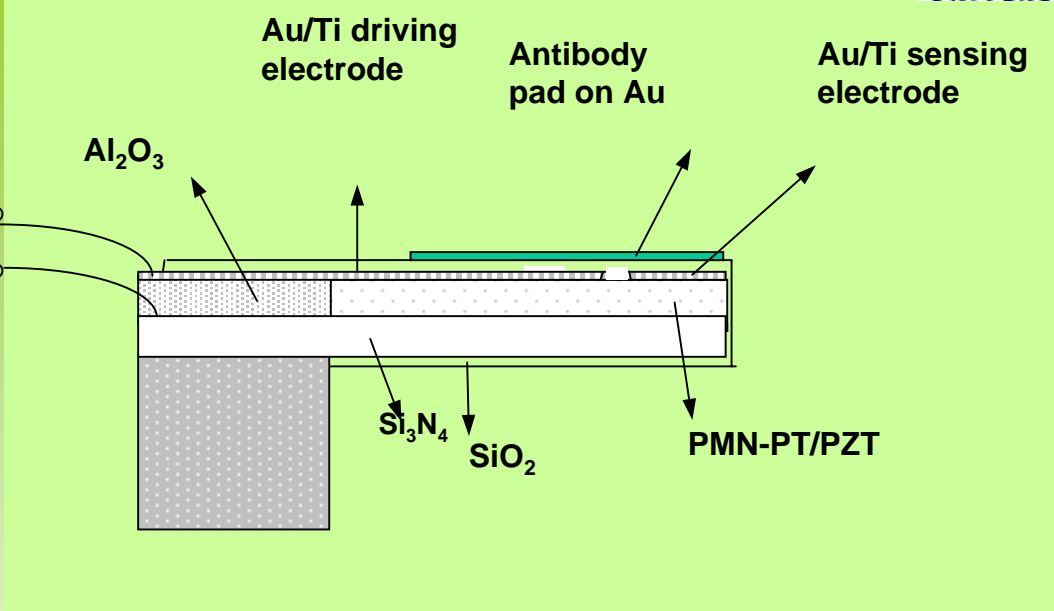


$$\Delta m / \Delta f \sim 10^{-12} \text{ g/Hz}$$

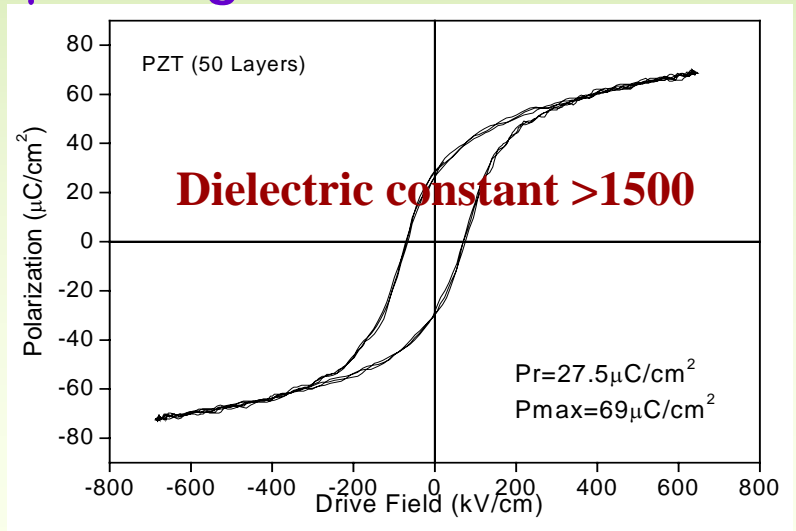
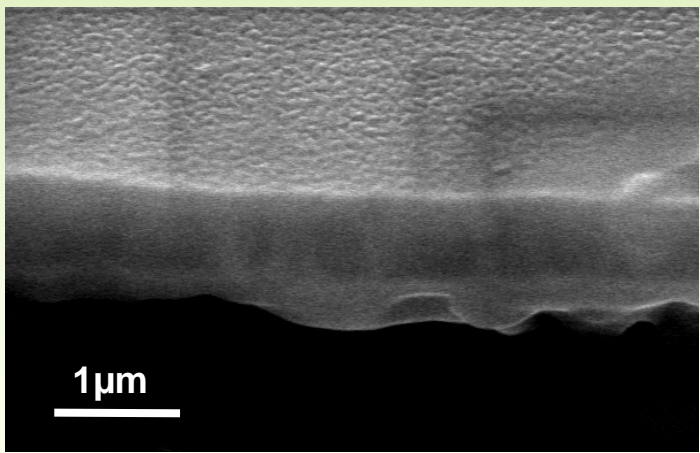
# Piezoelectric Microcantilever Fabrication



PZT/Mo microcantilevers



$\Delta m/\Delta f = 10^{-15} \text{ g/Hz}$  for 50  $\mu\text{m}$  long cantilever



# Conclusions



- ❑ High piezoelectric PMN-PT films have been successfully made with better than specially cut single-crystal piezoelectric properties
- ❑ Array PMN-PT/Cu microcantilevers 500  $\mu\text{m}$  long were made showing better than  $10^{-12}$  g/Hz
- ❑ we demonstrated in-situ real time detection of Salmonella in **standing water** with PZT/glass and PZT/Ti cantilevers with various immobilization schemes
- ❑ 0.4 mm long PZT/glass cantilevers of  $3 \times 10^{-11}$  g/Hz sensitivity **3-4** times more sensitive than 10MHz QCM in protein/molecular detection, **6-7** times more sensitive than 10MHz QCM's planar detection in cell detection
- ❑ The better than  **$10^4$  cfu/ml** detection sensitivity comparable to a 10 MHz QCM, better than commercial ELISA, and tapered optical fiber sensors
- ❑ Further sensitivity improvement includes using PMN-PT/Cu microcantilevers, using a flow cells



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