

US EPA ARCHIVE DOCUMENT

**ANAEROBIC BIODEGRADATION RATES
OF ORGANIC CHEMICALS
IN GROUNDWATER:
A SUMMARY OF
FIELD AND LABORATORY STUDIES**

Work Assignment Manager
and Technical Direction:

Dr. Zubair A. Saleem
U.S. Environmental Protection Agency
Office of Solid Waste
Washington, DC 20460

Prepared by:

HydroGeoLogic, Inc.
1155 Herndon Parkway, Suite 900
Herndon, VA 20170
Under Contract No. 68-W7-0035

U.S. Environmental Protection Agency
Office of Solid Waste
Washington, DC 20460

June 1999

TABLE OF CONTENTS

	Page
ACKNOWLEDGMENTS	iii
1.0 INTRODUCTION	1
2.0 REVIEW OF BIODEGRADATION STUDIES	2
3.0 REFERENCES	8
APPENDIX A CRITERIA FOR EVALUATING BIODEGRADATION RATE (LABORATORY STUDIES)	A-1
APPENDIX B CRITERIA FOR EVALUATING BIODEGRADATION RATE (FIELD STUDIES)	B-1
APPENDIX C CRITERIA FOR EVALUATING BIODEGRADATION RATE STUDIES (Syracuse Research Center)	C-1
APPENDIX D LABORATORY STUDIES FOR ALL COMPOUNDS	D-1
APPENDIX E FIELD STUDIES FOR ALL COMPOUNDS	E-1
APPENDIX F LABORATORY AND FIELD STUDIES FOR ALL COMPOUNDS	F-1
APPENDIX G REFERENCES FOR ALL COMPOUNDS	G-1

LIST OF FIGURES

	Page
Figure 2.1 Temperature of Groundwater in the United States at Depths of 10-20 meters (Collins, 1925)	4

LIST OF TABLES

	Page
Table 2.1 Summary of Rate Constants	5

ACKNOWLEDGMENTS

A number of individuals have been involved with the project. Dr. Zubair A. Saleem of the U.S. EPA, Office of Solid Waste, Washington, D.C., provided overall technical direction and review throughout this work. This report was prepared primarily by Mr. Jaideep Gadgil, with assistance from Mr. Sean Stanford all of HydroGeoLogic, Inc. Dr. Edward Bouwer of the Johns Hopkins University provided valuable technical consultation on the microbiological aspects throughout this effort. Useful inputs provided by many other EPA's scientists and microbiologists are greatly appreciated. Dr. John Wilson of the U.S. EPA, NRMRL, Ada, OK, provided the continued leadership on microbiological issues based on his extensive field as well as laboratory experiences. Dr. John Rogers of the U.S. EPA, NHEERL, Gulf Breeze, FL, helped with the conceptualization of microbial processes and mechanisms represented in the protocol, played a key role in the laboratory protocol developed earlier and published in the Federal Register and provided continuity with the previous efforts. Drs. Parmelli Pritchard, formerly with U.S. EPA, and Bob Boethling, OPPTS, U.S. EPA, Dr. Jack Jones of the U.S. EPA, Ecosystems Support Division, Athens, GA, was helpful throughout this review project. The Chemical Manufacturers Association, along with other industrial groups: American Petroleum Institute; American Forest and Paper Association; Edison Electric Institute; National Council of the Paper Industry for Air and Stream Improvement; coordinated and provided initial draft criteria and developed the data through the Syracuse Research Institute.

DISCLAIMER

The work presented in this document has been funded by the United States Environmental Protection Agency. Mention of trade names or commercial products does not constitute endorsement or recommendation for use by the Agency.

1.0 INTRODUCTION

Solid and hazardous wastes may pose a considerable threat to the health of humans and the environment. Under the Resource Conservation and Recovery Act (RCRA) of 1976 the U.S. EPA is appointed the task of managing these risks through the development and implementation of regulations. To this end, the EPA develops and uses fate and transport mathematical/computer models to aid in the assessment of risk from waste management practices. The input parameters of these models may be used to describe, in mathematical terms, the physical and chemical properties of actual landfill sites. These models may be used to develop a probability distribution of concentrations by performing a myriad of simulations, each time selecting sites parameters from a nation-wide database. This technique is known as Monte Carlo analysis and has proven a useful tool in risk analysis.

The EPA's Composite Model for Leachate Migration with Transformation Products (EPACMTP: U.S. EPA, 1996) is the subsurface fate and transport model which EPA's Office of Solid Waste implements on a nationwide basis using Monte Carlo analysis techniques. The implementation procedure requires nationally representative distributions of first order biodegradation rates.

EPA uses anaerobic biodegradation rates in the subsurface fate and transport model. The Agency considers that although anaerobic biodegradation is not the slowest activity, it is less likely that data collected under anaerobic conditions would lead to an overestimation of the degradation rate. In general, the concentrations of oxygen and nitrate are insignificant in ground waters that have been impacted with leachate from landfills. This condition results from prior microbial consumption of oxygen or nitrate that was supported by the electron acceptor demand of materials in the landfill leachate. Consequently, the rate of supply of oxygen and nitrate into impacted ground waters is slow. The rate of supply is limited by mass transfer processes from ground water that has not been impacted (and as a consequence has oxygen and nitrate available) into the plume impacted with leachate from a landfill. As a result, the only microbial processes that are generally available for biodegradation of hazardous organic compounds in ground water impacted with leachate from landfills are the processes of iron reduction, sulfate reduction, and methanogenesis (Krumholz et al, 1996). Therefore, EPA considers selected rate constants provided by laboratory and field studies that were conducted under the iron reducing, sulfate reducing, or methanogenic conditions to be the rate constants that are appropriate for use in EPACMTP. EPA does not consider rate constants provided by laboratory and field studies that were conducted under aerobic or nitrate reducing conditions to be appropriate for use in EPACMTP.

EPA developed a protocol to generate national distributions of anaerobic biodegradation rates for organic chemicals for use in the model (53 FR 22300, June 15, 1988). The protocol requires collecting samples from six sites: three sites in the northern half of the United States; and three sites located in the southern half of the country. Ideally, these six sites will represent the various pH and redox environments prevalent in the country. However, the protocol has not been implemented widely by the industry so far. Meanwhile, many laboratory and field studies on the anaerobic biodegradation of chemicals have become available since the protocol was developed more than ten years ago. Suggestions were made to critically evaluate these studies and incorporate the results in modeling. Therefore, EPA convened a workshop in Atlanta, Georgia, on anaerobic biodegradation of organic chemicals in 1997. Representatives from academia, industry and the EPA participated in discussions. Written comments from an environmental group were also considered at the workshop. Based on these discussions and other inputs from EPA scientists, the Agency developed criteria for the evaluation of the field as well as laboratory studies. The criteria are listed in Appendices A and B (laboratory and field studies, respectively).

At the workshop, industry representatives provided a report summarizing field and laboratory studies on anaerobic biodegradation of organic compounds. The report entitled "Anaerobic Biodegradation of Organic Chemicals in Groundwater: A Summary of Field and Laboratory Studies" was prepared for the American Petroleum Institute (API) and others by the Syracuse Research Center, 1997. All the research articles reviewed in the report were also submitted to EPA by the API. The criteria used by Syracuse Research Corporation are presented in Appendix C. Since the criteria used by Syracuse Research Center (Appendix C) to review research papers was less stringent than the criteria decided upon at the workshop, EPA conducted a second review of the API-submitted Syracuse Research Center Report. In addition, EPA conducted a literature review to collect additional studies on biodegradation of various organic chemicals (both laboratory as well as field). For purposes of this report, biodegradation is defined as "removal of a compound from ground water through biological activity". Only studies which were conducted with aquifer materials under anaerobic conditions were selected for review. In addition studies carried out on mixtures of compounds, and studies where the aquifer material was seeded with microorganisms from other sources were not included. The studies were then evaluated to see if they satisfied the proposed EPA criteria. Biodegradation rates from studies which met the proposed criteria were used to develop a distribution of first order rates to be used as potential input to the EPA's subsurface fate and transport model.

2.0 REVIEW OF BIODEGRADATION STUDIES

The studies submitted by Syracuse Research Center were divided into field and laboratory studies and the results from these studies are summarized separately. Results of a study were rejected if the study did not satisfy the criteria indicated as unacceptable in Appendix A or B. Appendices D, E, F, and G summarize the review of available biodegradation studies satisfying the proposed EPA criteria (Appendices A & B). The laboratory studies are summarized in Appendix D and the field studies are summarized in Appendix E. Appendix F has tables summarizing both field and laboratory studies for each compound. The references for each compound are listed in Appendix G. A summary of the distribution of rates for each compound is provided in Table 2.1.

Results of both field and laboratory studies are considered for the development of biodegradation rates for use in the model. Each category (field or laboratory) is further subdivided based on the temperature, pH and the redox regime. The biodegradation rate of a chemical depends, among other factors, on both the temperature and pH of the subsurface environment at the site. The subsurface reducing environment was assumed to be grouped into two broad categories: methanogenic; and sulfate reducing. Studies which identified iron reducing conditions were grouped under sulfate reducing. Studies which were purely denitrifying were not included as denitrification is believed to occur predominantly in the vadose zone (Krumholz et al, 1996). If a study met all criteria but was missing either temperature or pH or both, then pH and temperature were assigned to the study as follows:

- 1) pH: assume neutral range (6-8), for both laboratory and field; and
- 2) Temperature: assume 25 °C for laboratory, and for field studies refer to the nationwide distribution of temperature shown in Figure 2.1.

The pH regimes were grouped as: acidic (< 6), neutral (6-8), and alkaline (> 8). Two distinct temperature ranges were considered (# 15 °C and > 15 °C). Each table includes the lag time for degradation where reported and any special observations regarding the study. If a multiple redox regime was reported in the study (e.g., SO₄/CH₄/NO₃), the study was classified under the first relevant redox regime reported, in this case sulfate reducing. If no redox regime was reported the study was classified as sulfate reducing. If multiple studies were conducted at a site, a simple average was computed for the biodegradation rate and the average was used. The average was computed so that one site may not unduly bias the distribution of rate constants. The individual rates are also reported in parentheses following the average.

Information from the tables in Appendix F for each compound is used in the subsurface Monte-Carlo fate and transport modeling. In a Monte-Carlo realization, a site is selected along with the subsurface temperature and the pH. The reducing environment is randomly chosen, each environment having an equal likelihood of being selected. Then an anaerobic biodegradation rate is picked from the appropriate cell of the table corresponding to the selected reducing environment (the rates listed within a cell all have an equal likelihood of being chosen). The rate is then used for that site in the analysis. For the next Monte Carlo site realization, the model then selects a rate from the table based on the temperature, pH, and either methanogenic or sulfate reducing environment at random with replacement. The process is repeated for the total number of Monte Carlo realizations.

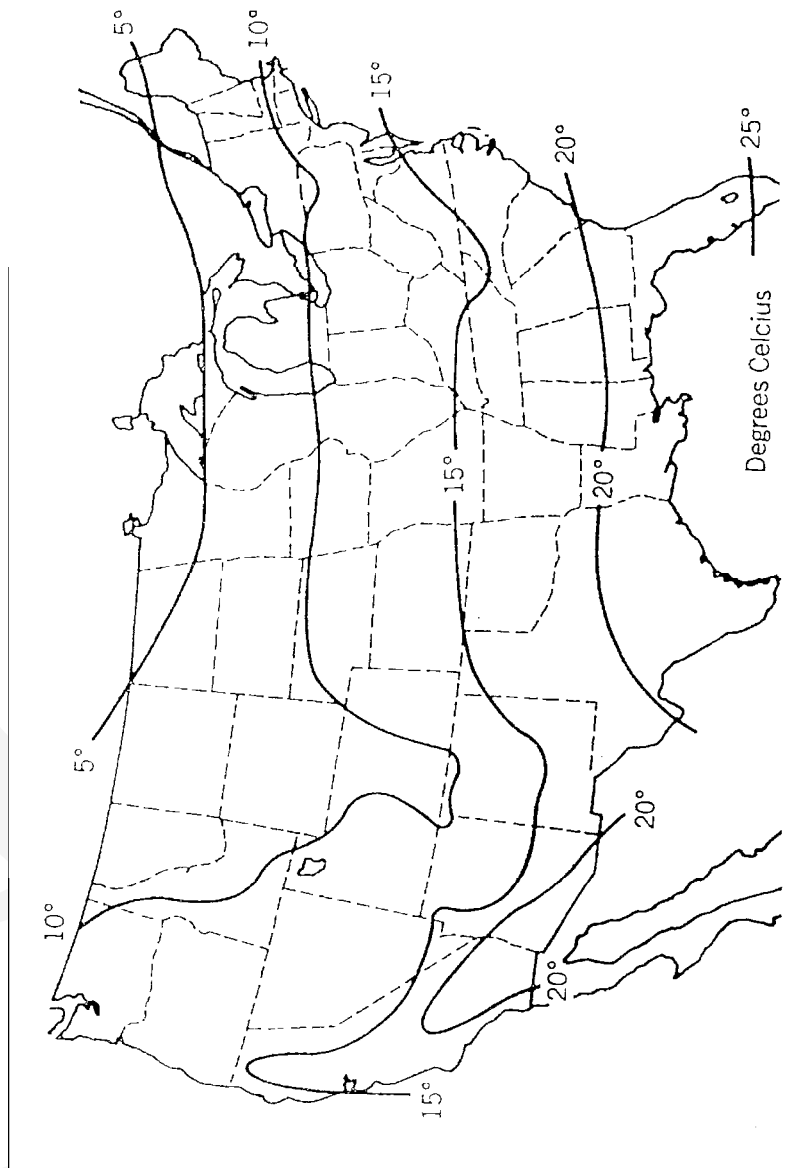


Figure 2.1 Temperature of Groundwater in the United States at Depths of 10-20 meters (Collins, 1925)

Table 2.1
Summary of Rate Constants

CAS Number / Chemical Name	Rate Constant (1/day)					Distribution Type	Molar Yield	Reaction Product
	Most Likely Estimate	Min	Max	Median	Standard Deviation			
000071-43-2 / Benzene	0	0	0.071	0	0.0152	--	--	Not Identified
000108-88-3 / Toluene	0.02	0	0.186	0.02	0.0372	--	--	Not Identified
000100-41-4 / Ethylbenzene	0.0031	0	0.46	0.0031	0.0762	--	--	Not Identified
000108-38-3 / m-Xylene	0.006	0	0.32	0.006	0.0675	--	--	Not Identified
000095-47-6 / o-Xylene	0.004	0	0.21	0.004	0.0468	--	--	Not Identified
000106-42-3 / p-Xylene	0.0052	0	0.17	0.0052	0.0367	--	--	Not Identified
000056-23-5 / Carbon Tetrachloride	0.16343	0	1.73	0.16343	0.572	--	--	Not Identified
000067-66-3 / Chloroform	0.0315	0.004	0.25	0.0315	0.0884	--	--	Not Identified
000107-06-2 / 1,2-Dichloroethane	0.0076	0.0076	0.0076	0.0076	N.A.	--	--	Not Identified
000075-09-2 / Dichloromethane	0.0064	0.0064	0.0064	0.0064	N.A.	--	--	Not Identified
000079-34-5 / 1,1,2,2-Tetrachloroethane	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000127-18-4 / Tetrachloroethylene	0.00186	0	0.071	0.00186	0.0223	--	--	Trichloroethylene
000071-55-6 / 1,1,1-Trichloroethane	0.00355	0	0.041	0.00355	0.0130	--	--	Not Identified
000079-00-5 / 1,1,2-Trichloroethane	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000079-01-6 / Trichloroethylene	0.0016	0.00082	0.04	0.0016	0.00889	--	--	Not Identified
000075-01-4 / Vinyl Chloride	0.00405	0	0.0582	0.00405	0.0139	--	--	Not Identified

Table 2.1 (continued)
Summary of Rate Constants

CAS Number / Chemical Name	Rate Constant (1/day)					Distribution Type	Molar Yield	Reaction Product
	Most Likely Estimate	Min	Max	Median	Standard Deviation			
000108-95-2 / Phenol	0.032	0	0.2	0.032	0.0651	--	--	Not Identified
000095-48-7 / o-Cresol	0.005	0	0.034	0.005	0.0172	--	--	Not Identified
000108-39-4 / m-Cresol	0.029	0.0029	0.033	0.029	0.0138	--	--	Not Identified
000106-44-5 / p-Cresol	0.037	0.035	0.048	0.037	0.007	--	--	Not Identified
000120-83-2 / 2,4-Dichlorophenol	0.016	0	0.12	0.016	0.0501	--	--	Not Identified
000088-06-2 / 2,4,6-Trichlorophenol	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000087-86-5 / Pentachlorophenol	0	0	0	0	0	--	--	Not Identified
000075-69-4 / Trichloroflouromethane (CFC-11)	0.0016	0.0016	0.0016	0.0016	N.A.	--	--	Not Identified
000075-71-8 / Dichlorodifluoromethane (CFC-12)	0	0	0	0	N.A.	--	--	Not Identified
000076-13-1 / 1,1,2-Trichloro-1,2,2-trifluoroethane (CFC 113)	0	0	0	0	N.A.	--	--	Not Identified
000067-64-1 / Acetone	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000078-93-3 / Methyl Ethyl Ketone	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000108-10-1 / Methyl Isobutyl Ketone	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000064-19-7 / Aceic Acid	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000103-32-2 / Phenylacetic Acid	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000083-32-9 / Acenaphthalene	0.0043	0.0043	0.0043	0.0043	N.A.	--	--	Not Identified
000086-73-7 / Fluorene	0.00015	0	0.00145	0.00015	0.00069	--	--	Not Identified

Table 2.1 (continued)
Summary of Rate Constants

CAS Number / Chemical Name	Rate Constant (1/day)					Distribution Type	Molar Yield	Reaction Product
	Most Likely Estimate	Min	Max	Median	Standard Deviation			
000090-12-0 / 1-Methylnaphthalene	0	0	0.057	0	0.0214	--	--	Not Identified
000091-20-3 / Naphthalene	0	0	0.03	0	0.00791	--	--	Not Identified
000085-01-8 / Phenanthrene	N.A.	N.A.	N.A.	N.A.	N.A.	--	--	Not Identified
000092-52-4 / 1,1'-Biphenyl	0.00016	0	0.019	0.00016	0.00944	--	--	Not Identified
000098-82-8 / Cumene	0	0	0	0	N.A.	--	--	Not Identified
000123-91-1 / Dioxane	0	0	0	0	N.A.	--	--	Not Identified
000067-56-1 / Methanol	0.036	0	0.34	0.036	0.0697	--	--	Not Identified
000098-95-3 / NitroBenzene	0.0037	0.0037	0.1168	0.0037	0.0427	--	--	Not Identified
000110-86-1 / Pyridine	0.01	0	0.02	0.01	0.0102	--	--	Not Identified
000100-42-5 / Styrene	0	0	0	0	N.A.	--	--	Not Identified
000108-67-8 / 1,3,5-Trimethylbenzene	0	0	0.0039	0	0.00174	--	--	Not Identified

Notes:

C An N.A. in all columns for a compound indicates no study met the proposed criteria.

C An N.A. in the standard deviation column for a compound indicates that there was only one study that met the proposed criteria and so it is not possible to calculate a standard deviation.

3.0 REFERENCES

Collins , W.D., Temperature of water available for individual use in the United States: U.S. Geol. Surv. Water Supply Papers 520-F, p97-104, 1925.

Krumholz, L.R., Caldwell, M.E., and Suflita, J.M., Biodegradation of 'BTEX' hydrocarbons under anaerobic conditions, pages 61 through 99 in Bioremediation: Principles and Applications, R.L. Crawford and D.L. Crawford, Cambridge University Press, 1996.

Syracuse Research Center. Anaerobic Biodegradation of Organic Chemicals in Groundwater: A Summary of Field and Laboratory Studies, SRC TR-97-0223F, 1997.

U.S. EPA, EPA's Composite Model for Leachate Migration with Transformation Products (EPACMTP): Background Document, Office of Solid Waste, Washington, D.C., 20460, 1996.

APPENDIX A CRITERIA FOR EVALUATING BIODEGRADATION RATE (LABORATORY STUDIES)

Italicized criteria are common with Syracuse Research Corporation's criteria.

A.1 LABORATORY MICROCOSMS

1. **The experiment should have been conducted with aquifer materials and aquifer bacteria.*
2. **A proper control study should have been conducted to separate abiotic losses from biotransformation.*
3. ** Scaling factors for normalizing reaction rate (e.g. dry cell mass) should not have been used. If a scaling factor was used, it should be reported so that it is possible to calculate a first order rate.*
4. ** Concentration decrease with time should be documented in the study.*
5. Experimental Conditions
 - C ** No artificial growth media should be used in the microcosm study.*
 - C ** The experiment should be carried out under anaerobic conditions.*
 - C ** In case of a volatile contaminants, there should be no head space in the flask in which the study is carried out.*
 - C The incubation should be carried out under conditions that closely replicate the subsurface environment in terms of temperature, pH, and light conditions. The incubation should be carried out in the dark.
6. The experiment should have good mass balance. That is, the loss of parent organic carbon should be accounted for by the intermediates and products. Are other products generated in stoichiometric proportions?
7. If the target compounds were administered to the microcosm using a carrier solvent, how was this accomplished? If a carrier solvent was used, this could potentially alter the consumption of electron donors and acceptors. Was a control done to account for the effect of the carrier solvent?
8. The soil/water ratio in the microcosms and the rate constants derived from the data should accurately reflect the in situ soil/water ratio.
9. Time dependency of rate data. Criteria for statistical analysis of data (e.g., questions regarding the degree of replication or standard error).

(To fit a first order model ideally there should be a concentration decrease of at least two orders of magnitude. However, it is recognized that in most cases it is possible to get only an order of magnitude decrease.)

A.2 BATCH AND COLUMN STUDIES:

1. The column should be packed and operated under anaerobic conditions. The column must be large enough to prevent wall effects (i.e. column diameter 20 times the grain diameter).
2. The chemistry of the water passed through the column (or batch reactor) must be similar to the composition of the groundwater at the site.
3. * A tracer study must be conducted to characterize the hydraulic properties of the column (e.g., mean fluid residence time and hydrodynamic dispersion).
4. The flow velocity through the column/batch reactor should mimic the groundwater flow velocity at the site.
5. The feed reservoirs, feed lines, and effluent lines need to be periodically sterilized to prevent/minimize microbial contamination of the column.
6. The column needs to be operated long enough to obtain stable effluent concentrations in order to establish proper breakthrough and steady-state reaction processes.
7. In order to accurately assess the biodegradation rate within the column, samples along the column length (using multiple side ports) must be collected to determine the concentration profile(s) within the column (i.e., concentration versus length).

These criteria are in addition to the criteria for evaluating laboratory microcosm studies.

*Criteria for acceptance/rejection of study.

APPENDIX B
CRITERIA FOR EVALUATING BIODEGRADATION RATE
(FIELD STUDIES)

1. * *The study should be conducted in the anaerobic portion of the plume.*
2. * *A tracer study should be carried out that can be used to estimate the processes of dilution, sorption, or volatilization that can contribute to attenuation of contaminants.*
3. * *If a transport model was employed to estimate a biodegradation rate, reasonable parameter values should be used to describe dispersion and sorption processes. The model used should be appropriate to the hydrogeologic setting and should have been verified.*
4. **Data on concentration decrease with time should be reported in the study.*
5. No scaling factor should be used to normalize the data. If a factor is used the study must document the factor so that a first order rate can be calculated.
6. Time dependency of rate data. Criteria for statistical analysis of data (e.g., questions regarding the degree of replication or standard error).

(To fit a first order model ideally there should be a concentration decrease of at least two orders of magnitude. However, it is recognized that in most cases it is possible to get only an order of magnitude decrease.)

APPENDIX C
CRITERIA FOR EVALUATING BIODEGRADATION RATE STUDIES
(Syracuse Research Center)

LABORATORY MICROCOSMS

1. Study Conducted Under Anaerobic Conditions
2. Experiment Conducted With Grab Samples of Aquifer Material or Ground Water
3. Study Has Controls to Determine Abiotic Losses
4. Need Data on Concentration Decrease With Time

FIELD STUDIES

1. Studies of Anaerobic Portion of Aquifer System
2. Suitable Conservative Tracer Other Than Target Compound Available to Correct for Dilution, Sorption, Advection and Dispersion Processes
3. Need Data on Concentration Decrease With Time/Distance (Need flow velocity to correlate distance with time)

APPENDIX D
LABORATORY STUDIES FOR ALL COMPOUNDS

Draft

Table D.1
Summary Table for Anaerobic Biodegradation Rate Constants for Benzene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0	20	0 ¹	15	¹ No Redox Regime Given
	6-8	0.0015 ²	9			² Meth/Fe/Mn
		0 ³	2	0.047*	11	³ Meth/SO ₄
						*8 days lag
		0.052*	11			* 21 days lag
		0.071 ¹⁴	27	0 ¹¹	3	¹⁴ Meth/Fe
						¹¹ No Redox Regime Given
		0	19	0.0029 ¹⁰	17	¹⁰ No Redox Regime Given
		0.0 ⁶ (0; 0)	1	0.0065 ¹³	28	⁶ 1 site 2 studies of Meth/SO ₄ and Meth/NO ₃
						¹³ Fe
	0 ⁷	18	0 ¹²	2	⁷ Meth/Fe/NO ₃	
	3			¹² Fe		
> 8	No Studies		No Studies			

Table D.1 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Benzene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	No Studies		No Studies		
	6-8	0	7	0.0041 ⁸	6	⁸ SO ₄ /Fe; 121 days lag
		0	14	0.003	8	
				0.0237 ¹⁵	6	¹⁵ SO ₄ /Fe; 184 days lag
		0	8	0	4	
		0.0074	26			*140 days lag
	0.0 ¹⁷ (0; 0)	13	¹⁷ Meth/SO ₄			
> 8	No Studies		No Studies			

Table D.2
Summary Table for Anaerobic Biodegradation Rate Constants for Toluene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments	
		Rate (1/day)	Ref.	Rate (1/day)	Ref.		
# 15	< 6	No Studies		No Studies			
	6-8	0.043 ¹²	27	0.016 ¹³	10	¹² Meth/Fe	
							¹³ No Redox Regime Given
		0.093 ¹⁴	8	0.087 ¹⁵	14	¹⁴ Meth/Fe/Mn	
							¹⁵ Fe
		0 ¹⁶	19	0.0045 ¹⁷	17	¹⁶ Meth/Fe/NO ₃	
							¹⁷ No Redox Regime Given
		0 ²¹ (0 ²² ; 0)	1	0.010 ²⁰	28	²¹ 1 site; 2 studies one with ²² Meth/NO ₃	
						²⁰ Fe	
0.10 ²⁴	2	²⁴ Meth/Fe/Mn					
> 8	No Studies		No Studies				

Table D.2 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Toluene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	No Studies		No Studies		
	6-8	0	7	0.11	3	
		0.034 ²⁵ (0.036* ¹ ; 0; 0.032* ² ; 0.06* ³ ; 0.012; 0.063)	11	0.0446 ²⁶	9	²⁵ 1 site; 6 studies; * ¹ 208 days lag; * ² 99 days lag * ³ 302 days lag
						²⁶ SO ₄ /Fe
		0.028 ²⁸ (0.036; 0.020)	5	0.011	22	²⁸ 1 site; 2 studies
		0.0098	4			²⁹ No Redox Regime Given
		0	12			
		0.018	28			
	0	14				
> 8	No Studies		No Studies			

Table D.3
Summary Table for Anaerobic Biodegradation Rate Constants for Ethylbenzene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ⁹	2	0	2	⁹ Meth/SO ₄
	6-8	0.0055 ¹⁰	7	0 ¹¹	9	¹⁰ Meth/Fe/Mn
		0 ¹² (0 ^{12a} ; 0 ^{12b})	1			¹¹ No Redox Regime Given
	> 8	No Studies		No Studies		¹² 1 site 2 studies; ^{12a} Meth; ^{12b} Meth/NO ₃
> 15	< 6	No Studies		No Studies		
	6-8	0	10	0.0019 ¹⁴	8	¹⁴ SO ₄ /Fe
		0.0029	5			*140 days lag
		0.0076*	16			
		0.29 ¹⁵ (0.12; 0.46)	6			¹⁵ 1 site; 2 studies
> 8	No Studies		No Studies			

Table D.4
Summary Table for Anaerobic Biodegradation Rate Constants for m-Xylene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ¹⁰	2	0 ¹¹	2	¹⁰ Meth/SO ₄
						¹¹ Fe
	6-8	0	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0	7	0.0204 ¹²	6	¹² SO ₄ /Fe
		0.0006	5	0.17*	3	*17 days lag
	> 8	No Studies		No Studies		

Table D.5
Summary Table for Anaerobic Biodegradation Rate Constants for o-Xylene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		0 ⁷	2	⁷ - Fe reducing
	6-8	0	1	0 ⁶	4	⁶ - No redox regime given
		0.071 ⁸	19			⁸ - Meth/Fe
		0.0091 ⁹	8			⁹ -Meth/Fe/Mn
		0 ¹⁰	13			¹⁰ -Meth/Fe/NO ₃
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0006	6	0	3	
		0	10	0.00559* ¹¹	5	* ¹¹ - SO ₄ /Fe; 37 days lag
		0.0087*	18			* - 140 days lag
	> 8	No Studies		No Studies		

Table D.6
Summary Table for Anaerobic Biodegradation Rate Constants for p-Xylene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ⁷	1	0 ⁹	1	⁷ - Meth/SO ₄
						⁹ - Fe
	6-8	0.057 ⁸	12	No Studies		⁸ - Meth/Fe
		0.0006	3			
> 8	No Studies		No Studies			
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0.17*	2	* - 21 days lag
	> 8	No Studies		No Studies		

Table D.7
Summary Table for Anaerobic Biodegradation Rate Constants for Carbon Tetrachloride According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.23	2	0 ³	2	³ Fe/NO ₃ /Mn
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.8
Summary Table for Anaerobic Biodegradation Rate Constants for Chloroform According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.01562 ² (0.025 ^{*a} ; 0.02 ^{*a} ; 0.025 ^{*a} ; 0.004 ^{*b} ; 0.0041 ^{*b})	2	² 1 site; 5 studies with No Redox Regime Given ^{*a} 56 days lag ^{*b} 112 days lag
	6-8	No Studies		0.142 ³ (0.099; 0.25 [*] ; 0.033; 0.099; .25 [*] ; .12)	2	³ 1 site; 6 studies with No Redox Regime Given [*] 14 days lag
	> 8	No Studies		No Studies		

Table D.9
Summary Table for Anaerobic Biodegradation Rate Constants for 1,2-Dichloroethane According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.10
Summary Table for Anaerobic Biodegradation Rate Constants for Dichloromethane According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.11
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 1, 2, 2, -Tetrachloroethane According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.12
Summary Table for Anaerobic Biodegradation Rate Constants for Tetrachloroethylene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.071 ⁶	2	No Studies		⁶ Meth/SO ₄
		0 ⁷	3			⁷ Meth/Fe/NO ₃ /Mn
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.00073	2	
	6-8	0.0084	2	0.054 ⁸	6	⁸ No Redox Regime Given
				0.0065 ⁹ (0; 0.013)	9	⁹ 1 site; 2 studies No Redox Regime Given
	> 8	No Studies		No Studies		

Table D.13
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,1-Trichloroethane According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0037	4	0 ⁶	4	⁶ Fe/NO ₃ /Mn
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0092 ⁷ (0.015; 0.0034)	1	0.0092 ⁸ (0.015; 0.0034)	1	⁷ 1 site; 2 studies
						⁸ 1 site; 2 studies
				0.0099 ⁹	6	⁹ No Redox Regime Given
> 8	No Studies		No Studies			

Table D.14
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 1, 2-Trichloroethane According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.15
Summary Table for Anaerobic Biodegradation Rate Constants for Trichloroethylene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ¹⁴	8	0.010 ¹⁵	19	¹⁴ Meth/Fe/NO ₃ /Mn ¹⁵ Fe
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0 ¹⁶ (0; 0; 0; 0)	10	¹⁶ 1 site; 4 studies with No Redox Regime Given
				0.0011 ¹⁷	18	¹⁷ SO ₄ /Fe
	6-8	0.016 ¹⁹ (0.011-0.021)	1	0.0015 ¹⁸ (0.0017; 0; 0.0029)	18	¹⁸ 1 site; 3 studies with SO ₄ /Fe ¹⁹ Meth/Fe/SO ₄
		0.039 ²⁰ (0.038* ¹ ; 0.040* ²)	7	0.0090 ²¹ (0.0077; 0.0082-0.011)	1	²⁰ 1 site; 2 studies; * ¹ 110 days lag; * ² 108 days lag
		0.013* (0.0020-0.024)	17	0.0029 ²² (0.00057-0.005)	4	* 112 days lag ²² SO ₄ /Fe
				0.010	19	
	> 8	No Studies		No Studies		

Table D.16
Summary Table for Anaerobic Biodegradation Rate Constants for Vinyl Chloride According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0.0069 ³ (0.0082; 0.0057)	1	³ 1 site; 2 studies with No Redox Regime Given
				0.0082 ⁴	1	⁴ - No Redox Regime Given
	> 8	No Studies		No Studies		

Table D.17
Summary Table for Anaerobic Biodegradation Rate Constants for Phenol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		0.20 ²	7	² - No redox regime given
	6-8	0 ³	6	0 ⁴	4	⁴ - No redox regime given ³ - Meth/Fe/NO ₃
	> 8	No Studies		No Studies		
> 15	< 6	0.13*	3	No Studies		* - 50 days lag
	6-8	0.0695 ⁶ (0.068*, 0.071**)	1, 2	No Studies		⁶ 1 site; 2 studies, * - 40 days lag, ** - 30 days lag
		0.10 ⁵	5			
	> 8	No Studies		No Studies		

Table D.18
Summary Table for Anaerobic Biodegradation Rate Constants for o-Cresol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ²	4	0 ³	3	² - Meth/Fe/NO ₃
	> 8	No Studies		No Studies		³ - No redox regime given
> 15	< 6	No Studies		No Studies		
	6-8	0.0050	5	No Studies		⁴ 1 site; 2 studies * - 100 days lag
		0.033 ⁴ (0.032*; 0.034*)	2, 1			
> 8	No Studies		No Studies			

Table D.19
Summary Table for Anaerobic Biodegradation Rate Constants for m-Cresol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0203 ¹ (0.0029, 0.029*, 0.029*)	3, 2,1	No Studies		¹ 1 site; 3 studies * - 100 days lag
	> 8	No Studies		No Studies		

Table D.20
Summary Table for Anaerobic Biodegradation Rate Constants for p-Cresol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.036 ¹ (0.035*, 0.037*)	1, 2	No Studies		¹ 1 site; 2 studies * - 100 days lag
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.21
Summary Table for Anaerobic Biodegradation Rate Constants for 2,4-Dichlorophenol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.015	2	No Studies		
	6-8	0.017	2	0 ³		³ -Fe/NO ₃
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.090	5	0.0055	1	
		0.12 ⁴	3	0	5	⁴ - Meth/SO ₄
		0.12	1			² No Redox Regime Given
	> 8	No Studies		No Studies		

Table D.22
Summary Table for Anaerobic Biodegradation Rate Constants for 2,4,6-Trichlorophenol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.23
Summary Table for Anaerobic Biodegradation Rate Constants for Pentachlorophenol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	1.98e-03 ¹ (1.61e-03-2.36e-03)	1	No Studies		¹ - 1 site 2 studies
		1.414e-03 ¹ (1.01e-03-2.17e-03; 6.86e-04-1.79e-03)	1			
	6-8	1.97e-03 ³ (1.60e-03-2.42e-03; 1.06e-03-2.80e-03)	1	No Studies		³ - 1 site 2 studies
	> 8	No Studies		No Studies		

Table D.23 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Pentachlorophenol According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	4.94e-03 (7.38e-03 - 2.50e-03)	1	0 ²	3	² No Redox Regime Given
	6-8	0(0; 0)	4	No Studies		⁴ - 1 site 2 studies
		5.09e-03 ⁵ (2.91e-03- 7.27e-03)	1			⁵ - 1 site 2 studies
		5.75e-03 ⁴ (1.63e-03- 2.15e-03; 0.0162-3.0e-03)	1			
> 8	No Studies		No Studies			

Table D.24
Summary Table for Anaerobic Biodegradation Rate Constants for Trichlorofluoromethane According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.25
Summary Table for Anaerobic Biodegradation Rate Constants for Dichlorodifluoromethane According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.26
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,2-Trichloro-1,2,2-trifluoroethane According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.27
Summary Table for Anaerobic Biodegradation Rate Constants for Acetone According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.28
Summary Table for Anaerobic Biodegradation Rate Constants for Methyl Ethyl Ketone According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.29
Summary Table for Anaerobic Biodegradation Rate Constants for Methyl Isobutyl Ketone According to EPA Protocol (Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.30
Summary Table for Anaerobic Biodegradation Rate Constants for Acetic Acid According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.31
Summary Table for Anaerobic Biodegradation Rate Constants for Phenylacetic Acid According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.32
Summary Table for Anaerobic Biodegradation Rate Constants for Acenaphthene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.33
Summary Table for Anaerobic Biodegradation Rate Constants for Fluorene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0	2	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0 ³	3	No Studies		³ - Meth/NO ₃ /SO ₄
	> 8	No Studies		No Studies		

Table D.34
Summary Table for Anaerobic Biodegradation Rate Constants for 1 - Methylanthalene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ⁴	1	0 ⁵	1	⁴ Meth/SO ₄
						⁵ Fe
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.35
Summary Table for Anaerobic Biodegradation Rate Constants for Naphthalene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ⁹	2	0 ¹⁰	2	⁹ Meth/SO ₄
						¹⁰ Fe
	6-8	0 ¹¹	10	0 ¹²	1	¹¹ Meth/Fe/NO ₃
						¹² No Redox Regime Given
		0 ¹³	3	0 ¹⁴	6	¹³ Meth/Fe/Mn
						¹⁴ No Redox Regime Given
> 8	No Studies		No Studies			
> 15	< 6	No Studies		No Studies		
	6-8	0	9	0	8	
	> 8	No Studies		No Studies		

Table D.36
Summary Table for Anaerobic Biodegradation Rate Constants for Phenanthrene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.37
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1' Biphenyl According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ⁴	2	No Studies		⁴ - Meth/Fe/NO ₃
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.38
Summary Table for Anaerobic Biodegradation Rate Constants for Cumene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		0 ²	1	² No Redox Regime Given
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.39
Summary Table for Anaerobic Biodegradation Rate Constants for Dioxane According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.40
Summary Table for Anaerobic Biodegradation Rate Constants for Methanol According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0067	5	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.034 ¹ (0.043; 0.0022; 0.0089; 0.0039; 0.072; 0.072)	4	¹ - 1 Site 6 studies; No redox regime given
				0.034 ⁴ (0.028*; 0.030*; 0.054*; 0.036*; 0.016; 0.029*; 0.039; 0.039)	1, 6, 7	⁴ 1 site; 8 studies *-No redox regime given
	6-8	0.089 ⁵	3	0.071 ²	1	² - No redox regime given
				0.067 ³	1	³ - No redox regime given
				0.34	3	⁵ Meth/SO ₄
				0.058 (0.1*; 0.016**)	4	* - 24 days lag
						** - 30 days lag
	> 8	No Studies		No Studies		

Table D.41
Summary Table for Anaerobic Biodegradation Rate Constants for Nitrobenzene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0037	1	0.0602 ² (0.0037 ^{2a} ; 0.0037- 0.23 ^{2a})	1	² - 1 site 2 studies ^{2a} - Fe reducing
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.42
Summary Table for Anaerobic Biodegradation Rate Constants for Pyridine According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.010 ¹ (0.020*, 0)	2, 1	0.014**	2	¹ 1 site; 2 studies * - 90 days lag
	> 8	No Studies		No Studies		** - 30 days lag

Table D.43
Summary Table for Anaerobic Biodegradation Rate Constants for Styrene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table D.44
Summary Table for Anaerobic Biodegradation Rate Constants for 1,3,5-Trimethylbenzene According to EPA Protocol
(Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ¹	1	0 ²	1	¹ Meth/SO ₄
						² Fe
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

**APPENDIX E
FIELD STUDIES FOR ALL COMPOUNDS**

Draft

Table E.1
Summary Table for Anaerobic Biodegradation Rate Constants for Benzene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ¹	16	No Studies		¹ Meth/SO ₄ /Fe
	6-8	0	21	0.0063 ⁷	31	⁷ No Redox Regime Given
		0.002 ³ (0-0.002;0.002-0.004)	7	0.00134 ⁴ (0.0017; 0.0022; 0.00011)	28	³ 1 site 2 studies for Meth/NO ₃ /SO ₄
		0.017 ⁵	10			⁴ 1 site 3 studies for Fe
		0 ⁶	22			⁵ Meth/Fe/Mn
		0.0001075 ² (0; 0; 0; 0.00043)	30			² 1 site; 4 studies for Meth/NO ₃ /SO ₄
		0.00714	29			
	> 8	No Studies				No Studies
> 15	< 6	No Studies		0 ¹⁰	23	¹⁰ SO ₄ /Fe
	6-8	0.005 ⁹ (0.01; 0)	25	0.0308 ⁸ (0.0072-0.046; 0.038; 0.028)	25	⁹ 1 site; 2 studies
		0	30			⁸ 1 site; 3 studies
	> 8	No Studies		No Studies		

Table E.2
Summary Table for Anaerobic Biodegradation Rate Constants for Toluene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.043 ¹	15	0 ¹¹	16	¹¹ No Redox Regime Given
		0	18			¹ Meth/SO ₄ /Fe
	6-8	0.042 ² (0.067; 0.026; 0.053; 0.023)	30	0.0024 ³ (0.00099; 0.0039; 0.0023)	28	² 1 site; 4 studies for Meth/NO ₃ /SO ₄
		0.186	27	0.019 ⁴	31	⁴ No Redox Regime Given
		0.0532 ¹⁸ (0.1 ¹⁹ ; 0.0064)	1	0.066	20	¹⁸ 1 site; 2 studies one with ¹⁹ Meth/SO ₄ /Fe
		0.042 ⁷ (0.053-0.067; 0.023-0.026)	4	0.083 ²³	1	⁵ Meth/SO ₄ /Fe
						⁷ 1 site; 2 studies for Meth/NO ₃ /SO ₄
					²³ NO ₃ /SO ₄	
> 8	No Studies		No Studies			
> 15	< 6	No Studies		0.0086 ⁸ (0.0052-0.012)	23	⁸ SO ₄ /Fe
	6-8	0.003	24	0.027 ⁹ (0.023; 0.031)	25	⁹ 1 site; 2 studies
				0.045	24	
		0.032 ¹⁰ (0.05; 0.013)	29	0.091	6	¹⁰ 1 site; 2 studies
	0.011 ²⁷	9	0.018 (0.0075-0.03)	7	²⁷ SO ₄ /Fe	
> 8	No Studies		No Studies			

Table E.3
Summary Table for Anaerobic Biodegradation Rate Constants for Ethylbenzene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0066 ¹ (0.011; 0.0024; 0.0099; 0.0031)	19	0.00187 ² (0.0032; 0.00060; 0.0018)	17	¹ 1 site; 4 studies for Meth/NO ₃ /SO ₄
						² 1 site; 3 studies for Fe
		0.00675 ³ (0.003-0.011; 0.003-0.010)	5	0.019 ⁶	20	³ 1 site; 2 studies for Meth/NO ₃ /SO ₄
		0.025 ¹⁵ (0.0011 ^{15a} ; 0.028* ^{15a} ; 0 ^{15b} ; 0.0053 ^{15c} ; 0.067 ^{15c})	1			¹⁵ 1 site; 4 studies; ^{15a} Meth; ^{15b} Meth/NO ₃ ; ^{15c} Meth/SO ₄ ; * 4 days lag
						⁵ Meth/SO ₄ /Fe
		0	12			⁶ No Redox Regime Given
		0 ⁷	13			⁷ Meth/SO ₄ /Fe
	0.024 ⁵	11				
> 8	No Studies		No Studies			
> 15	< 6	No Studies		No Studies		
	6-8	0.0031	14	0.012 ⁴ (0; 0.029; 0.009; 0.010)	14, 15	⁴ 1 site; 4 studies
		0.04 ⁸ (0.03; 0.05)	18	0 ¹³	4	¹³ SO ₄ /Fe
					⁸ 1 site; 2 studies	
> 8	No Studies		No Studies			

Table E.4
Summary Table for Anaerobic Biodegradation Rate Constants for m-Xylene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.057 ¹	8	No Studies		¹ Meth/SO ₄ /Fe
	6-8	0.0077 ² (0.0037; 0.014; 0.0083; 0.0046)	13	0.00203 ³ (0.0033; 0.0012; 0.0016)	14	² 1 site; 4 studies for Meth/NO ₃ /SO ₄
						³ 1 site; 3 studies for Fe
		0.121 ⁴ (0; 0.32 ^{4a} ; 0.044)	1	0.019 ⁵	15	⁴ 1 site; 3 studies ^{4a} Meth/SO ₄
						⁵ No Redox Regime Given
		0 ⁶	10			⁶ Meth/SO ₄ /Fe
	0.019 ⁷ (0.004-0.008; 0.005-0.014)	5	⁷ 1 site; 2 studies for Meth/NO ₃ /SO ₄			
> 8	No Studies		No Studies			
> 15	< 6	No Studies		No Studies		
	6-8	0.003	11	0.024	11	
		0.06 ⁸ (0.02; 0.1)	12	0.050	9	⁸ 1 site; 2 studies
	0.0143 ⁹			6	⁹ SO ₄ /Fe 121 days lag	
> 8	No Studies		No Studies			

Table E.5
Summary Table for Anaerobic Biodegradation Rate Constants for o-Xylene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.029 ⁶	11	0 ¹	12	¹ - No redox regime given
						⁶ - Meth/SO ₄ /Fe
	6-8	0.0043	19	0.063 ²	23	² - No redox regime given
		0	13	0.0015 ³ (0.0022; 0.0015; 0.0082)	20	³ - Fe reducing
		0.00795 ⁴ (0.004-0.011; 0.009- 0.016; 0.004; 0.015; 0.0011; 0.0086)	6, 22			⁴ - 1 site 6 studies for Meth/NO ₃ /SO ₄
		0 ⁵	16			⁵ - Meth/SO ₄ /Fe
> 8	No Studies		No Studies			
> 15	< 6	0	14	No Studies		
	6-8	0.21	21	0.16*	7	* - 17 days lag
		0.003	17	0.077*	15	* - 17 days lag
				0.02	17	
				0	9	
	> 8	No Studies		No Studies		

Table E.6
Summary Table for Anaerobic Biodegradation Rate Constants for p-Xylene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.057 ¹	5	No Studies		¹ - Meth/SO ₄ /Fe
	6-8	0.0043	12	0.019 ⁴	13	² - Meth/SO ₄ /Fe
		0 ²	6			⁴ - No redox regime given
		0.007 ³ (0.005-0.014; 0.002-0.010; 0.0094; 0.0024; 0.0051; 0.0096)	3, 10	0.0015 ⁵ (0.00085; 0.0019; 0.0018)	9	³ - 1 Site 6 studies for Meth/NO ₃ /SO ₄
	> 8	No Studies		No Studies		⁵ - 1 Site 3 studies for Fe
> 15	< 6	No Studies		No Studies		
	6-8	0.05 (0.02;0.08)	10	0.016 ⁶ (0; 0.032)	8	⁶ - 1 Site 2 studies
		0.0029	8	0.0143*	4	* - 121 days lag
		0.0053 (0.0023- 0.0083)		7		
> 8	No Studies		No Studies			

Table E.7
Summary Table for Anaerobic Biodegradation Rate Constants for Carbon Tetrachloride According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.11	1	No Studies		
	6-8	0.32 (0.15-0.49)	2	0.0602 ¹ (0.0037; 0.0037-0.23)	2	¹ 1 site; 2 studies for Fe
		0.21	1			
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		1.73 ²	3	² NO ₃ /SO ₄
	> 8	No Studies		No Studies		

Table E.8
Summary Table for Anaerobic Biodegradation Rate Constants for Chloroform According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0.03	1	¹ No Redox Regime Given *20-30 days lag
	> 8	No Studies		No Studies		

Table E.9
Summary Table for Anaerobic Biodegradation Rate Constants for 1,2-Dichloroethane According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0076 (0.0042-0.011)	1	No Studies		
	> 8	No Studies		No Studies		

Table E.10
Summary Table for Anaerobic Biodegradation Rate Constants for Dichloromethane According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0064	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.11
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 1, 2, 2, -Tetrachloroethane According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.12
Summary Table for Anaerobic Biodegradation Rate Constants for Tetrachloroethylene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0	4	No Studies		
	6-8	0.000735 (0.00068- 0.00079)	1	0.00405 ¹ (0.0035- 0.0046)	5	¹ NO ₃ /Fe/SO ₄ /Meth
		0 ²	8	0 ³	3	² Meth/SO ₄ /Fe
						³ Fe/NO ₃ /Mn
	0.0109 ⁴ (0; 0.0097-0.034)	3		⁴ 1 site; 2 studies		
> 8	No Studies		No Studies			
> 15	< 6	No Studies		No Studies		
	6-8	0.003 ⁵	7	No Studies		⁵ No Redox Regime Given
	> 8	No Studies		No Studies		

Table E.13
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,1-Trichloroethane According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.011 ¹ (0; 0.022)	3	No Studies		¹ 1 site; 2 studies
	6-8	0.0013	1	0 ⁴	4	⁴ Fe/NO ₃
		0 ²	8	0.010 ³	5	² Meth/SO ₄ /Fe
		0.041*	5			³ Fe
		0.0375 (0.029-0.046)	4			*30 days lag
	> 8	No Studies				No Studies
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0.003 ⁵	7	⁵ No Redox Regime Given
	> 8	No Studies		No Studies		

Table E.14
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,2 -Trichloroethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.15
Summary Table for Anaerobic Biodegradation Rate Constants for Trichloroethylene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0	9	No Studies		
	6-8	0.00062 (0.00045-0.00079)	5	0.0023 ¹ (0.0017-0.0029)	4	¹ SO ₄ /Fe
		0	12	0.0036 ² (0.00014-0.0071)	3	² SO ₄ /Fe
		0.0034 ⁴ (0.0047;0.00082; 0.0047)	15	³ 0.00086	6	³ Fe/ Meth/SO ₄
						⁴ 1 site; 3 studies for Meth/SO ₄
		0.0023 ⁶ (0.0011;0.0034; 0.0010;0.0036; 0.0025)	20, 21	0.00074 ⁵ (0.00019-0.0024; 0.00014-0.00024)	13	⁵ 1 site; 2 studies with No Redox Regime Given
						⁶ 1 site; 5 studies for Meth/SO ₄
		0 ⁷	8	0.00137 ⁸ (0.00082; 0.0033; 0.0014)	16	⁷ Meth/Fe/SO ₄
						⁸ 1 site; 3 studies for NO ₃ /Fe/SO ₄ /Meth
			⁹ (0.0016; 0.0015; 0.0011)	17	⁹ 1 site; 3 studies for Fe	
¹⁰ 1 site; 3 studies for SO ₄ /Fe						
		0.0033 ¹⁰ (0.0038; 0.0033; 0.0027)	21			
> 8	No Studies		No Studies			

Table E.15 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Trichloroethylene According to EPA Protocol (Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
	6-8	No Studies		0.00069 ¹¹ (0.00059-0.00079)	14	¹¹ Fe/Meth/SO ₄
				0.003 ¹²	11	¹² No Redox Regime Given
				0.0015 ¹³ (0.0026; 0.0005)	2	¹³ 1 site; 2 studies with No Redox Regime Given
	> 8	No Studies		No Studies		

Table E.16
Summary Table for Anaerobic Biodegradation Rate Constants for Vinyl Chloride According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.00093 (0.00086-0.0010)	2	0.0008 ¹ (0.0012; 0.0013; 0)	3	¹ - 1 site 3 studies for NO ₃ /Fe/SO ₄ /Meth
		0.00297 ² (0.0024; 0.0060; 0.00049)	4			² 1 site; 3 studies for Meth/SO ₄
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.17
Summary Table for Anaerobic Biodegradation Rate Constants for Phenol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0	6	0.0135 ¹ (0; 0.027*)	6	¹ 1 Site 2 studies of Fe, * - 0-70 days lag
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.032	2	No Studies		
	> 8	No Studies		No Studies		

Table E.18
Summary Table for Anaerobic Biodegradation Rate Constants for o-Cresol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ¹	4	No Studies		¹ - Meth/Fe/NO ₃
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.034	2	No Studies		
	> 8	No Studies		No Studies		

Table E.19
Summary Table for Anaerobic Biodegradation Rate Constants for m-Cresol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.033	2	No Studies		
	> 8	No Studies		No Studies		

Table E.20
Summary Table for Anaerobic Biodegradation Rate Constants for p-Cresol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.048	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.21
Summary Table for Anaerobic Biodegradation Rate Constants for 2,4-Dichlorophenol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.027	4	0 ¹	4	¹ - Fe/NO ₃
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.22
Summary Table for Anaerobic Biodegradation Rate Constants for 2,4,6-Trichlorophenol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.033	2	No Studies		
	> 8	No Studies		No Studies		

Table E.23
Summary Table for Anaerobic Biodegradation Rate Constants for Pentachlorophenol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0	2	No Studies		
	> 8	No Studies		No Studies		

Table E.24
Summary Table for Anaerobic Biodegradation Rate Constants for Trichlorofluoromethane According to EPA Protocol (Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0016	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.25
Summary Table for Anaerobic Biodegradation Rate Constants for Dichlorodifluoromethane According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		0	1	¹ No Redox Regime Given
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.26
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,2-Trichloro-1,2,2-trifluoroethane According to EPA Protocol (Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.27
Summary Table for Anaerobic Biodegradation Rate Constants for Acetone According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.28
Summary Table for Anaerobic Biodegradation Rate Constants for Methyl Ethyl Ketone According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.29
Summary Table for Anaerobic Biodegradation Rate Constants for Methyl Isobutyl Ketone According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.30
Summary Table for Anaerobic Biodegradation Rate Constants for Acetic Acid According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.31
Summary Table for Anaerobic Biodegradation Rate Constants for Phenylacetic Acid According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.32
Summary Table for Anaerobic Biodegradation Rate Constants for Acenaphthene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0043	1	No Studies		
	> 8	No Studies		No Studies		

Table E.33
Summary Table for Anaerobic Biodegradation Rate Constants for Fluorene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.00030 ¹	1	¹ - No redox regime given
	6-8	No Studies		0.00145 ² (0.0013 - 0.0016)	4	² - No redox regime given
	> 8	No Studies		No Studies		

Table E.34
Summary Table for Anaerobic Biodegradation Rate Constants for 1 - Methyl-naphthalene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.057 ¹	4	No Studies		¹ Meth/SO ₄ /Fe
	6-8	0	5	0.00031 ²	2	² No Redox Regime Given
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0		0.00054 ³ (0.00040-0.00068)		³ No Redox Regime Given
	> 8	No Studies		No Studies		

Table E.35
Summary Table for Anaerobic Biodegradation Rate Constants for Naphthalene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.026 ¹	7	No Studies		¹ Meth/SO ₄ /Fe
		0	11			
	6-8	0 ⁵	10	0.0063 ⁴	15	⁴ No Redox Regime Given
		0	12	0 ⁶	8	⁶ No Redox Regime Given
				0.0018 ⁷ (0.0015-0.0021)	14	⁷ No Redox Regime Given
				0.0050 ⁸	12	⁸ Fe
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.03 ² (0.017-0.043)	13	² SO ₄ /Fe
				0.00018 ³	4	³ No Redox Regime Given
	6-8	0	5	No Studies		
	> 8	No Studies		No Studies		

Table E.36
Summary Table for Anaerobic Biodegradation Rate Constants for Phenanthrene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.37
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1' Biphenyl According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ³	2	0.019 ¹	3	¹ - No redox regime given
	> 8	No Studies		No Studies		³ - Meth/Fe/NO ₃
> 15	< 6	No Studies		0.00032 ²	1	² - No redox regime given
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.38
Summary Table for Anaerobic Biodegradation Rate Constants for Cumene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ¹	1	No Studies		¹ Meth/SO ₄
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.39
Summary Table for Anaerobic Biodegradation Rate Constants for Dioxane According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.40
Summary Table for Anaerobic Biodegradation Rate Constants for Methanol According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.41
Summary Table for Anaerobic Biodegradation Rate Constants for Nitrobenzene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0037	1	0.0037 ¹ (0.0037 ^{1a} ; 0.0037 ^{1a} ; 0.0037 ^{1b})	1	¹ - 1 site 3 studies ^{1a} - NO ₃ /Mn ^{1a} - Fe reducing
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.42
Summary Table for Anaerobic Biodegradation Rate Constants for Pyridine According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table E.43
Summary Table for Anaerobic Biodegradation Rate Constants for Styrene According to EPA Protocol
(Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0 ¹	1	¹ No Redox Regime Given
	> 8	No Studies		No Studies		

Table E.44
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 3, 5-Trimethylbenzene According to EPA Protocol (Field Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0	4	0	3	
				0.0039	2	
	> 8	No Studies		No Studies		

APPENDIX F
LABORATORY AND FIELD STUDIES FOR ALL COMPOUNDS

Draft

Table F.1
Summary Table for Anaerobic Biodegradation Rate Constants for Benzene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments		
		Rate (1/day)	Ref.	Rate (1/day)	Ref.			
# 15	< 6	0	20	0 ¹	15	¹ No Redox Regime Given		
		0 ¹⁸	16			¹⁸ Meth/SO ₄ /Fe		
	6-8	0.0015 ¹	9	0.0041	6	² Meth/Fe/Mn		
		0	21	0.0063 ¹⁹	31	¹⁹ No Redox Regime Given		
		0.002 ²⁰ (0-0.002;0.002-0.004)	7	0.00134 ²¹ (> 0.0017; 0.0022; 0.00011)	28	²⁰ 1 site 2 studies for Meth/NO ₃ /SO ₄		
						²¹ 1 site 3 studies for Fe		
		0.017 ²²	10			²² Meth/Fe/Mn		
		0 ²⁴	22			²⁴ Meth/SO ₄ /Fe		
		0.0001075 ²³ (0; 0; 0; 0.00043)	30			²³ 1 site; 4 studies for Meth/NO ₃ /SO ₄		
		0.00714	28					
		0 ³	2			0.047*	11	³ Meth/SO ₄
								*8 days lag
	0.052*	11			*21 days lag			
	0.071 ¹⁴	27	0 ¹¹	3	¹¹ No Redox Regime Given			
					¹⁴ Meth/Fe			

Table F.1 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Benzene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15 (cont.)	6-8 (cont.)	0.0 ⁶ (0; 0)	1	0.0065 ¹³	28	⁶ 1 site 2 studies of Meth/SO ₄ and Meth/NO ₃
						¹³ Fe
		0 ⁷	18	0 ¹²	2	⁷ Meth/Fe/NO ₃
	0.031	3			¹² Fe	
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0	23	
	6-8	0	7	0.0308 ²⁶ (0.0072-0.046; 0.038; 0.028)	25	²⁶ 1 site; 3 studies
				0.0041 ⁸	6	⁸ SO ₄ /Fe; 121 days lag
				0.0237 ¹⁵	6	¹⁵ SO ₄ /Fe; 184 days lag
		0	14	0	4	
		0	8	0.003	8	
		0.0074*	26			* 140 days lag
		0.0 ¹⁷ (0;0)	13			¹⁷ Meth/SO ₄
		0.005 ²⁵ (0.01; 0)	25			²⁵ 1 site; 2 studies
	0	30				
	> 8	No Studies		No Studies		

Table F.2
Summary Table for Anaerobic Biodegradation Rate Constants for Toluene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0	18	0 ¹¹	16	¹¹ No Redox Regime Given
		0.043 ³¹	15			³¹ Meth/SO ₄ /Fe
	6-8	0.043 ¹²	27	0.016 ¹³	10	¹² Meth/Fe
						¹³ No Redox Regime Given
						¹⁴ Meth/Fe/Mn
		0.093 ¹⁴	8	0.087 ¹⁵	14	¹⁵ Fe
						¹⁶ Meth/Fe/NO ₃
		0 ¹⁶	19	0.0045 ¹⁷	17	¹⁷ No Redox Regime Given
						¹⁸ 1 site; 2 studies one with ¹⁹ Meth/SO ₄ /Fe
		0.0532 ¹⁸ (0.1 ¹⁹ ; 0.0064)	1	0.010 ²⁰	28	²⁰ Fe
		0 ²¹ (0 ²² ; 0)	1	0.083 ²³	1	²¹ 1 site; 2 studies one with ²² Meth/NO ₃
		0.10 ²⁴	2			²³ NO ₃ /SO ₄
						²⁴ Meth/Fe/Mn
		0.042 ³² (0.067; 0.026; 0.053; 0.023)	30	0.0024 ³³ (0.00099; 0.0039; 0.0023)	28	³² 1 site; 4 studies for Meth/NO ₃ /SO ₄
						³³ 1 site; 3 studies for Fe
0.186	27	0.019 ³⁴	31	³⁴ No Redox Regime Given		
0 ³⁵	21	0.066	20	³⁵ Meth/SO ₄ /Fe		
0.042 ³⁷ (0.053-0.067; 0.023-0.026)	4			³⁷ 1 site; 2 studies for Meth/NO ₃ /SO ₄		
> 8	No Studies			No Studies		

Table F.2 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Toluene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	No Studies		0.0086 ³⁸ (0.0052-0.012)	23	³⁸ SO ₄ /Fe
	6-8	0	7	0.11	3	
		0.034 ²⁵ (0.036* ¹ ; 0; 0.032* ² ; 0.06* ³ ; 0.012; 0.063)	11	0.0446 ²⁶	9	²⁵ 1 site; 6 studies; * ¹ 208 days lag; * ² 99 days lag; * ³ 302 days lag
						²⁶ SO ₄ /Fe
		0.0115 ²⁷	9	0.011	22	²⁷ SO ₄ /Fe
		0.028 ²⁸ (0.036; 0.020)	5	0.027 ³⁹ (0.023; 0.031)	25	²⁸ 1 site; 2 studies
		0.0098	4	0.091	6	²⁹ No Redox Regime Given
		0	12	0.018 (0.0075-0.03)	7	³⁹ 1 site; 2 studies
		0.018	28	0.045	24	
		0.003	24			
		0.032 ⁴⁰ (0.05; 0.013)	29			⁴⁰ 1 site; 2 studies
	0	14				
	> 8	No Studies		No Studies		

Table F.3
Summary Table for Anaerobic Biodegradation Rate Constants for Ethylbenzene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments		
		Rate (1/day)	Ref.	Rate (1/day)	Ref.			
# 15	< 6	0 ⁹	2	0	2	⁹ Meth/SO ₄		
	6-8	0.0055 ¹⁰	7	0 ¹¹	9	¹⁰ Meth/Fe/Mn ¹¹ No Redox Regime Given		
		0.0066 ¹ (0.011; 0.0024; 0.0099; 0.0031)	19	0.00187 ² (0.0032; 0.00060; 0.0018)	17	¹ 1 site; 4 studies for Meth/NO ₃ /SO ₄ ² 1 site; 3 studies for Fe		
		0.00675 ³ (0.003-0.011; 0.003-0.010)	5	0.019 ⁶	20	³ 1 site; 2 studies for Meth/NO ₃ /SO ₄		
		0.025 ¹⁵ (0.0011 ^{15a} , 0.028* ^{15a} ; 0 ^{15b} , 0.0053 ^{15c} ; 0.067 ^{15c})	1			¹⁵ 1 site; 4 studies; ^{15a} Meth; ^{15b} Meth/NO ₃ ; ^{15c} Meth/SO ₄ ; * 4 days lag		
		0 ¹² (0 ^{12a} ; 0 ^{12b})	1			¹² 1 site 2 studies; ^{12a} Meth; ^{12b} Meth/NO ₃		
		0 ⁷	13			⁵ Meth/SO ₄ /Fe ⁶ No Redox Regime Given		
		0.024 ⁵	11			⁷ Meth/SO ₄ /Fe ⁵ Meth/SO ₄ /Fe		
		> 8	No Studies				No Studies	

Table F.3 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Ethylbenzene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	No Studies		No Studies		
	6-8	0	10	0 ¹³	4	¹³ SO ₄ /Fe
		0.0031	14	0.012 ⁴ (0; 0.029; 0.009; 0.010)	14, 15	⁴ 1 site; 4 studies
		0.04 ⁸ (0.03; 0.05)	18			⁸ 1 site; 2 studies
		0.0029	5	0.0019 ¹⁴	8	¹⁴ SO ₄ /Fe
		0.0076*	16			*140 days lag
		.29 ¹⁵ (0.12; 0.46)	6			¹⁵ 1 site; 2 studies
	> 8	No Studies		No Studies		

Table F.4
Summary Table for Anaerobic Biodegradation Rate Constants for m-Xylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.057 ¹	8	0 ¹¹	2	¹ Meth/SO ₄ /Fe
		0 ¹⁰	2			¹⁰ Meth/SO ₄
						¹¹ Fe
	6-8	0.0077 ² (0.0037; 0.014; 0.0083; 0.0046)	13	0.00203 ³ (0.0033; 0.0012; 0.0016)	14	² 1 site; 4 studies for Meth/NO ₃ /SO ₄
						³ 1 site; 3 studies for Fe
		0	1	0.019 ⁵	15	⁴ 1 site; 3 studies; ^{4a} Meth/SO ₄
		0.121 ⁴ (0; 0.32 ^{4a} ; 0.044)	1			⁵ No Redox Regime Given
		0 ⁶	10			⁶ Meth/SO ₄ /Fe
	0.019 ⁷ (0.004-0.008; 0.005-0.014)	5		⁷ 1 site; 2 studies for Meth/NO ₃ /SO ₄		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.003	11	0.024	11	
		0	7	0.0204 ¹²	6	¹² SO ₄ /Fe
		0.0006	5	0.17*	3	*17 days lag
		0.06 ⁸ (0.02; 0.1)	12	0.050	9	⁸ 1 site; 2 studies
	0.0143 ⁹			6	⁹ SO ₄ /Fe; 121 days lag	
> 8	No Studies		No Studies			

Table F.5
Summary Table for Anaerobic Biodegradation Rate Constants for o-Xylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments	
		Rate (1/day)	Ref.	Rate (1/day)	Ref.		
# 15	< 6	0.029 ⁶	11	0 ¹	12	¹ - No redox regime given	
				0 ⁷	2	⁶ - Meth/SO ₄ /Fe	
						⁷ - Fe reducing	
	6-8	0.0043	19	0.063 ²	23	² - No redox regime given	
				0 ⁶	4	⁶ - No redox regime given	
		0	13	0.0015 ³ (0.0022; 0.0015; 0.0082)	20	³ - Fe reducing	
		0.00795 ⁴ (0.004-0.011; 0.009-0.016; 0.004; 0.015; 0.0011; 0.0086)	6, 22			⁴ - 1 site 6 studies for Meth/NO ₃ /SO ₄	
		0 ⁵	16			⁵ - Meth/SO ₄ /Fe	
		0	1				
		0.071 ⁸	19			⁸ - Meth/Fe	
		0.0091 ⁹	8			⁹ -Meth/Fe/Mn	
		0 ¹⁰	13			¹⁰ -Meth/Fe/NO ₃	
> 8	No Studies		No Studies				

Table F.5 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for o-Xylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
	6-8	0.21	21	0.16*	7	* - 17 days lag
		0.003	17	0.077*	15	* - 17 days lag
		0.0006	6	0.02	17	
		0	10	0	9	* ¹¹ - SO ₄ /Fe; 37 days lag
		0.0087*	18	0.00559* ¹¹	5	* - 140 days lag
		0		3		
	> 8	No Studies		No Studies		

Table F.6
Summary Table for Anaerobic Biodegradation Rate Constants for p-Xylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.057 ¹	5	0 ⁹	1	¹ - Meth/SO ₄ /Fe
						⁹ - Fe
		0 ⁷	1			⁷ - Meth/SO ₄
	6-8	0.0043	12	0.019 ⁴	13	² - Meth/SO ₄ /Fe
		0.057 ⁸	12			⁸ - Meth/Fe
		0.0006	3			
		0 ²	6			⁴ - No redox regime given
		0.007 ³ (0.005-0.014; 0.002-0.010; 0.0094; 0.0024; 0.0051; 0.0096)	3, 10			0.0015 ⁵ (0.00085; 0.0019; 0.0018)
					⁵ - 1 Site 3 studies for Fe	
	> 8	No Studies		No Studies		

Table F.6 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for p-Xylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	No Studies		No Studies		
	6-8	0.05 (0.02;0.08)	10	0.016 ⁶ (0; 0.032)	8	⁶ - 1 Site 2 studies
				0.17*	2	* - 21 days lag
		0.0029	8	0.0143*	4	* - 121 days lag
				0.0053 (0.0023- 0.0083)	7	
	> 8	No Studies		No Studies		

Table F.7
Summary Table for Anaerobic Biodegradation Rate Constants for Carbon Tetrachloride According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.11	1	No Studies		
	6-8	0.32 (0.15-0.49)	2	0.0602 ¹ (0.0037; 0.0037-0.23)	2	¹ 1 site; 2 studies for Fe
		0.21	1			
		0.23	2	0 ³	2	
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		1.73 ²	3	² NO ₃ /SO ₄
	> 8	No Studies		No Studies		

Table F.8
Summary Table for Anaerobic Biodegradation Rate Constants for Chloroform According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.01562 ² (0.025 ^{*a} ; 0.02 ^{*a} ; 0.025 ^{*a} ; 0.004 ^{*b} ; 0.0041 ^{*b})	2	² 1 site; 5 studies with No Redox Regime Given * ^a 56 days lag * ^b 112 days lag
	6-8	No Studies		0.142 ³ (0.099; 0.25 [*] ; 0.033; 0.099; .25 [*] ; .12)	2	³ 1 site; 6 studies with No Redox Regime Given *14 days lag
				0.03	1	¹ No Redox Regime Given *20-30 days lag
	> 8	No Studies		No Studies		

Table F.9
Summary Table for Anaerobic Biodegradation Rate Constants for 1,2-Dichloroethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0076 (0.0042-0.011)	1	No Studies		
	> 8	No Studies		No Studies		

Table F.10
Summary Table for Anaerobic Biodegradation Rate Constants for Dichloromethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0064	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.11
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 1, 2, 2, -Tetrachloroethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.12
Summary Table for Anaerobic Biodegradation Rate Constants for Tetrachloroethylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0	4	No Studies		
	6-8	0.000735 (0.00068-0.00079)	1	0.00405 ¹ (0.0035-0.0046)	5	¹ NO ₃ /Fe/SO ₄ /Meth
		0 ²	8	0 ³	3	² Meth/SO ₄ /Fe
		0.071 ⁶	2			³ Fe/NO ₃ /Mn
		0 ⁷	3			⁶ Meth/SO ₄
		0.0109 ⁴ (0; 0.0097-0.034)	3			⁷ Meth/Fe/NO ₃ /Mn
			⁴ 1 site; 2 studies			
> 8	No Studies		No Studies			
> 15	< 6	No Studies		0.00073	2	
	6-8	0.003 ⁵	7	0.054 ⁸	6	⁵ No Redox Regime Given
		0.0084	2			⁸ No Redox Regime Given
				0.0065 ⁹ (0; 0.013)	9	⁹ 1 site; 2 studies No Redox Regime Given
> 8	No Studies		No Studies			

Table F.13
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,1-Trichloroethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.011 ¹ (0; 0.022)	3	No Studies		¹ 1 site; 2 studies
	6-8	0.0013	1	0 ⁴	4	⁴ Fe/NO ₃
		0.0037	4	0 ⁶	4	⁶ Fe/NO ₃ /Mn
		0 ²	8	0.010 ³	5	² Meth/SO ₄ /Fe
		0.041*	5			³ Fe
		0.0375 (0.029-0.046)	4			*30 days lag
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0.003 ⁵	7	⁵ No Redox Regime Given
		0.0092 ⁷ (0.015; 0.0034)	1	0.0092 ⁸ (0.015; 0.0034)	1	⁷ 1 site; 2 studies
						⁸ 1 site; 2 studies
	0.0099 ⁹		0.0099 ⁹	6	⁹ No Redox Regime Given	
> 8	No Studies		No Studies			

Table F.14
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 1, 2-Trichloroethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.15
Summary Table for Anaerobic Biodegradation Rate Constants for Trichloroethylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0	9	No Studies		
	6-8	0.00062 (0.00045-0.00079)	5	0.0023 ¹ (0.0017-0.0029)	4	¹ SO ₄ /Fe
		0	12	0.0036 ² (0.00014-0.0071)	3	² SO ₄ /Fe
		0.0034 ⁴ (0.0047; 0.00082; 0.0047)	15	³ 0.00086	6	³ Fe/ Meth/SO ₄
						⁴ 1 site; 3 studies for Meth/SO ₄
		0.0023 ⁶ (0.0011; 0.0034; 0.0010; 0.0036; 0.0025)	20, 21	0.00074 ⁵ (0.00019-0.0024; 0.00014-0.00024)	13	⁵ 1 site; 2 studies with No Redox Regime Given
						⁶ 1 site; 5 studies for Meth/SO ₄
		0 ⁷	8	0.0013 ⁸ (0.00082; 0.0033; 0.0014)	16	⁷ Meth/Fe/SO ₄
						⁸ 1 site; 3 studies for NO ₃ /Fe/SO ₄ /Meth
						⁹ 1 site; 3 studies for Fe
		0.014 ⁹ (0.0016; 0.0015; 0.0011)		0.0033 ¹⁰ (0.0038; 0.0033; 0.0027)	21	¹⁰ 1 site; 3 studies for SO ₄ /Fe
	0 ¹⁴	8	0.010 ¹⁵	19	¹⁴ Meth/Fe/NO ₃ /Mn	
¹⁵ Fe						
> 8	No Studies		No Studies			

Table F.15 (continued)
Summary Table for Anaerobic Biodegradation Rate Constants for Trichloroethylene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
> 15	< 6	No Studies		0 ¹⁶ (0; 0; 0; 0)	10	¹⁶ 1 site; 4 studies with No Redox Regime Given
				0.0011 ¹⁷	18	¹⁷ SO ₄ /Fe
	6-8	0.016 ¹⁹ (0.011-0.021)	1	0.00069 ¹¹ (0.00059-0.00079)	14	¹¹ Fe/Meth/SO ₄
				¹² 0.003	11	¹² No Redox Regime Given
				0.0015 ¹³ (0.0026; 0.0005)	2	¹³ 1 site; 2 studies with No Redox Regime Given
				0.00153 ¹⁸ (0.0017; 0; 0.0029)	18	¹⁸ 1 site; 3 studies with SO ₄ /Fe
						¹⁹ Meth/Fe/SO ₄
				0.039 ²⁰ (0.038* ¹ ; 0.040* ²)	7	0.0090 ²¹ (0.0077; 0.0082-0.011)
	0.013* (0.0020-0.024)	17	0.0029 ²² (0.00057-0.005)	4	*112 days lag	
			0.010	19	²² SO ₄ /Fe	
	> 8	No Studies		No Studies		

Table F.16
Summary Table for Anaerobic Biodegradation Rate Constants for Vinyl Chloride According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.00093 (0.00086-0.0010)	2	0.0008 ¹ (0.0012; 0.0013; 0)	3	¹ - 1 site 3 studies for NO ₃ /Fe/SO ₄ /Meth
		0.00297 ² (0.0024; 0.0060; 0.00049)	4			² 1 site; 3 studies for Meth/SO ₄
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0.0069 ³ (0.0082; 0.0057)	1	³ 1 site; 2 studies with No Redox Regime Given
				0.0082 ⁴	1	⁴ - No Redox Regime Given
	> 8	No Studies		No Studies		

Table F.17
Summary Table for Anaerobic Biodegradation Rate Constants for Phenol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		0.20 ²	7	² - No redox regime given
	6-8	0 ⁷ (0,0 ³)	6	0.0135 ¹ (0; 0.027*)	6	¹ - 1 Site 2 studies of Fe, * - 0-70 days lag
				0 ⁴	4	⁴ - No redox regime given
						⁷ 1 site; 2 studies, ³ - Meth/Fe/NO ₃
> 8	No Studies		No Studies			
> 15	< 6	0.13*	3	No Studies		* - 50 days lag
	6-8	0.057 ⁶ (0.032, 0.068*, 0.071**)	2, 1, 2	No Studies		⁶ 1 site; 3 studies, * - 40 days lag, ** - 30 days lag
				No Studies		
	> 8	No Studies		No Studies		

Table F.18
Summary Table for Anaerobic Biodegradation Rate Constants for o-Cresol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ¹ (0; 0)	4	0 ³	3	¹ 1 site; 2 studies for Meth/Fe/NO ₃
	> 8	No Studies		No Studies		³ - No redox regime given
> 15	< 6	No Studies		No Studies		
	6-8	0.0050	5	No Studies		² 1 site; 3 studies * - 100 days lag
		0.033 ² (0.032*; 0.034; 0.034*)	2, 1, 2			
> 8	No Studies		No Studies			

Table F.19
Summary Table for Anaerobic Biodegradation Rate Constants for m-Cresol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.033	2	No Studies		¹ 1 site; 3 studies * - 100 days lag
		0.0203 ¹ (0.0029, 0.029*, 0.029*)	3, 2,1			
	> 8	No Studies		No Studies		

Table F.20
Summary Table for Anaerobic Biodegradation Rate Constants for p-Cresol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.04 (0.035*, 0.048, 0.037*)	1,1,2	No Studies		¹ 1 site; 3 studies * - 100 days lag
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.21
Summary Table for Anaerobic Biodegradation Rate Constants for 2,4-Dichlorophenol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref	Rate (1/day)	Ref	
# 15	< 6	0.015	2	No Studies		
	6-8	0.017	2	0 ¹ (0, 0)	4	¹ 1 site; 2 studies for Fe/NO ₃
		0.027	4			
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.090	5	0.0055	1	
		0.12 ⁴	3	0 ²	5	⁴ - Meth/SO ₄
		0.12	1			² No Redox Regime Given
	> 8	No Studies		No Studies		

Table F.22
Summary Table for Anaerobic Biodegradation Rate Constants for 2,4,6-Trichlorophenol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.23
Summary Table for Anaerobic Biodegradation Rate Constants for Pentachlorophenol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	1.98e-03 ¹ (1.61e-03-2.36e-03)	1	No Studies		¹ - 1 site 2 studies
		1.414e-03 ¹ (1.01e-03-2.17e-03; 6.86e-04-1.79e-03)	1			
	6-8	1.97e-03 ³ (1.60e-03-2.42e-03; 1.06e-03-2.80e-03)	1	No Studies		³ - 1 site 2 studies
	> 8	No Studies		No Studies		
> 15	< 6	4.94e-03 (7.38e-03 -2.50e-03)	1	0 ²	3	² No Redox Regime Given
	6-8	0 (0; 0)	4	No Studies		⁴ - 1 site 2 studies
		5.09e-03 ⁵ (2.91e-03-7.27e-03)	1			⁵ - 1 site 2 studies
		0	2			
		5.75e-03 ⁴ (1.63e-03-2.15e-03; 0.0162-3.0e-03)	1			
	> 8	No Studies		No Studies		

Table F.24
Summary Table for Anaerobic Biodegradation Rate Constants for Trichlorofluoromethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0016	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.25
Summary Table for Anaerobic Biodegradation Rate Constants for Dichlorodifluoromethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref	Rate (1/day)	Ref	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		0	1	¹ No Redox Regime Given
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.26
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1,2-Trichloro-1,2,2-trifluoroethane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref	Rate (1/day)	Ref	
# 15	< 6	No Studies		No Studies		
	6-8	0	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.27
Summary Table for Anaerobic Biodegradation Rate Constants for Acetone According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.28
Summary Table for Anaerobic Biodegradation Rate Constants for Methyl Ethyl Ketone According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.29
Summary Table for Anaerobic Biodegradation Rate Constants for Methyl Isobutyl Ketone According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.30
Summary Table for Anaerobic Biodegradation Rate Constants for Acetic Acid According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.31
Summary Table for Anaerobic Biodegradation Rate Constants for Phenylacetic Acid According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.32
Summary Table for Anaerobic Biodegradation Rate Constants for Acenaphthene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.0043	1	No Studies		
	> 8	No Studies		No Studies		

Table F.33
Summary Table for Anaerobic Biodegradation Rate Constants for Fluorene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0	2	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.00030 ¹	1	¹ - No redox regime given
	6-8	0 ³	3	0.00145 ² (0.0013 - 0.0016)	4	² - No redox regime given
						³ - Meth/NO ₃ /SO ₄
	> 8	No Studies		No Studies		

Table F.34
Summary Table for Anaerobic Biodegradation Rate Constants for 1-Methylnaphthalene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.057 ¹	4	0 ³	1	¹ Meth/SO ₄ /Fe
		0 ²	1			² Meth/SO ₄
						³ Fe
	6-8	0	5	0.00031 ⁴	2	⁴ No Redox Regime Given
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0		0.00054 ⁵ (0.00040-0.00068)		⁵ No Redox Regime Given
	> 8	No Studies		No Studies		

Table F.35
Summary Table for Anaerobic Biodegradation Rate Constants for Naphthalene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0.026 ¹	7	0 ¹⁰	2	¹ Meth/SO ₄ /Fe
		0 ⁹	2			⁹ Meth/SO ₄
		0	11			¹⁰ Fe
	6-8	0 ⁵	10	0.0063 ⁴	15	⁴ No Redox Regime Given
			10			⁵ Meth/Fe/SO ₃
		0 ¹¹	10	0 ¹²	1	¹¹ Meth/Fe/NO ₃
		0 ¹³	3	0 ¹⁴	6	¹² No Redox Regime Given
						¹³ Meth/Fe/Mn
						¹⁴ No Redox Regime Given
		0	12	0 ⁶	8	⁶ No Redox Regime Given
	0.0018 ⁷ (0.0015-0.0021)			14	⁷ No Redox Regime Given	
	0.0050 ⁸	12	⁸ Fe			
	> 8	No Studies	No Studies	No Studies	No Studies	No Studies
> 15	< 6	0	9	0.03 ² (0.017-0.043)	13	² SO ₄ /Fe
				0.00018 ³	4	³ No Redox Regime Given
				0	8	
	6-8	0	5	No Studies	No Studies	No Studies
	> 8	No Studies	No Studies	No Studies	No Studies	No Studies

Table F.36
Summary Table for Anaerobic Biodegradation Rate Constants for Phenanthrene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.37
Summary Table for Anaerobic Biodegradation Rate Constants for 1,1' Biphenyl According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref	Rate (1/day)	Ref	
# 15	< 6	No Studies		No Studies		
	6-8	0 ³ (0, 0)	2	0.019 ¹	3	¹ - No redox regime given
	> 8	No Studies		No Studies		³ 1 site; 2 studies of Meth/Fe/NO ₃
> 15	< 6	No Studies		0.00032 ²	1	² - No redox regime given
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.38
Summary Table for Anaerobic Biodegradation Rate Constants for Cumene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0 ¹	1	0 ²	1	¹ Meth/SO ₄
	> 8	No Studies		No Studies		² No Redox Regime Given
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.39
Summary Table for Anaerobic Biodegradation Rate Constants for Dioxane According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0	1	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.40
Summary Table for Anaerobic Biodegradation Rate Constants for Methanol According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0067	5	No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		0.034 ¹ (0.043; 0.0022; 0.0089; 0.0039; 0.072; 0.072)	4	¹ - 1 Site 6 studies; No redox regime given
				0.034 (0.028; 0.030; 0.054; 0.036; 0.016; 0.029; 0.039; 0.039)	1, 6, 7	⁴ 1 site; 8 studies *-No redox regime given
	6-8	0.089 ⁵	3	0.071 ²	1	² - No redox regime given
				0.067 ³	1	³ - No redox regime given
				0.34	3	⁵ Meth/SO ₄
				0.058 (0.1*; 0.016**)	4	* - 24 days lag
						** - 30 days lag
	> 8	No Studies		No Studies		

Table F.41
Summary Table for Anaerobic Biodegradation Rate Constants for Nitrobenzene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	0.0037 ² (0.0037; 0.0037)	1	0.00263 ¹ (0.0037 ^{1a} ; 0.0037 ^{1a} ; 0.0037 ^{1b} ; 0.037- 0.23 ^{1b})	1	¹ - 1 site 5 studies ^{1a} - NO ₃ /Mn ^{1b} - Fe reducing
	> 8	No Studies		No Studies		² - 1 site 2 studies
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		

Table F.42
Summary Table for Anaerobic Biodegradation Rate Constants for Pyridine According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref	Rate (1/day)	Ref	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0.010 ¹ (0.020*, 0)	2, 1	0.014**	2	¹ 1 site; 2 studies * - 90 days lag
	> 8	No Studies		No Studies		** - 30 days lag

Table F.43
Summary Table for Anaerobic Biodegradation Rate Constants for Styrene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	No Studies		No Studies		
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	No Studies		0 ¹	1	¹ No Redox Regime Given
	> 8	No Studies		No Studies		

Table F.44
Summary Table for Anaerobic Biodegradation Rate Constants for 1, 3, 5-Trimethylbenzene According to EPA Protocol
(Field and Laboratory Studies)

Temp. (°C)	pH	Methanogenic		Sulfate Reducing		Comments
		Rate (1/day)	Ref.	Rate (1/day)	Ref.	
# 15	< 6	0 ¹	1	0 ²	1	¹ Meth/SO ₄
						² Fe
	6-8	No Studies		No Studies		
	> 8	No Studies		No Studies		
> 15	< 6	No Studies		No Studies		
	6-8	0	4	0	3	
				0.0039	2	
	> 8	No Studies		No Studies		

**APPENDIX G
REFERENCES FOR ALL COMPOUNDS**

Draft

REFERENCES FOR BENZENE

1. Acton, D.W. & Barker, J.F., *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters: *Journal of Contaminant Hydrology*, v. 9, p. 325-52 (1992)
2. Baedecker, M.J., et al. Crude oil in a shallow sand and gravel aquifer-M. Biogeochemical reactions and mass balance modeling in anoxic groundwater. *Applied Geochem* 8: 569-586 (1993)
3. Ball, H.A. & Reinhard, M. Monoaromatic hydrocarbon transformation under anaerobic conditions at Seal Beach, California: laboratory studies. *Environ Toxicol Chem* 15: 114-22 (1996)
4. Barker, J.F., et al. Natural attenuation of aromatic hydrocarbons in a shallow sand aquifer. *Ground Water Monit Rev* 7: 64-72 (1987)
5. Barlaz, M.A., et al. Intrinsic bioremediation of a gasoline plume: comparisons of field and laboratory results. In: *Bioremediation of Hazardous Wastes. Research, Development, and Field Evaluations*. USEPA. EPA/540/R-95-532 (1995)
6. Barlaz, M.A., et al. Rate and extent of natural anaerobic bioremediation of BTEX compounds in ground water plumes. In: *Symposium on Bioremediation of Hazardous Wastes: Research, Development, and Field Evaluations*; Dallas, TX US EPA. EPA/600/R-93/054 (1993)
7. Chapelle, F.H. Identifying redox conditions that favor the natural attenuation of chlorinated ethenes in contaminated ground-water systems. In: *Symposium on Natural Attenuation of Chlorinated Organics in Ground Water*. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX. September 11 - 13 (1996)
8. Cozzarelli, I.M., et al. The geochemical evolution of low-molecular-weight organic acids derived from the degradation of petroleum contaminants in groundwater. *Geochim Cosmochim Acta* 58: 863-877 (1994)
9. Cozzarelli, I.M., et al. Transformation of monoaromatic hydrocarbons to organic acids in anoxic groundwater environment. *Environ Geol Water Sci* 16: 135-141 (1990)
10. Davis, J.W., et al. Natural Biological Attenuation of Benzene in Ground Water Beneath a Manufacturing Facility. *Ground Water* 32: 215-226 (1994)
11. Hunt, M.J., et al. Anaerobic BTEX biodegradation in laboratory microcosms and in situ columns. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 101-107 (1995)
12. Kao, C.M. & Borden, R.C. Site specific variability in biodegradation under denitrifying conditions. *Ground Water* 35(2): 305-311 (1997)
13. Kazumi, J., et al. Anaerobic degradation of benzene in diverse anoxic environments. *Environ Sci Technol* 31: 813-18 (1997)
14. Lovley, D.R., et al. Rapid anaerobic benzene oxidation with a variety of chelated Fe(III) forms. *Appl Environ Microbiol* 62: 288-291 (1996)
15. Lyngkilde, J., et al. Degradation of specific organic compounds in leachate-polluted groundwater. In: Landfilling, Waste: Leachate. Christensen, T.H., et al. (eds.). Elsevier: London, UK pp. 485-95 (1992)

16. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
17. Major, D.W., et al. Biotransformation of benzene by denitrification in aquifer sand. *Ground Water* 26: 8-14 (1988)
18. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
19. Nielsen, P.H. & Christensen, T.H. In situ measurement of degradation of specific organic compounds under aerobic, denitrifying, iron(RD-reducing, and methanogenic groundwater conditions. In: Bioremediation of Chlorinated and PAH Compounds. Hinchee, R.E. (ed.). Lewis pp 416-422 (1994)
20. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449462 (1992)
21. Reinhard, M., et al. Occurrence and distribution of organic chemicals in two landfill leachate plumes. *Environ Sci Technol* 18: 953-961 (1984)
22. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
23. Thierrin, J., et al. A ground-water tracer test with deuterated compounds for monitoring in situ biodegradation and retardation of aromatic hydrocarbons. *Ground Water* 33: 469-475 (1995)
24. Wiedemeler, T.H., et al. Approximation of biodegradation rate constants for monoaromatic hydrocarbons (BTEX) in ground water. *Ground Water Monit Remediat.* 16: 186-194 (1996)
25. Wiedemeier, T.H., et al. Patterns of intrinsic bioremediation at two US Air Force Bases. In: Intrinsic Bioremediation. Hinchee, RE et al. (eds.). Battelle Press: Columbus, OH. pp. 31-51 (1995)
26. Wilson, B.H., et al. Biotransformation of selected alkylbenzenes and halogenated aliphatic hydrocarbons in methanogenic aquifer material: a microcosm study. *Environ Sci Technol* 20: 997-1002 (1986)
27. Wilson, B.H., et al. Biotransformation of monoaromatic and chlorinated hydrocarbons at an aviation gasoline spill site. *Geomicrobiol J* 8: 225-40 (1990)
28. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: *Symposium on Natural Attenuation of Chlorinated Organics in Ground Water*. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)
29. Wilson, J.T., et al. Intrinsic bioremediation of JP-4 jet fuel. In: *Symposium on Intrinsic Bioremediation of Ground Water*. Denver, CO. EPA/540/R-94/515. Washington, DC: USEPA (1994A)

30. Wilson, J.T., et al. Natural bioreclamation of alkylbenzenes (BTEX) from a gasoline spill in methanogenic groundwater. In: Hydrocarbon Bioremediation. Hinchee, R.E., et al. (eds.). Lewis Publishers: Boca Raton, FL. pp. 201-218 (1994B)
31. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR TOLUENE

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)
2. Baedecker, M.J., et al. Crude oil in a shallow sand and gravel aquifer-M. Biogeochemical reactions and mass balance modeling in anoxic groundwater. *Applied Geochem* 8: 569-586 (1993)
3. Ball, H.A. & Reinhard, M. Monoaromatic hydrocarbon transformation under anaerobic conditions at Seal Beach, California: laboratory studies. *Environ Toxicol Chem* 15: 114-22 (1996)
4. Barlaz, M.A., et al. Rate and extent of natural anaerobic bioremediation of BTEX compounds in ground water plumes. In: *Symposium on Bioremediation of Hazardous Wastes: Research, Development, and Field Evaluations*; Dallas, TX US EPA. EPA/600/R-93/054 (1993)
5. Beller, H.R., et al. Byproducts of anaerobic alkylbenzene metabolism useful as indicators of in situ bioremediation. *Environ Sci Technol* 29: 2864-2870 (1995)
6. Beller, H.R., et al. Microbial degradation of alkylbenzenes under sulfate-reducing and methanogenic conditions. Robert S. Kerr Environmental Research Laboratory. US EPA Report EPA/600/S2-91/027. Robert S Kerr Environmental Research Laboratory. Ada, OK (1991)
7. Chapelle, F.H. Identifying redox conditions that favor the natural attenuation of chlorinated ethenes in contaminated ground-water systems. In: *Symposium on Natural Attenuation of Chlorinated Organics in Ground Water*. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX. September 11 - 13 (1996)
8. Cozzarelli, I.M., et al. The geochemical evolution of low-molecular-weight organic acids derived from the degradation of petroleum contaminants in groundwater. *Geochim Cosmochim Acta* 58: 863-877 (1994)
9. Hunt, M.J., et al. Anaerobic BTEX biodegradation in laboratory microcosms and in situ columns. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 101-107 (1995)
10. Hutchins, S.R. Biodegradation of monoaromatic hydrocarbons by aquifer microorganisms using oxygen, nitrate, or nitrous oxide as the terminal electron acceptor. *Appl Environ Microbiol* 57: 2403-2407 (1991)
11. Johnston, J.J., et al. Anaerobic biodegradation of alkylbenzenes and trichloroethylene in aquifer sediment down gradient of a sanitary landfill. *J Contam Hydrol* 23: 263-283 (1996)
12. Kao, C.M. & Borden, R.C. Site specific variability in biodegradation under denitrifying conditions. *Ground Water* 35(2): 305-311 (1997)

13. Lovley, D.R., et al. Simulated anoxic biodegradation of aromatic hydrocarbons using Fe(II) ligands. *Nature* 370: 128-131 (1994)
14. Lovley, D.R., et al. Oxidation of aromatic contaminants coupled to microbial iron reduction. *Nature* 339: 297-299 (1989)
15. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
16. Lyngkilde, J., et al. Degradation of specific organic compounds in leachate-polluted groundwater. In: Landfilling, Waste: Leachate. Christensen, T.H., et al. (eds.). Elsevier: London, UK pp. 485-95 (1992)
17. Major, D.W., et al. Biotransformation of benzene by denitrification in aquifer sand. *Ground Water* 26: 8-14 (1988)
18. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
19. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449462 (1992)
20. Reinhard, M., et al. In situ BTEX biotransformation under intrinsic and nitrate- and sulfatereducing conditions. American Chemical Society. Division of Environmental Chemistry Preprints of Extended Abstracts, 211th ACS National Meeting. 36: 210-212 (1996)
21. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
22. Sewell, G.W. & Gibson, S.A. Stimulation of the reductive dechlorination of tetrachloroethene in anaerobic aquifer microcosms by the addition of toluene. *Environ Sci Technol* 25: 982-984 (1991)
23. Thierrin, J., et al. A ground-water tracer test with deuterated compounds for monitoring in situ biodegradation and retardation of aromatic hydrocarbons. *Ground Water* 33: 469-475 (1995)
24. Wiedemeier, T.H., et al. Patterns of intrinsic bioremediation at two US Air Force Bases. In: Intrinsic Bioremediation. Hinchee, R.E., et al. (eds.). Battelle Press: Columbus, OH. pp. 31-51 (1995)
25. Wiedemeier, T.H., et al. Approximation of biodegradation rate constants for monoaromatic hydrocarbons (BTEX) in ground water. *Ground Water Monit Remediat.* 16: 186-194 (1996)
26. Wilson, B.H., et al. Biotransformation of monoaromatic and chlorinated hydrocarbons at an aviation gasoline spill site. *Geomicrobiol J* 8: 225-40 (1990)
27. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)

28. Wilson, B.H., et al. Biotransformation of selected alkylbenzenes and halogenated aliphatic hydrocarbons in methanogenic aquifer material: a microcosm study. *Environ Sci Technol* 20: 997-1002 (1986)
29. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR ETHYLBENZENE

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)
2. Albrechtsen, H.J., et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation, Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hincsee, R.E., et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1993)
3. Ball, H.A. & Reinhard, M. Monoaromatic hydrocarbon transformation under anaerobic conditions at Seal Beach, California: laboratory studies. *Environ Toxicol Chem* 15: 114-22 (1996)
4. Barlaz, M.A., et al. Intrinsic bioremediation of a gasoline plume: comparisons of field and laboratory results. In: *Bioremediation of Hazardous Wastes. Research, Development, and Field Evaluations*. USEPA. EPA/540/R-95-532 (1995)
5. Barlaz, M.A., et al. Rate and extent of natural anaerobic bioremediation of BTEX compounds in ground water plumes. In: *Symposium on Bioremediation of Hazardous Wastes: Research, Development, and Field Evaluations*; Dallas, TX US EPA. EPA/600/R-93/054 (1993)
6. Beller, H.R., et al. Microbial degradation of alkylbenzenes under sulfate-reducing and methanogenic conditions. Robert S. Kerr Environmental Research Laboratory. US EPA Report EPA/600/S2-91/027. Robert S Kerr Environmental Research Laboratory. Ada, OK (1991)
7. Cozzarelli, I.M., et al. The geochemical evolution of low-molecular-weight organic acids derived from the degradation of petroleum contaminants in groundwater. *Geochim Cosmochim Acta* 58: 863-877 (1994)
8. Hunt, M.J., et al. Anaerobic BTEX biodegradation in laboratory microcosms and in situ columns. In: Intrinsic Bioremediation. Hincsee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 101-107 (1995)
9. Hutchins, S.R. Biodegradation of monoaromatic hydrocarbons by aquifer microorganisms using oxygen, nitrate, or nitrous oxide as the terminal electron acceptor. *Appl Environ Microbiol* 57: 2403-2407 (1991)
10. Kao, C.M. & Borden, R.C. Site specific variability in biodegradation under denitrifying conditions. *Ground Water* 35(2): 305-311 (1997)
11. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
12. Reinhard, M., et al. Occurrence and distribution of organic chemicals in two landfill leachate plumes. *Environ Sci Technol* 18: 953-961 (1984)

13. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
14. Wiedemeier, T.H., et al. Patterns of intrinsic bioremediation at two US Air Force Bases. In: Intrinsic Bioremediation. Hinchee, RE et al. (eds.). Battelle Press; Columbus, OH. pp. 31-51 (1995)
15. Wiedemeler, T.H., et al. Approximation of biodegradation rate constants for monoaromatic hydrocarbons (BTEX) in ground water. *Ground Water Monit Remediat.* 16: 186-194 (1996)
16. Wilson, B.H., et al. Biotransformation of selected alkylbenzenes and halogenated aliphatic hydrocarbons in methanogenic aquifer material: a microcosm study. *Environ Sci Technol* 20: 997-1002 (1986)
17. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)
18. Wilson, J.T., et al. Intrinsic bioremediation of JP-4 jet fuel. In: Symposium on Intrinsic Bioremediation of Ground Water. Denver, CO. EPA/540/R-94/515. Washington, DC: USEPA (1994A)
19. Wilson, J.T., et al. Natural bioreclamation of alkylbenzenes (BTEX) from a gasoline spill in methanogenic groundwater. In: Hydrocarbon Bioremediation. Hinchee, R.E., et al. (eds.). Lewis Publishers: Boca Raton, FL. pp. 201-218 (1994B)
20. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR m-XYLENE

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)
2. Albrechtsen, H.J., et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation. Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hinchee, RE et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1993)
3. Ball, H.A. & Reinhard, M. Monoaromatic hydrocarbon transformation under anaerobic conditions at Seal Beach, California: laboratory studies. *Environ Toxicol Chem* 15: 114-22 (1996)
4. Barlaz, M.A., et al. Intrinsic bioremediation of a gasoline plume: comparisons of field and laboratory results. In: Bioremediation of Hazardous Wastes. Research, Development, and Field Evaluations. USEPA. EPA/540/R-95-532 (1995)
5. Barlaz, M.A., et al. Rate and extent of natural anaerobic bioremediation of BTEX compounds in ground water plumes. In: Symposium on Bioremediation of Hazardous Wastes: Research, Development, and Field Evaluations; Dallas, TX US EPA. EPA/600/R-93/054 (1993)
6. Hunt, M.J., et al. Anaerobic BTEX biodegradation in laboratory microcosms and in situ columns. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 101-107 (1995)

7. Kao, C.M. & Borden, R.C. Site specific variability in biodegradation under denitrifying conditions. *Ground Water* 35(2): 305-311 (1997)
8. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
9. Reinhard, M., et al. In situ BTEX biotransformation under intrinsic and nitrate- and sulfatereducing conditions. American Chemical Society. Division of Environmental Chemistry Preprints of Extended Abstracts, 211th ACS National Meeting. 36: 210-212 (1996)
10. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
11. Wiedemeier, T.H., et al. Patterns of intrinsic bioremediation at two US Air Force Bases. In: Intrinsic Bioremediation. Hinchee, R.E., et al. (eds.). Battelle Press: Columbus, OH. pp. 31-51 (1995)
12. Wilson, J.T., et al. Intrinsic bioremediation of JP-4 jet fuel. In: Symposium on Intrinsic Bioremediation of Ground Water. Denver, CO. EPA/540/R-94/515. Washington, DC: USEPA (1994A)
13. Wilson, J.T., et al. Natural bioreclamation of alkylbenzenes (BTEX) from a gasoline spill in methanogenic groundwater. In: Hydrocarbon Bioremediation. Hinchee, R.E., et al. (eds.). Lewis Publishers: Boca Raton, FL. pp. 201-218 (1994B)
14. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)
15. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR o-XYLENE

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)
2. Albrechtsen, H. J. et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation. Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hinchee, RE et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1994)
3. Ball, H.A. & Reinhard, M. Monoaromatic hydrocarbon transformation under anaerobic conditions at Seal Beach, California: laboratory studies. *Environ Toxicol Chem* 15: 114-22 (1996)
4. Barker, J.F., et al. Natural attenuation of aromatic hydrocarbons in a shallow sand aquifer. *Ground Water Monit Rev* 7: 64-72 (1987)

5. Barlaz, M.A., et al. Intrinsic bioremediation of a gasoline plume: comparisons of field and laboratory results. In: *Bioremediation of Hazardous Wastes. Research, Development, and Field Evaluations*. USEPA. EPA/540/R-95-532 (1995)
6. Barlaz, M.A., et al. Rate and extent of natural anaerobic bioremediation of BTEX compounds in ground water plumes. In: *Symposium on Bioremediation of Hazardous Wastes: Research, Development, and Field Evaluations*; Dallas, TX US EPA. EPA/600/R-93/054 (1993)
7. Beller, H.R., et al. Byproducts of anaerobic alkylbenzene metabolism useful as indicators of in situ bioremediation. *Environ Sci Technol* 29: 2864-2870 (1995)
8. Cozzarelli, I.M., et al. The geochemical evolution of low-molecular-weight organic acids derived from the degradation of petroleum contaminants in groundwater. *Geochim Cosmochim Acta* 58: 863-877 (1994)
9. Hunt, M.J., et al. Anaerobic BTEX biodegradation in laboratory microcosms and in situ columns. In: Intrinsic Bioremediation, Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 101-107 (1995)
10. Hutchins, S.R. Biodegradation of monoaromatic hydrocarbons by aquifer microorganisms using oxygen, nitrate, or nitrous oxide as the terminal electron acceptor. *Appl Environ Microbiol* 57: 2403-2407 (1991)
11. Kao, C.M. & Borden, R.C. Site specific variability in biodegradation under denitrifying conditions. *Ground Water* 35(2): 305-311 (1997)
12. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
13. Lyngkilde, J., et al. Degradation of specific organic compounds in leachate-polluted groundwater. In: Landfilling. Waste. Leachate, Christensen, T.H., et al. (eds.). Elsevier: London, UK pp. 485-95 (1992)
14. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
15. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449-462 (1992)
16. Reinhard, M., et al. In situ BTEX biotransformation under intrinsic and nitrate- and sulfatereducing conditions. American Chemical Society. Division of Environmental Chemistry Preprints of Extended Abstracts, 211th ACS National Meeting. 36: 210-212 (1996)
17. Rugge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation, Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
18. Wiedemeier, T.H., et al. Patterns of intrinsic bioremediation at two US Air Force Bases. In: Intrinsic Bioremediation, Hinchee, R.E., et al. (eds.). Battelle Press: Columbus, OH. pp. 31-51 (1995)

19. Wilson, B.H., et al. Biotransformation of selected alkylbenzenes and halogenated aliphatic hydrocarbons in methanogenic aquifer material: a microcosm study. *Environ Sci Technol* 20: 997-1002 (1986)
20. Wilson, B.H., et al. Biotransformation of monoaromatic and chlorinated hydrocarbons at an aviation gasoline spill site. *Geomicrobiol J* 8: 225-40 (1990)
21. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: *Symposium on Natural Attenuation of Chlorinated Organics in Ground Water*. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)
22. Wilson, J.T., et al. Intrinsic bioremediation of JP-4 jet fuel. In: *Symposium on Intrinsic Bioremediation of Ground Water*. Denver, CO. EPA/540/R-94/515. Washington, DC: USEPA (1994A)
23. Wilson, J.T., et al. Natural bioreclamation of alkylbenzenes (BTEX) from a gasoline spill in methanogenic groundwater. In: Hydrocarbon Bioremediation, Hinchee, R.E., et al. (eds.). Lewis Publishers: Boca Raton, FL. pp. 201-218 (1994B)
24. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR p-XYLENE

1. Albrechtsen, H.J., et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation, Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hinchee, R.E., et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1993)
2. Ball, H.A. & Reinhard, M. Monoaromatic hydrocarbon transformation under anaerobic conditions at Seal Beach, California: laboratory studies. *Environ Toxicol Chem* 15: 114-22 (1996)
3. Barlaz, M.A., et al. Rate and extent of natural anaerobic bioremediation of BTEX compounds in ground water plumes. In: *Symposium on Bioremediation of Hazardous Wastes: Research, Development, and Field Evaluations*; Dallas, TX US EPA. EPA/600/R-93/054 (1993)
4. Hunt, M.J., et al. Anaerobic BTEX biodegradation in laboratory microcosms and in situ columns. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 101-107 (1995)
5. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contarn Hydrol* 10: 291-307 (1992)
6. Rugge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
7. Thierrin, J., et al. A ground-water tracer test with deuterated compounds for monitoring in situ biodegradation and retardation of aromatic hydrocarbons. *Ground Water* 33: 469-475 (1995)

8. Wiedemeier, T.H., et al. Patterns of intrinsic bioremediation at two US Air Force Bases. In: Intrinsic Bioremediation. Hinchee, R.E., et al. (eds.). Battelle Press: Columbus, OH. pp. 31-51 (1995)
9. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)
10. Wilson, J.T., et al. Intrinsic bioremediation of JP-4 jet fuel. In: Symposium on Intrinsic Bioremediation of Ground Water. Denver, CO. EPA/540/R-94/515. Washington, DC: USEPA (1994A)
11. Wilson, J.T., et al. Natural bioreclamation of alkylbenzenes (BTEX) from a gasoline spill in methanogenic groundwater. In: Hydrocarbon Bioremediation. Hinchee, R.E., et al. (eds.). Lewis Publishers: Boca Raton, FL. pp. 201-218 (1994B)
12. Wilson, B.H., et al. Biotransformation of monoaromatic and chlorinated hydrocarbons at an aviation gasoline spill site. *Geomicrobiol J* 8: 225-40 (1990)
13. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR CARBON TETRACHLORIDE

1. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449462 (1992)
2. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
3. Semprini, L., et al. In-situ transformation of carbon tetrachloride and other halogenated compounds resulting from biostimulation under anoxic conditions. *Environ Sci Technol* 26: 2454-2461 (1992)

REFERENCES FOR CHLOROFORM

1. Roberts, P.V., et al. Field study of organic water quality changes during groundwater recharge in the Palo Alto baylands. *Water Res* 16: 1025-1035 (1982)
2. Saunders, F., et al. Results of laboratory microcosm studies of the anaerobic biodegradation of chloroform in subsurface environments. NCASI Technical Bulletin No. 716. Research Triangle Park, NC. (1996)

REFERENCES FOR 1, 2-DICHLOROETHANE

1. Lee, M. D. et al. Intrinsic bioremediation of 1,2-dichloroethane. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11 - 13. USEPA. EPA/540/R-96/509 p. 159 (1996)

REFERENCE FOR DICHLOROMETHANE

1. Fiorenza, S., et al. Natural anaerobic degradation of chlorinated solvents at a Canadian manufacturing plant. In: Bioremediation of Chlorinated Polycyclic Aromatic Hydrocarbons, Hinchee, R.E., et al. (eds.) Boca Raton, FL: Lewis Publishers. pp. 277-286 (1994)
2. Lehmicke, L.L., et al. Involvement of dichloromethane in the intrinsic biodegradation of chlorinated ethenes and ethanes. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11 - 13. USEPA. EPA/540/R-96/509 p. 158 (1996)

REFERENCES FOR TETRACHLOROETHYLENE

1. Ellis, D.E., et al. Remediation technology forum intrinsic remediation project at Dover Air Force Base, Delaware. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Dallas, TX, September 11 - 13. USEPA. EPA/540/R-96/509 (1996)
2. Major, D.W., et al. Field and laboratory evidence of in situ biotransformation of tetrachloroethene to ethene and ethane at a chemical transfer facility in North Toronto. In: on-Site Bioreclamation, Hinchee, R.E. & Olfenbuttel, R.F. (eds): Stoneham, MA (1991)
3. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. J Contam Hydrol 20: 51-66 (1995B)
4. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. Chemosphere 25: 449462 (1992)
5. Parsons, F., et al. Transformations of tetrachloroethene and trichloroethene in microcosms and groundwater. J AWWA 76: 56-59 (1984)
6. Roberts, P.V., et al. Field study of organic water quality changes during groundwater recharge in the Palo Alto baylands. Water Res 16: 1025-1035 (1982)
7. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation, Hinchee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
8. Sewell, G.W. & Gibson, S.A. Stimulation of the reductive dechlorination of tetrachloroethene in anaerobic aquifer microcosms by the addition of toluene. Environ Sci Technol 25: 982-984 (1991)
9. Suflita, J.M., et al. Anaerobic biotransformations of pollutant chemicals in aquifers. J Ind Microbiol 3: 179-194 (1988)

REFERENCES FOR 1, 1, 1-TRICHLOROETHANE

1. Fiorenza, S., et al. Natural anaerobic degradation of chlorinated solvents at a Canadian manufacturing plant. In: Bioremediation of Chlorinated Polycyclic Aromatic Hydrocarbons, Hinchee, R.E. et al. (eds.) Boca Raton, FL: Lewis Publishers. pp. 277-286 (1994)
2. Klecka, G.M., et al. Biological transformations of 1,1,1-trichloroethane in subsurface soils and ground water. Environ Toxicol Chem 9: 1437-1451 (1990)

3. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449-462 (1992)
4. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
5. Nielsen, P.H. & Christensen, T.H. In situ measurement of degradation of specific organic compounds under aerobic, denitrifying, iron(RD-reducing, and methanogenic groundwater conditions. In: Bioremediation of Chlorinated and PAH Compounds. Hinchee, R.E. (ed.). Lewis pp 416-422 (1994)
6. Parsons, F., et al. Biotransformation of chlorinated organic solvents in static microcosms. *Environ Toxicol Chem* 4: 739-742 (1985)
7. Roberts, P.V., et al. Field study of organic water quality changes during groundwater recharge in the Palo Alto baylands. *Water Res* 16: 1025-1035 (1982)
8. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hinchee, RE et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)

REFERENCES FOR TRICHLOROETHYLENE

1. Barrio-Lage, G., et al. Kinetics of the depletion of trichloroethene. *Environ Sci Technol* 21(4): 366-370 (1987)
2. Dupont, R.R., et al. Case study: Eielson Air Force Base, Alaska. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX. September 11-13 (1996)
3. Ehlke, T. A. et al. In situ biotransformation of trichloroethylene and cis-1,2-dichloroethylene at Picatinny Arsenal, New Jersey. In: Proceedings of the USGS Toxic Substances Hydrology Program, Colorado Springs, CO. Sept 20-24, 1993. Water Resources Investigations Report 944015 pp. 347-354 (1994)
4. Ehlke, T.A. & Imbrigiotta, T.E. Estimation of laboratory and in situ degradation rates for trichloroethene and cis- 1,2-dichloroethene in a contaminated aquifer at Picatinny Arsenal, New Jersey. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 141-142 (1996)
5. Ellis, D.E., et al. Remediation technology forum intrinsic remediation project at Dover Air Force Base, Delaware. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Dallas, TX, September 11 - 13. USEPA. EPA/540/R-96/509 (1996)
6. Imbrigiotta, T.E., et al. Case study: natural attenuation of a trichloroethene plume at Picatinny Arsenal, New Jersey. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX. September 11-13 (1996)

7. Johnston, J.J., et al. Anaerobic biodegradation of alkylbenzenes and trichloroethylene in aquifer sediment down gradient of a sanitary landfill. *J Contam Hydrol* 23: 263-283 (1996)
8. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
9. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449462 (1992)
10. Odom, J.M., et al. Anaerobic biodegradation of chlorinated solvents: comparative laboratory study of aquifer microcosms. In: Bioremediation of Chlorinated Solvents. Pap Int In Situ On-Site Bioreclamation Symposium. 3rd. Hincsee, R.E. (ed.). Battelle Press: Columbus, OH. pp. 17-24 (1995)
11. Roberts, P.V., et al. Field study of organic water quality changes during groundwater recharge in the Palo Alto baylands. *Water Res* 16: 1025-1035 (1982)
12. Ruge, K., et al. Natural attenuation of xenobiotic compounds: anaerobic field injection experiment. In: Intrinsic Bioremediation. Hincsee, R.E., et al (eds.). Battelle Press; Columbus, OH pp. 127-133 (1995)
13. Silka, L.R. & Wallen, D.A. Observed rates of biotransformation of chlorinated aliphatics in groundwater. In: Superfund'88 Proceedings 9th National Conference. Published by Hazardous Material Control Research Institute 138-141 (1988)
14. Swanson, M., et al. Patterns of natural attenuation of chlorinated aliphatic hydrocarbons at Cape Canaveral Air Station, Florida. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11 - 13. USEPA. EPA/540/R96/509 p. 166 (1996)
15. Weaver, J.W., et al. Extraction of degradation rate constants from the St. Joseph, Michigan, trichloroethene site. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX September 11 - 13 (1996)
16. Wiedemeier, T.H., et al. Natural attenuation of chlorinated aliphatic hydrocarbons at Plattsburgh Air Force Base, New York. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX. September 11 - 13 (1996A)
17. Wilson, B.H., et al. Biotransformation of selected alkylbenzenes and halogenated aliphatic hydrocarbons in methanogenic aquifer material: a microcosm study. *Environ Sci Technol* 20: 997-1002 (1986)
18. Wilson, B.H., et al. Reductive dechlorination of trichloroethylene in anoxic aquifer material from Picatinny Arsenal, New Jersey. In: USGS Toxic Substances Hydrology Program - Proceedings of the Technical Meeting, Monterey, CA, March 11-15. Morganwalp, D.W. and Aronson, D.A. (eds.). USGS Water Resources Investigations Report. 91-4034 pp. 704-707 (1991)

19. Wilson, B.H., et al. Design and interpretation of microcosm studies for chlorinated compounds. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Hyatt Regency Dallas. Dallas, TX, September 11-13. USEPA. EPA/540/R-96/509 pp. 21-28 (1996)
20. Wilson, J.T., et al. Intrinsic bioremediation of TCE in ground water at an NPL site in St. Joseph, Michigan. In: Proceedings of the EPA Symposium on Intrinsic Bioremediation of Ground Water. USEPA. EPA-540/R-94-515 (1994C)
21. Wilson, J.T., et al. A review of intrinsic bioremediation of trichlorethylene in ground water at Picatinny Arsenal, New Jersey, and St. Joseph, Michigan. In: Bioremediation of Hazardous Wastes. Research, Development, and Field Evaluations. USEPA. EPA/540/R-95/532 (1995B)

REFERENCES FOR VINYL CHLORIDE

1. Bradley, P.M. & Chapelle, F.H. Anaerobic mineralization of vinyl chloride in Fe(III) reducing, aquifer sediments. *Environ Sci Technol* 30(6): 2084-2086 (1996B)
2. Ellis, D.E., et al. Remediation technology forum intrinsic remediation project at Dover Air Force Base, Delaware. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. Dallas, TX, September 11 - 13. USEPA. EPA/540/R-96/509 (1996)
3. Wiedemeier, T.H., et al. Natural attenuation of chlorinated aliphatic hydrocarbons at Plattsburgh Air Force Base, New York. In: Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. USEPA Office of Research and Development. EPA/540/R-96/509. Hyatt Regency Dallas, Dallas, TX. September 11 - 13 (1996A)
4. Wilson, J.T., et al. A review of intrinsic bioremediation of trichlorethylene in ground water at Picatinny Arsenal, New Jersey, and St. Joseph, Michigan. In: Bioremediation of Hazardous Wastes. Research, Development, and Field Evaluations. USEPA. EPA/540/R-95/532 (1995B)

REFERENCES FOR PHENOL

1. Arvin, E., et al. Microbial degradation of oil and creosote related aromatic compounds under aerobic and anaerobic conditions. *Int Conf Physiochemical Biol Detoxif Hazard Wastes*. 2: 828-847(1989)
2. Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)
3. Godsy, E.M., et al. Methanogenic degradation kinetics of phenolic compounds in aquifer-derived microcosms. *Biodegradation* 2: 211-221 (1992A)
4. Klecka, G.M., et al. Natural bioremediation of organic contaminants in ground water: Cliffs-Dow Superfund site. *Ground Water* 28(4): 534-543 (1990A)
5. Morris, M.S. Biodegradation of organic contaminants in subsurface systems: kinetic and metabolic considerations. Ph.D. Dissertation. Virginia Polytechnic Institute and State University (1988)

- Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 1. Experimental conditions and fate of phenolic compounds. *J Contam. Hydrol* 20: 27-50 (1995A)
- Smith, J.A. & Novak, J.T. Biodegradation of chlorinated phenols in subsurface soils. *Water, Air, and Soil Pollution* 33: 29-42 (1987)

REFERENCES FOR o-CRESOL

- Arvin, E., et al. Microbial degradation of oil and creosote related aromatic compounds under aerobic and anaerobic conditions. *Int Conf Physiochemical Biol Detoxif Hazard Wastes*. 2: 828-847(1989)
- Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)
- Klecka, G.M., et al. Natural bioremediation of organic contaminants in ground water: Cliffs-Dow Superfund site. *Ground Water* 28(4): 534-543 (1990A)
- Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 1. Experimental conditions and fate of phenolic compounds. *J Contam. Hydrol* 20: 27-50 (1995A)
- Troutman, D.E., et al. Phenolic contamination in the sand-and-gravel aquifer from a surface impoundment of wood treatment wastes, Pensacola, Florida. *USGS Water-Resources Investigations Report* 84-4230 (1984)

REFERENCES FOR m-CRESOL

- Arvin, E., et al. Microbial degradation of oil and creosote related aromatic compounds under aerobic and anaerobic conditions. *Int Conf Physiochemical Biol Detoxif Hazard Wastes*. 2: 828-847(1989)
- Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)
- Troutman, D.E., et al. Phenolic contamination in the sand-and-gravel aquifer from a surface impoundment of wood treatment wastes, Pensacola, Florida. *USGS Water-Resources Investigations Report* 84-4230 (1984)

REFERENCES FOR p-CRESOL

1. Arvin, E., et al. Microbial degradation of oil and creosote related aromatic compounds under aerobic and anaerobic conditions. *Int Conf Physiochemical Biol Detoxif Hazard Wastes*. 2: 828-847(1989)
2. Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)

REFERENCES FOR 2,4-DICHLOROPHENOL

1. Gibson, S.A. & Suflita, J.M. Extrapolation of biodegradation results to groundwater aquifers: reductive dehalogenation of aromatic compounds. *Appl Environ Microbiol* 52: 681-688 (1986)
2. Kjeldsen, P., et al. Sorption and degradation of chlorophenols, nitrophenols and organophosphorus pesticides in the subsoil under landfills-laboratory studies. *J Contam Hydrol* 6: 165-184 (1990)
3. Morris, M.S. Biodegradation of organic contaminants in subsurface systems: kinetic and metabolic considerations. Ph.D. Dissertation. Virginia Polytechnic Institute and State University (1988)
4. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 1. Experimental conditions and fate of phenolic compounds. *J Contam. Hydrol* 20: 27-50 (1995A)
5. Suflita, J.M. & Miller, G.D. Microbial metabolism of chlorophenolic compounds in ground water aquifers. *Environ Sci Technol* 4: 751-758 (1985)

REFERENCES FOR PENTACHLOROPHENOL

1. Bellcore. Investigation of the Anaerobic Biodegradability of Pentachlorophenol. SR-2741, March 1995.
2. Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992).
3. Thomas, J.M., et al. Microbial ecology of the subsurface at an abandoned creosote waste site. *J Ind Microbiol* 4: 109-120 (1989).
4. Troutman, D.E., et al. Phenolic contamination in the sand-and-gravel aquifer from a surface impoundment of wood treatment wastes, Pensacola, Florida. USGS Water-Resources Investigations Report 84-4230 (1984).

REFERENCES FOR TRICHLOROFLUOROMETHANE

1. Cook, P.G., et al. Chlorofluorocarbons as tracers of groundwater transport processes in a shallow, silty sand aquifer. *Water Resources Research* 31(3): 425-434 (1995)

REFERENCES FOR DICHLORODIFLUOROMETHANE

1. Cook, P.G., et al. Chlorofluorocarbons as tracers of groundwater transport processes in a shallow, silty sand aquifer. *Water Resources Research* 31(3): 425-434 (1995)

REFERENCES FOR 1,1,2-TRICHLORO-1,2,2-TRIFLUOROETHANE

1. Cook, P.G., et al. Chlorofluorocarbons as tracers of groundwater transport processes in a shallow, silty sand aquifer. *Water Resources Research* 31(3): 425-434 (1995)

REFERENCES FOR ACENAPHTHENE

1. Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)

REFERENCES FOR FLUORENE

1. Bedient, P.B., et al. Ground water quality at a creosote waste site. *Ground Water* 22: 318-329 (1984)
2. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
3. Sharak Genthner, B.R., et al. Persistence of polycyclic aromatic hydrocarbon components of creosote under anaerobic enrichment conditions. *Arch Environ Contam Toxicol* 32: 99-105 (1997)
4. Wilson, J.T., et al. Influence of microbial adaptation on the fate of organic pollutants in ground water. *Environ Toxicol Chem* 4: 721-726 (1985)

REFERENCES FOR 1-METHYLNAPHTHALENE

1. Albrechtsen, H.J., et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation. Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hinchee, R.E., et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1993)
2. Bedient, P.B., et al. Ground water quality at a creosote waste site. *Ground Water* 22: 318-329 (1984)
3. Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)
4. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
5. Reinhard, M., et al. Occurrence and distribution of organic chemicals in two landfill leachate plumes. *Environ Sci Technol* 18: 953-961 (1984)
6. Wilson, J.T., et al. Influence of microbial adaptation on the fate of organic pollutants in ground water. *Environ Toxicol Chem* 4: 721-726 (1985)

REFERENCES FOR NAPHTHALENE

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)

2. Albrechtsen, H.J., et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation. Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hinchee, R.E., et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1993)
3. Baedecker, M.J., et al. Crude oil in a shallow sand and gravel aquifer-M. Biogeochemical reactions and mass balance modeling in anoxic groundwater. *Applied Geochem* 8: 569-586 (1993)
4. Bedient, P.B., et al. Ground water quality at a creosote waste site. *Ground Water* 22: 318-329 (1984)
5. Godsy, E.M., et al. Methanogenic biodegradation of creosote contaminants in natural and simulated ground-water ecosystems. *Ground Water* 30(2): 232-242 (1992)
6. Klecka, G.M., et al. Natural bioremediation of organic contaminants in ground water: Cliffs-Dow Superfund site. *Ground Water* 28(4): 534-543 (1990A)
7. Lyngkilde, J. & Christensen, T.H. Fate of organic contaminants in the redox zones of a landfill leachate pollution plume (Vejen, Denmark). *J Contam Hydrol* 10: 291-307 (1992)
8. Madsen, E.L., et al. Oxygen limitations and aging as explanations for the field persistence of naphthalene in coal tar-contaminated surface sediments. *Environ Toxicol Chem* 15: 1876-1882 (1996)
9. Nielsen, P.H., et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)
10. Nielsen, P.H., et al. A field method for determination of groundwater and groundwater-sediment associated potentials for degradation of xenobiotic organic compounds. *Chemosphere* 25: 449462 (1992)
11. Nielsen, P.H. & Christensen, T.H. In situ measurement of degradation of specific organic compounds under aerobic, denitrifying, iron(RD-reducing, and methanogenic groundwater conditions. In: Bioremediation of Chlorinated and PAH Compounds. Hinchee, R.E. (ed.). Lewis pp 416-422 (1994)
12. Thierrin, J., et al. A ground-water tracer test with deuterated compounds for monitoring in situ biodegradation and retardation of aromatic hydrocarbons. *Ground Water* 33: 469-475 (1995)
13. Wilson, J.T., et al. Influence of microbial adaptation on the fate of organic pollutants in ground water. *Environ Toxicol Chem* 4: 721-726 (1985)
14. Zoeteman, B.C.J., et al. Persistency of organic contaminants in groundwater, lessons from soil pollution incidents in the Netherlands. *Sci Total Environ* 21: 187-202 (1981)

REFERENCES FOR 1,1'-BIPHENYL

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)

REFERENCES FOR CUMENE

1. Acton, D.W. & Barker, J.F. *In situ* biodegradation potential of aromatic hydrocarbons in anaerobic groundwaters. *J Contam Hydrol* 9: 325-52 (1992)

REFERENCES FOR DIOXANE

1. Nyer, E.K., et al. Biochemical effects on contaminant fate and transport. *GWMR Spring*: 80-82 (1991)

REFERENCES FOR METHANOL

1. API. Transport and fate of dissolved methanol, methyl-tertiary-butyl-ether, and monoaromatic hydrocarbons in a shallow sand aquifer. Appendix H: Laboratory biotransformation studies. American Petroleum Institute. Health Environ Sci Dept (1994)
2. Hickman, G.T., et al. Effects of site variations on subsurface biodegradation potential. *Journal WPCF* 61(9): 1564-1575 (1990)
3. Morris, M.S. & Novak, J.T. Mechanisms responsible for the biodegradation of organic chemicals in subsurface systems. In: *Toxic and Hazardous Wastes. Proceedings of the 19' Mid-Atlantic Industrial Waste Conference*. Evans, JC (ed.). 19: 123-136 (1987)
4. Morris, M.S. Biodegradation of organic contaminants in subsurface systems: kinetic and metabolic considerations. Ph.D. Dissertation. Virginia Polytechnic Institute and State University (1988)
5. Novak, J.T., et al. Biodegradation of methanol and tertiary butyl alcohol in subsurface systems. *Wat Sci Tech* 17: 71-85 (1985)
6. White, K.D., A comparison of subsurface biodegradation rates of methanol and tertiary butanol in contaminated and uncontaminated sites. Ph.D. Dissertation, Virginia Polytechnic Institute and State University (1986)
7. Wilson, W.G., et al. Enhancement of biodegradation of alcohols in groundwater systems. In: *Toxic and Hazardous Wastes, Proceedings of the Mid-Atlantic Industrial Waste Conference*. 18: 421430(1986)
8. Wilson, W.G. & Novak, J.T. Biodegradation of organic compounds in anoxic groundwater systems. In: *Proceedings of the 42nd Industrial Waste Conference May 12-14, West Lafayette, IN: Lewis Publishers Inc.* 197-205 (1988)

REFERENCES FOR NITROBENZENE

1. Nielsen, P. H. et al. In situ and laboratory studies on the fate of specific organic compounds in an anaerobic landfill leachate plume, 2. Fate of aromatic and chlorinated aliphatic compounds. *J Contam Hydrol* 20: 51-66 (1995B)

REFERENCE FOR PYRIDINE

1. Adrian, N.R. & Suflita, J.M. Anaerobic biodegradation of halogenated and nonhalogenated N-, S-, and O-heterocyclic compounds in aquifer slurries. *Environ Toxicol Chem* 13(10): 1551-1557 (1994)

2. Kuhn, E.P. & Suflita, J.M. Microbial degradation of nitrogen, oxygen and sulfur heterocyclic compounds under anaerobic conditions: Studies with aquifer samples. *Environ Toxicol Chem* 8: 1149-1158 (1989)

REFERENCES FOR STYRENE

1. Roberts, P.V., et al. Organic contaminant behavior during groundwater recharge. *J Water Pollut Control Fed.* 52: 161-72 (1980)

REFERENCES FOR TRIMETHYLBENZENE

1. Albrechtsen, H.J., et al. Landfill leachate-polluted groundwater evaluated as substrate for microbial degradation under different redox conditions. In: Applied Biotechnology Site Remediation, Pap Int Symp, In Site On-Site Bioreclam. 2nd. Hincbee, R.E., et al. (eds.). Lewis: Boca Raton, FL pp. 371-378 (1993)
2. Thierrin, J., et al. A ground-water tracer test with deuterated compounds for monitoring in situ biodegradation and retardation of aromatic hydrocarbons. *Ground Water* 33: 469-475 (1995)
3. Wiedemeler, T.H., et al. Approximation of biodegradation rate constants for monoaromatic hydrocarbons (BTEX) in ground water. *Ground Water Monit Remediat.* 16: 186-194 (1996)
4. Wilson, J.T., et al. Intrinsic bioremediation of JP-4 jet fuel. In: *Symposium on Intrinsic Bioremediation of Ground Water*. Denver, CO. EPA/540/R-94/515. Washington, DC: USEPA (1994A)