

US EPA ARCHIVE DOCUMENT



Ohio EPA HAB Response and Lessons Learned

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For assistance in accessing this document please send
an email to EPACyanoHABs@epa.gov

Summary of Ohio HAB Response

2010: Ohio EPA began sampling for cyanotoxins at public water systems (PWSs)

- Finished water detection at inland PWS.

2011: Ohio EPA/ODNR/ODH created Ohio HAB Response Strategy (updated annually)

2013: Finished water microcystins threshold exceedance at small PWS

- Drinking Water Advisory Issued.

2014: Finished water microcystins threshold exceedance at large PWS

- Drinking Water Advisory Issued.

2015: Revised Response Strategy to include U.S. EPA health advisories for microcystins and cylindrospermops

- Finished water microcystins detections at 5 PWSs

- No Drinking Water Advisories Issued.

- Ohio Senate Bill 1 passed

2016: HAB Monitoring and Reporting Rules

- Effective June 1, 2016
- Updated response strategies, new treatment optimization and general plan guidance for PWSs



Ohio HAB Rules Overview

HAB Rules: epa.ohio.gov/ddagw/rules.aspx

- PWS requirements - new rules in OAC Chapter 3745-90
 - Microcystins action levels in drinking water
 - Monitoring requirements: Microcystins and Cyanobacteria Screening
 - Treatment technique requirements
 - Public notification and Consumer Confidence Report (CCR) requirements
 - Recordkeeping requirements
- Laboratory Certification requirements –
New OAC rule 3745-90-04 and amended rules in Chapter 3745-89
 - Laboratory certification
 - Analytical techniques
 - Reporting deadlines



epa.ohio.gov/Portals/28/documents/labcert/TotalMicrocystins.pdf

Ohio Monitoring Requirements

- Total Microcystins

May – October

- Weekly raw and finished water
 - Eligible for monitoring reductions starting May 2017
- Raw water detections >5 ug/L and any finished water detections trigger additional sampling

November – April

- Raw water only every other week
- Detections trigger additional monitoring

- Cyanobacteria Screening (qPCR)

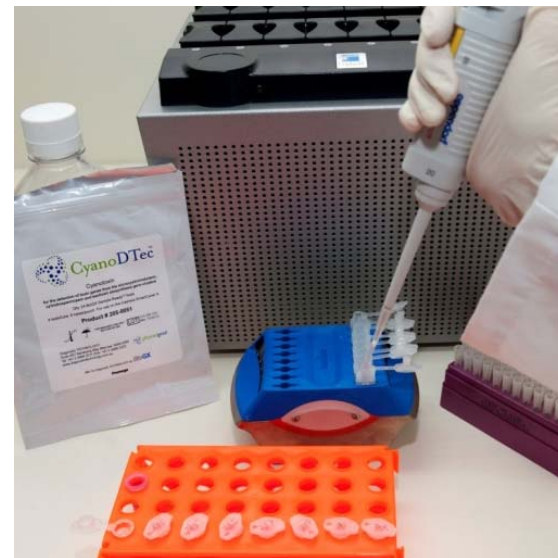
All year

- Biweekly raw water
- Triggers follow up sampling by Ohio EPA for other cyanotoxins



Cyanobacteria Screening: Molecular Methods (Multiplex qPCR)

- Quantitative polymerase chain reaction (qPCR) – identifies and quantifies the presence of genes unique to:
 - Cyanobacteria (16S rDNA)
 - Microcystin and Nodularin production (mcyE gene)
 - Cylindrospermopsin production (cyrA gene)
 - Saxitoxin production (sxtA gene)
 - Test completed within 2-3 hours
 - Scalable
 - Cost-effective
 - Utilizes certified reference material
 - Specific
- Ohio EPA method and certification in 2017
- Ohio EPA uses the data to trigger saxitoxins and cylindrospermopsin sampling and in 2017 will be used as trigger for microcystins monitoring.
- www.phytoxigene.com/products/



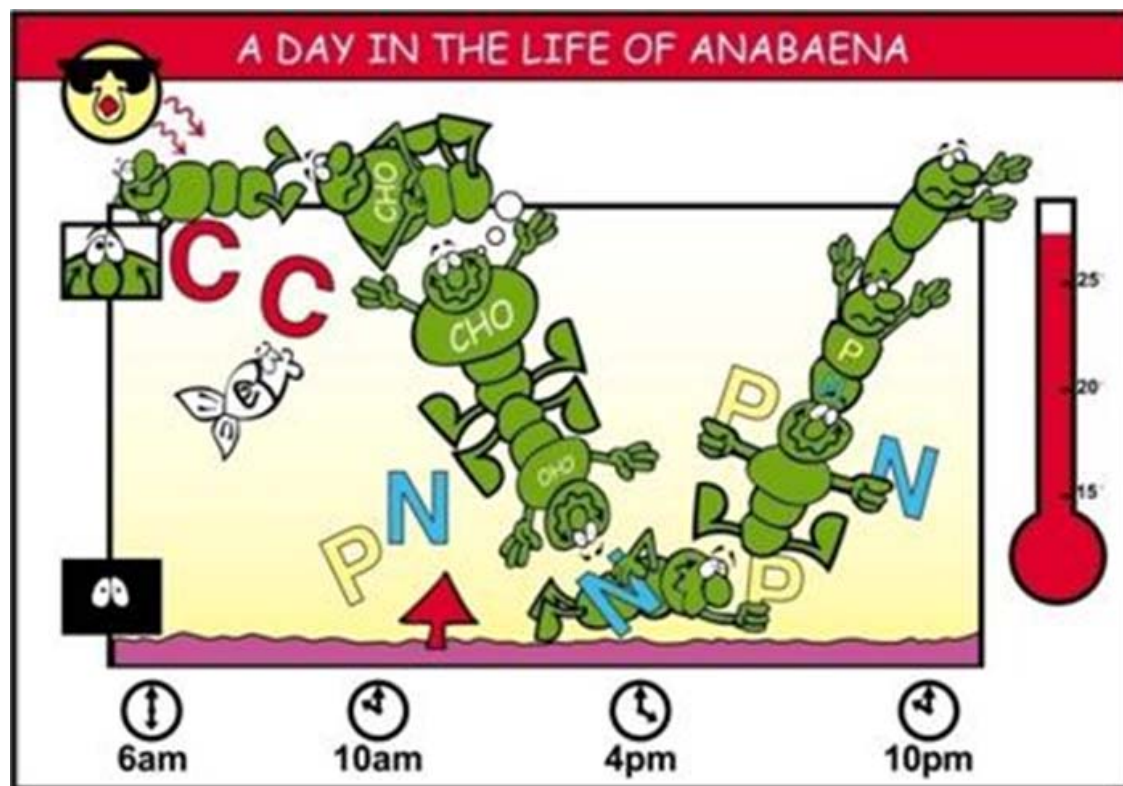
Rule Implementation & Monitoring Results

- 100% compliance with rules!
- Microcystins detected in raw water at 41 PWSs (33% of surface water systems)
- No finished water microcystins detections
- Saxitoxin genes (sxtA) detected in 33 PWSs (27%)
- Saxitoxins detected in raw water at 15 PWSs (12%); 6 finished water saxitoxins detections (none above threshold)
- Cylindrospermopsin genes detected at 1 PWS, no cylindrospermopsin detections.
- qPCR is an effective screening tool for microcystins and saxitoxins.



Lessons Learned – HABs are Not Always Visually Apparent

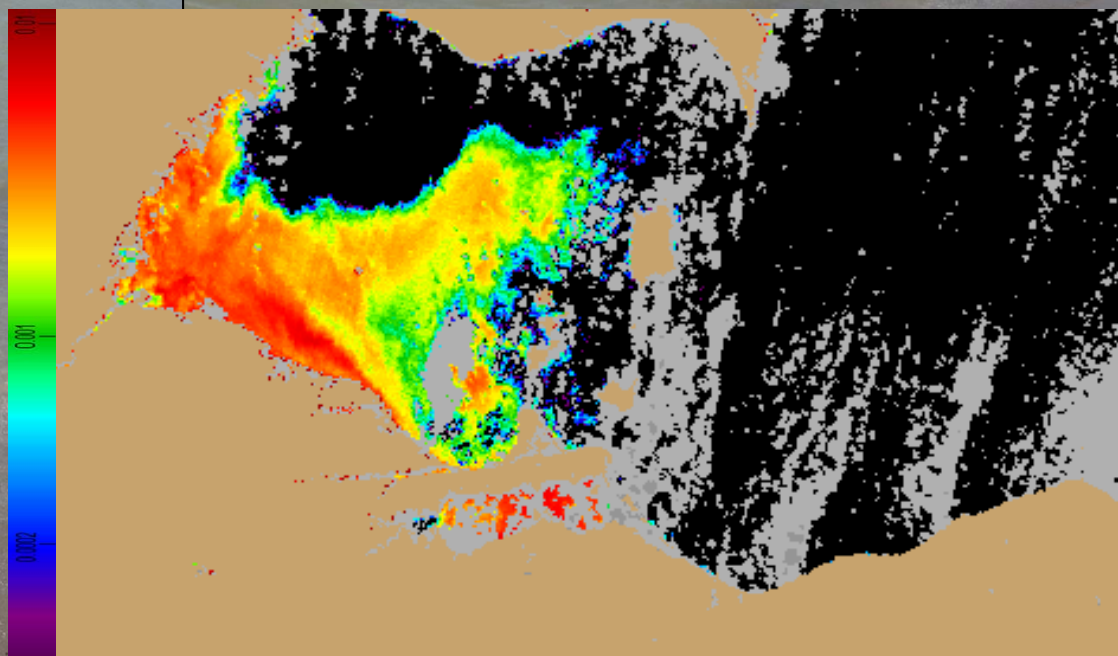
- Many cyanobacteria do not produce surface scums
- Some cyanobacteria occur at depth in the water column or attach to a substrate (benthic)
- Some cyanobacteria move up and down in the water column



Complicates Monitoring

Lake Erie Beach
Microcystis bloom
 August 31, 2011

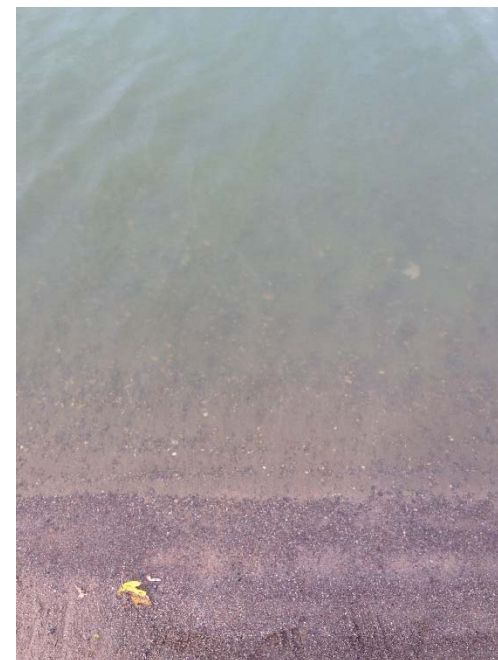
Microcystins >100 ug/L





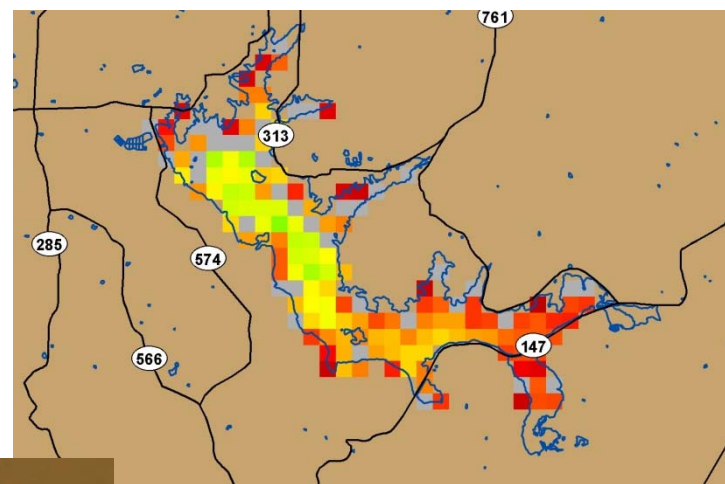
Celina Intake on Grand Lake Saint Mary,
September 2015, *Planktothrix* bloom

Microcystins 185 ug/L



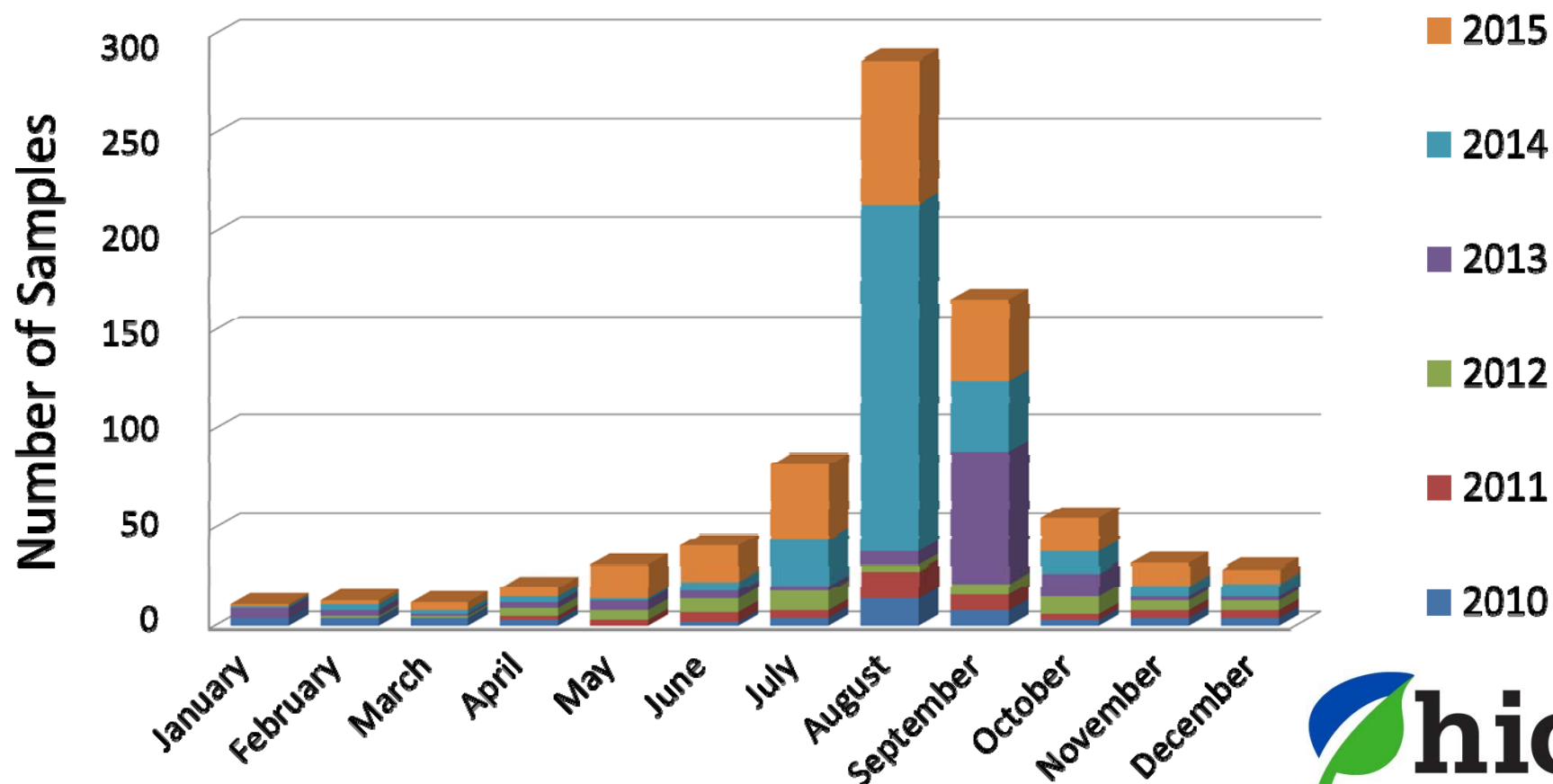
Chippewa Lake, November 2016
Aphanizomenon and *Planktothrix* bloom

Average Microcystins at Beaches 48-58 ug/L



Lessons Learned – HAB Occurrence Year-Round

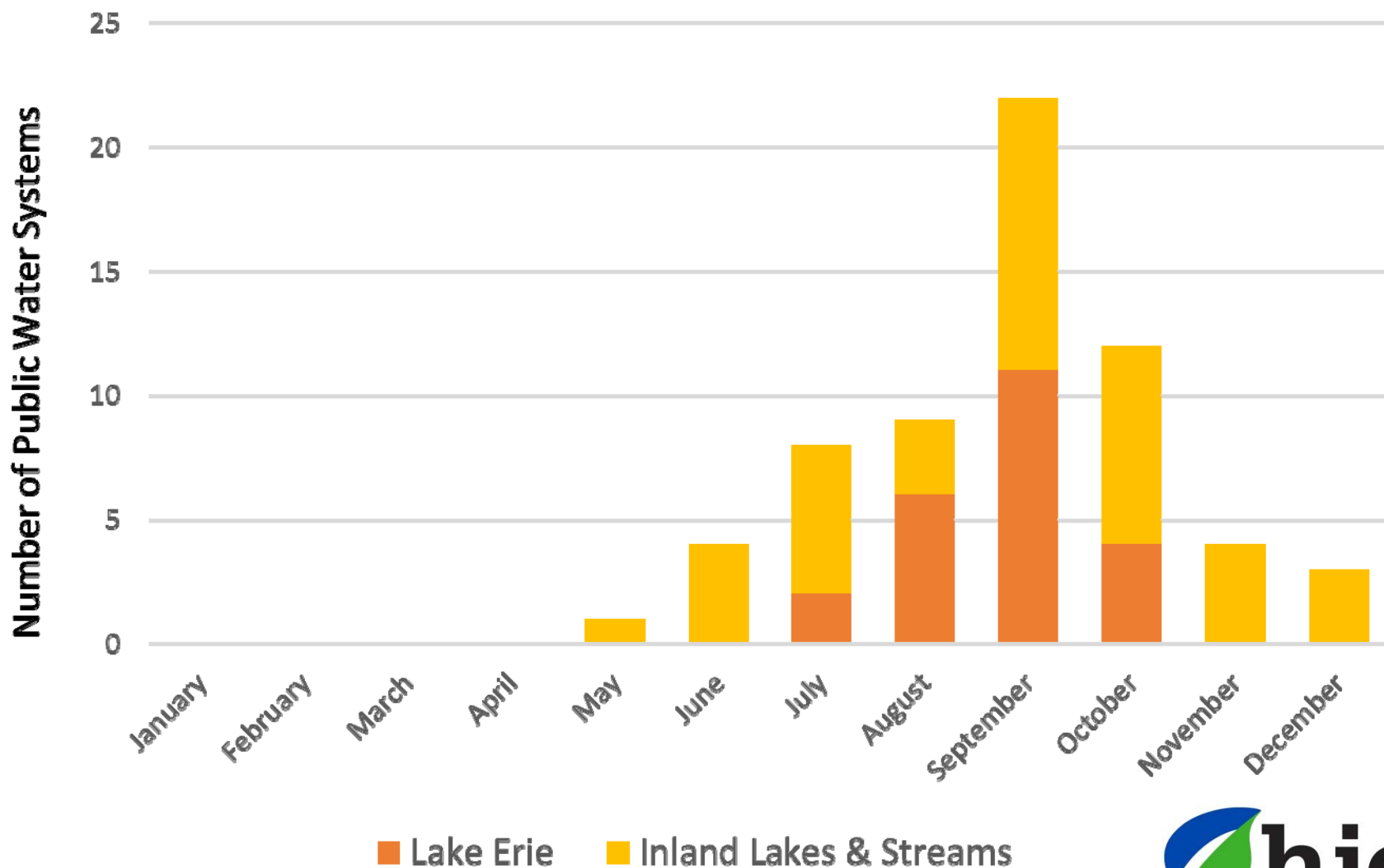
Frequency of Ohio PWS Source Water
Microcystins Detections > 1.6 ug/L



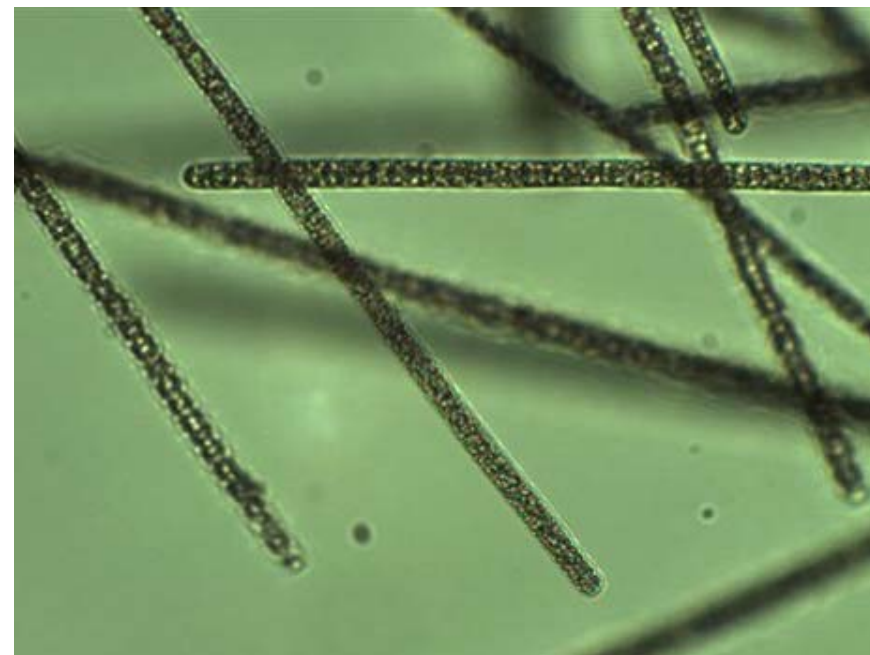
745 samples >1.6 ug/L microcystins, out of 3583 total samples (21 %).

44% of samples were > 0.30 ug/L microcystins. Does not include 2016 results.

Maximum Microcystins Detection By Month



Based on Microcystins Detections at 62 Public Water Systems
(2010 - 2016)

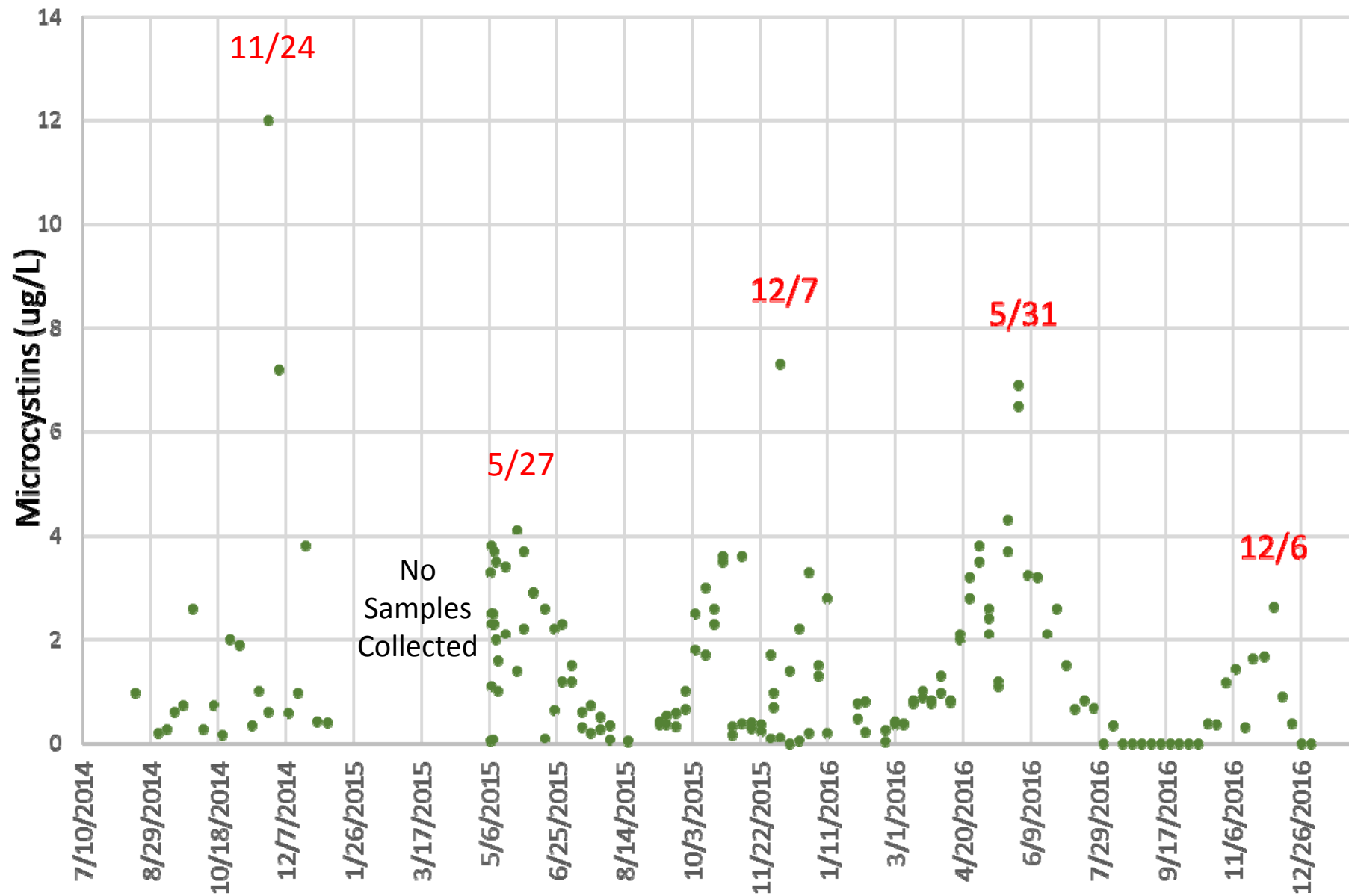


William's Reservoir November, 2012

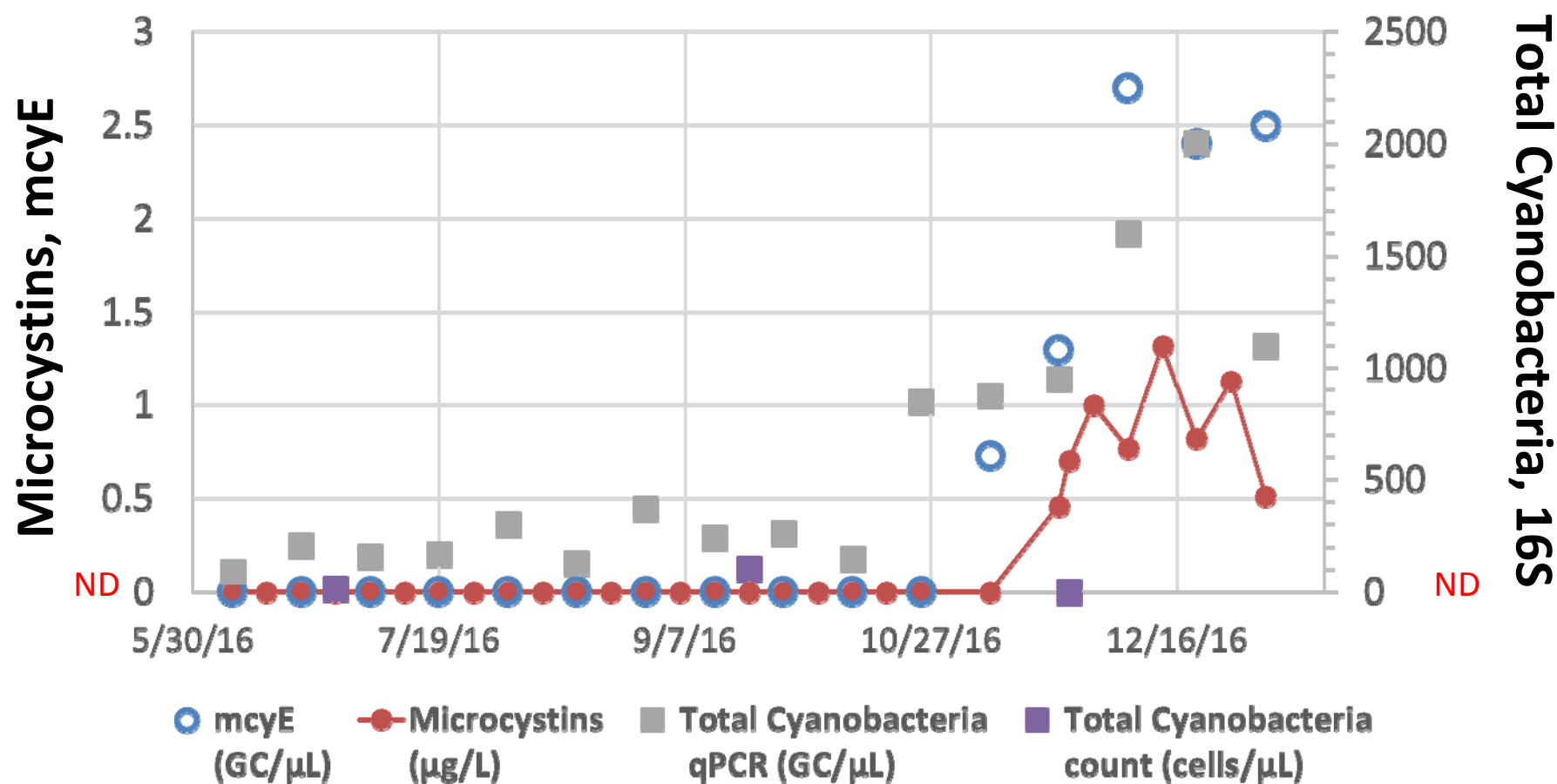
**Microcystin
Concentration: 1400 ug/L**



Raw Water Microcystins at an Inland PWS – Microcystins Do Not Always Peak in Summer



Late Fall-Winter Microcystins and qPCR Trends

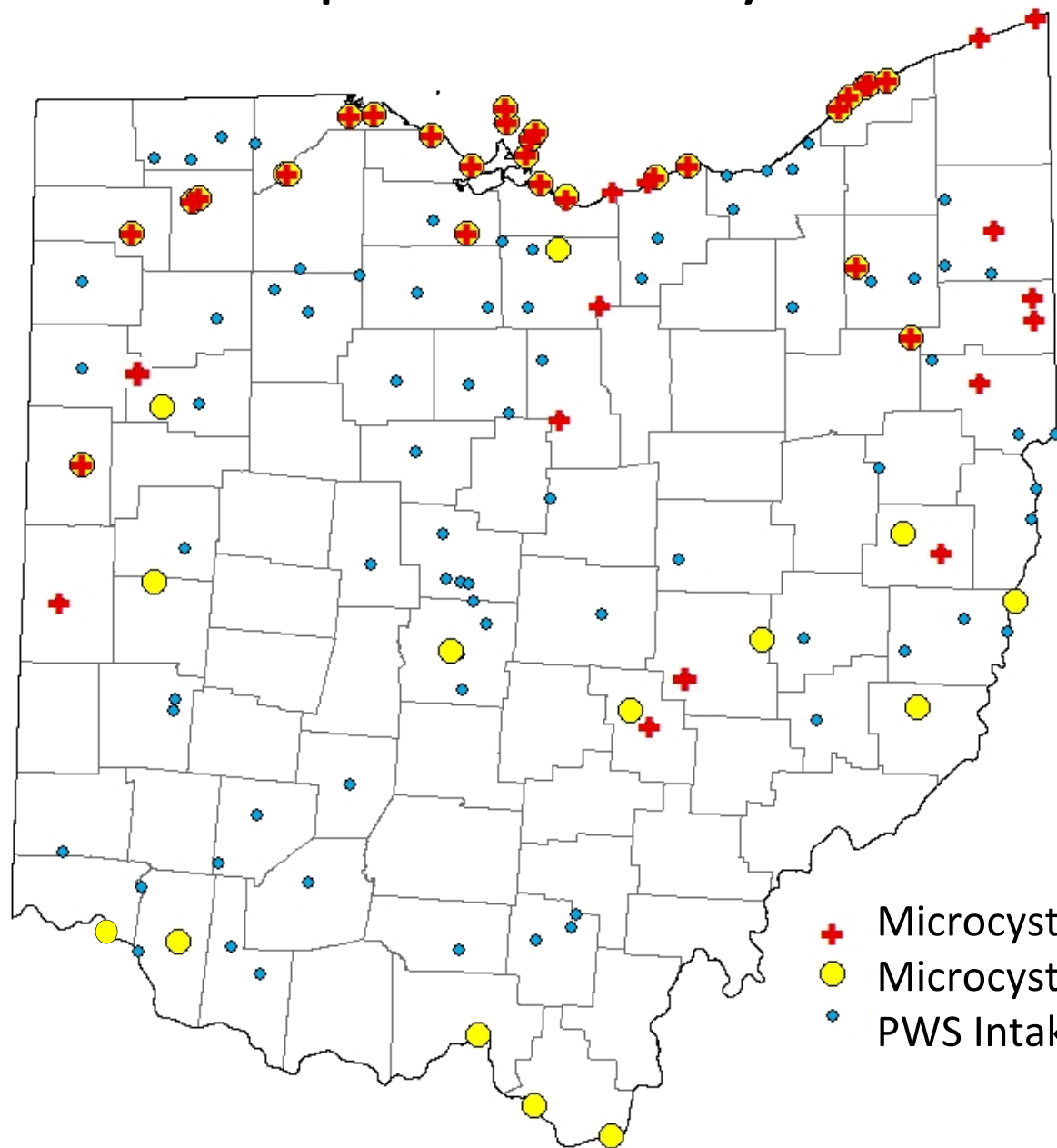


Widespread Microcystins Occurrence in Ohio

Source Water Microcystins Detections at Ohio PWSs

Data Available for 119
Public Water System
Source Waters

(includes ~2000
compliance samples
collected in 2016)

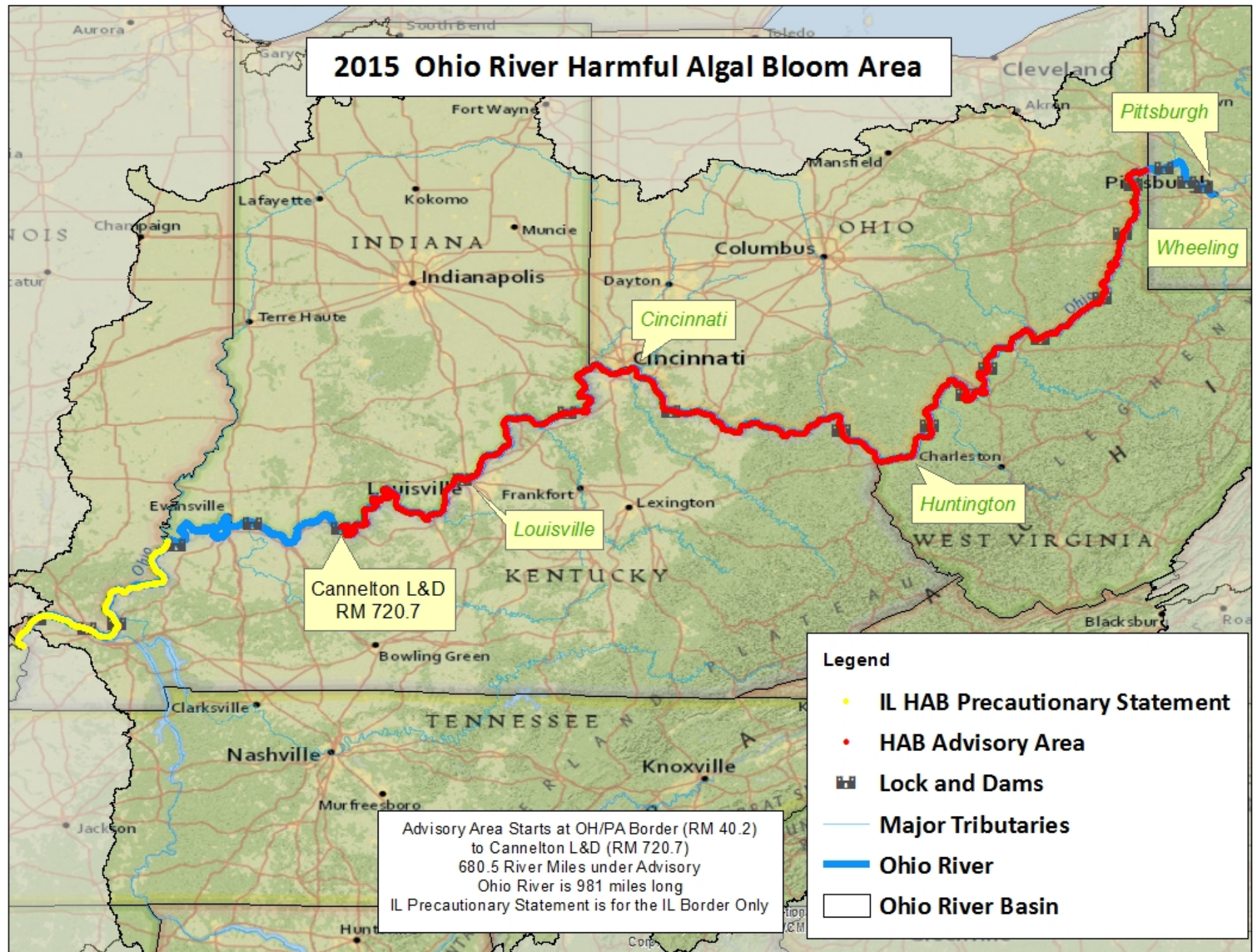


- Microcystins Detections June -Dec. 2016
- Microcystins Detections 2010-2015
- PWS Intakes

Widespread Occurrence: Lake Erie, Inland Lakes, Upground Reservoirs, Quarries, and Rivers

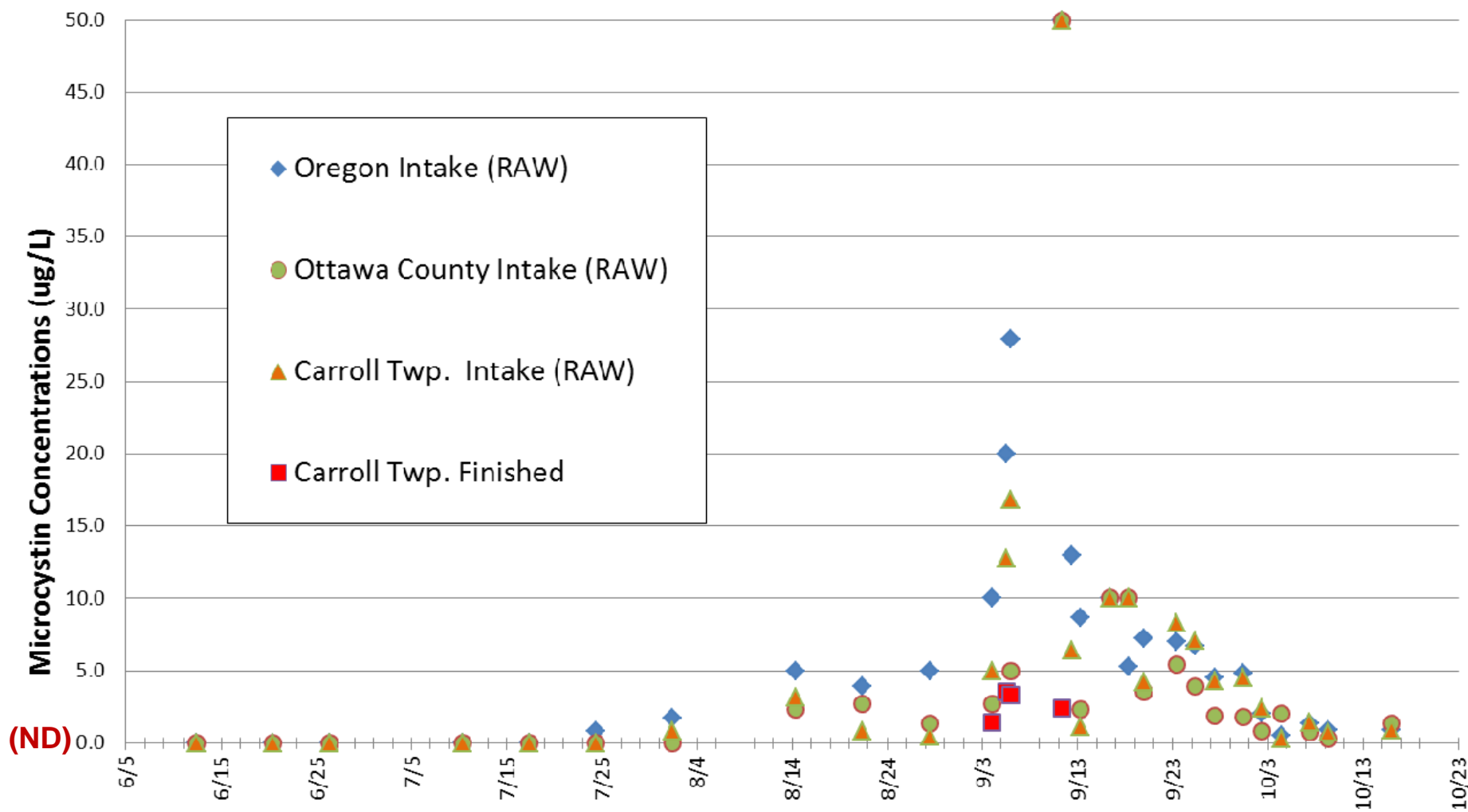


2015 Ohio River Harmful Algal Bloom Area



Microcystins Concentrations Can Increase Rapidly

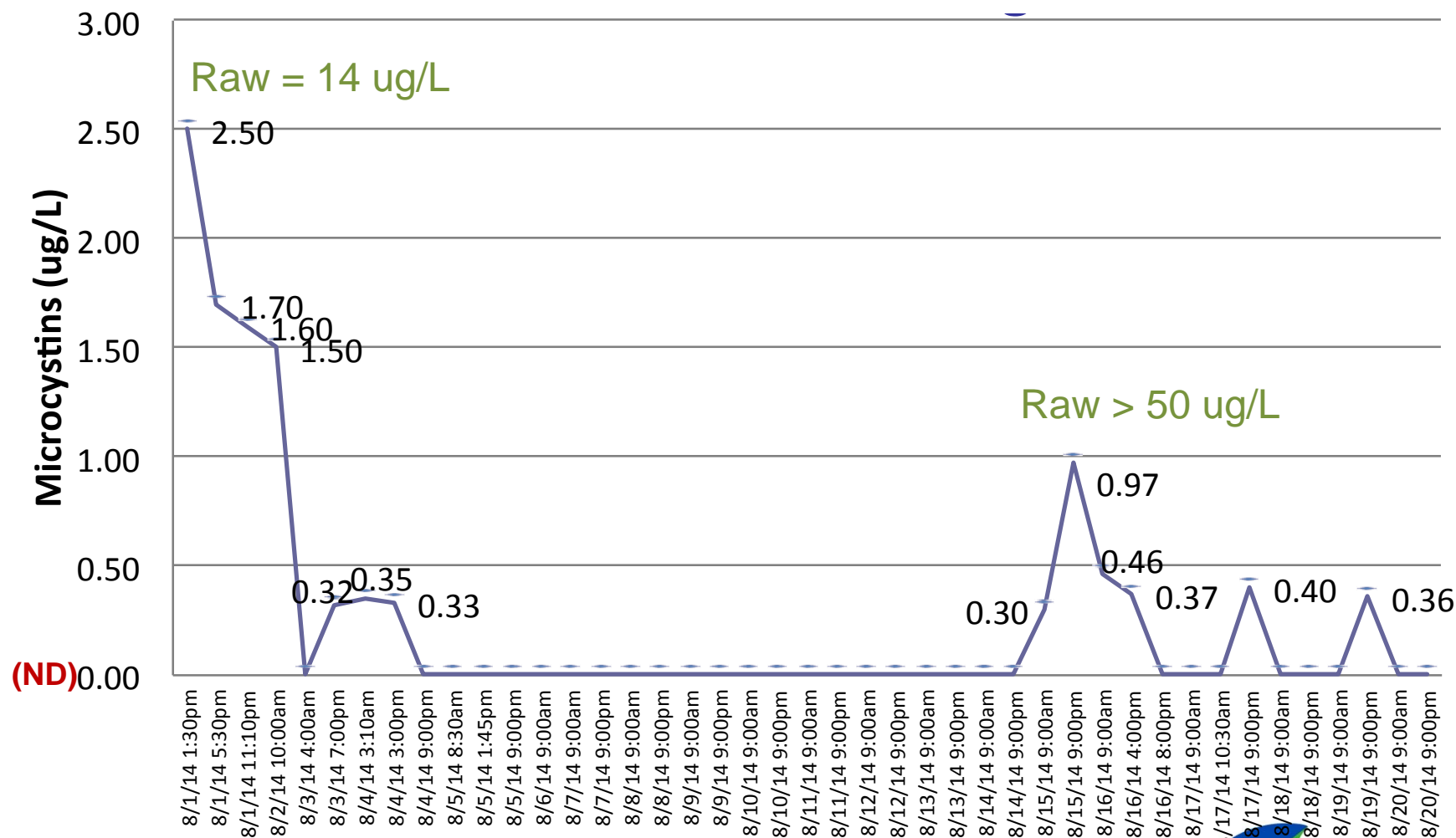
2013 Western Lake Erie Basin Microcystin Concentrations



ND= Not Detected (Concentration <0.3)

Source: Oregon

Microcystin Source Concentration and Treatment Capacity (Finished Water Concentrations Lake Erie PWS)



Microcystins also detected at 31
distribution sites

ND= Not Detected (Concentration <0.30)

Microcystins Concentration
Micrograms per Liter (ug/L)

Legend:

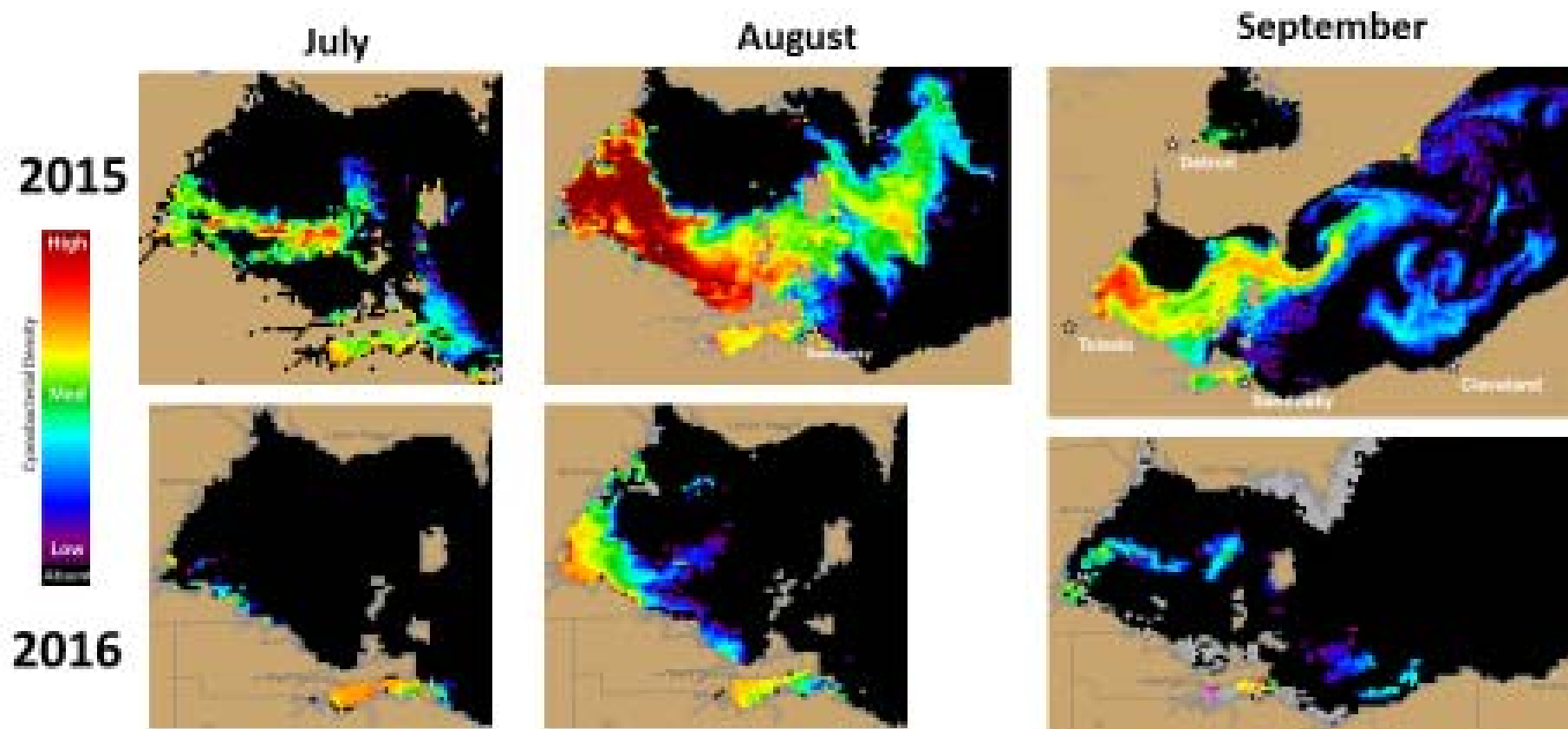
- 2011 (Blue diamond)
- 2012 (Green circle)
- 2013 (Green asterisk)
- 2014 (Red square)
- 2015 (Orange triangle)
- 2016 (Black diamond)

* Microcystins not detected at Cleveland Intakes

Location	2011	2012	2013	2014	2015	2016
Toledo	~5	~25	~25	~50	~5	~1.5
Oregon	~5	~0.3	~50	~35	~5	~1.5
Carroll Township	~4	~1.2	~50	~18	~10	~1.2
Ottawa County	~2.5	~2.5	~50	~2.5	~12	~2.5
Lake Erie Utilities	~0.3	~1.2	~1.2	~4	~4	~0.6
Put-in-Bay	~0.4	~1.2	~4	~1.2	~4	~1.2
Kelleys Island	~0.4	~1.2	~1.2	~1.2	~6	~0.4
Camp Patnos	~0.4	~2.5	~2.5	~2.5	~25	~0.4
Marblehead	~0.5	~0.5	~0.5	~0.8	~1.8	~0.8
Sandusky	~0.3	~0.5	~1.2	~1.5	~2.5	~0.5
Huron	~1.0	~0.8	~3	~0.8	~2	~1.0
Vermillion	~1.0	~1.0	~1.0	~1.0	~1.0	~1.0
Elyria	~1.0	~0.3	~0.3	~0.3	~0.3	~1.0
Lorain	~0.6	~0.6	~0.6	~0.6	~0.6	~0.6
Avon Lake	~0.8	~0.8	~0.8	~0.8	~0.8	~0.8
Cleveland*	Not detected	Not detected	Not detected	Not detected	Not detected	Not detected
Lake County West	~1.0	~1.0	~1.0	~1.0	~1.5	~1.0
Mentor	~0.7	~0.7	~0.7	~0.7	~2.0	~0.7
Painesville	~1.0	~1.0	~1.0	~1.0	~4.0	~1.0
Fairport Harbor	~1.2	~1.2	~1.2	~1.2	~1.2	~1.2
Lake County East	~0.7	~0.7	~0.7	~0.7	~0.8	~0.7
Ashtabula	~0.5	~0.5	~0.5	~0.5	~0.5	~0.5
Conneaut	~0.4	~0.4	~0.4	~0.4	~0.4	~0.4

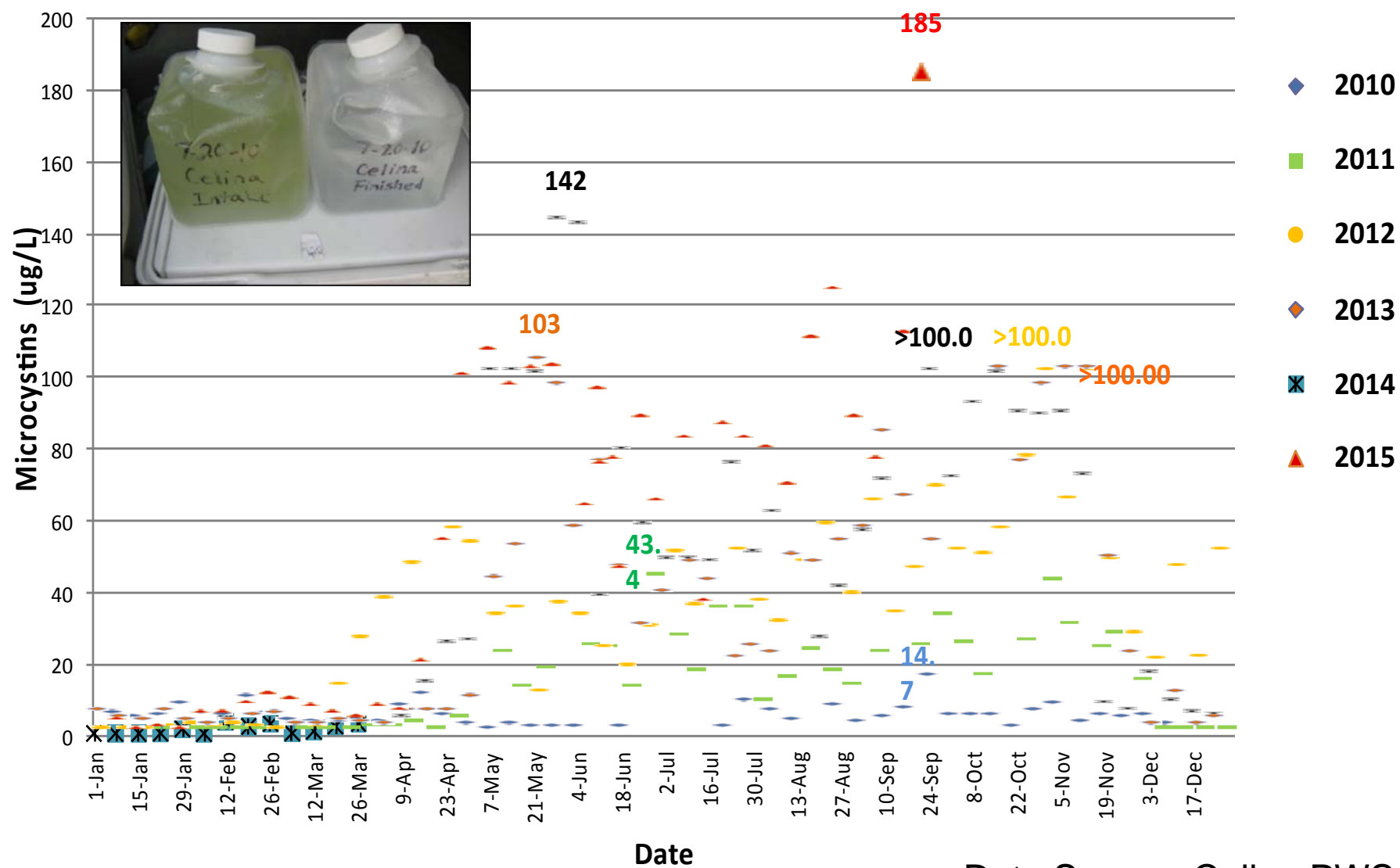
Microcystins
not detected
at Cleveland
Intakes**

Lake Erie HAB Comparison 2015 and 2016



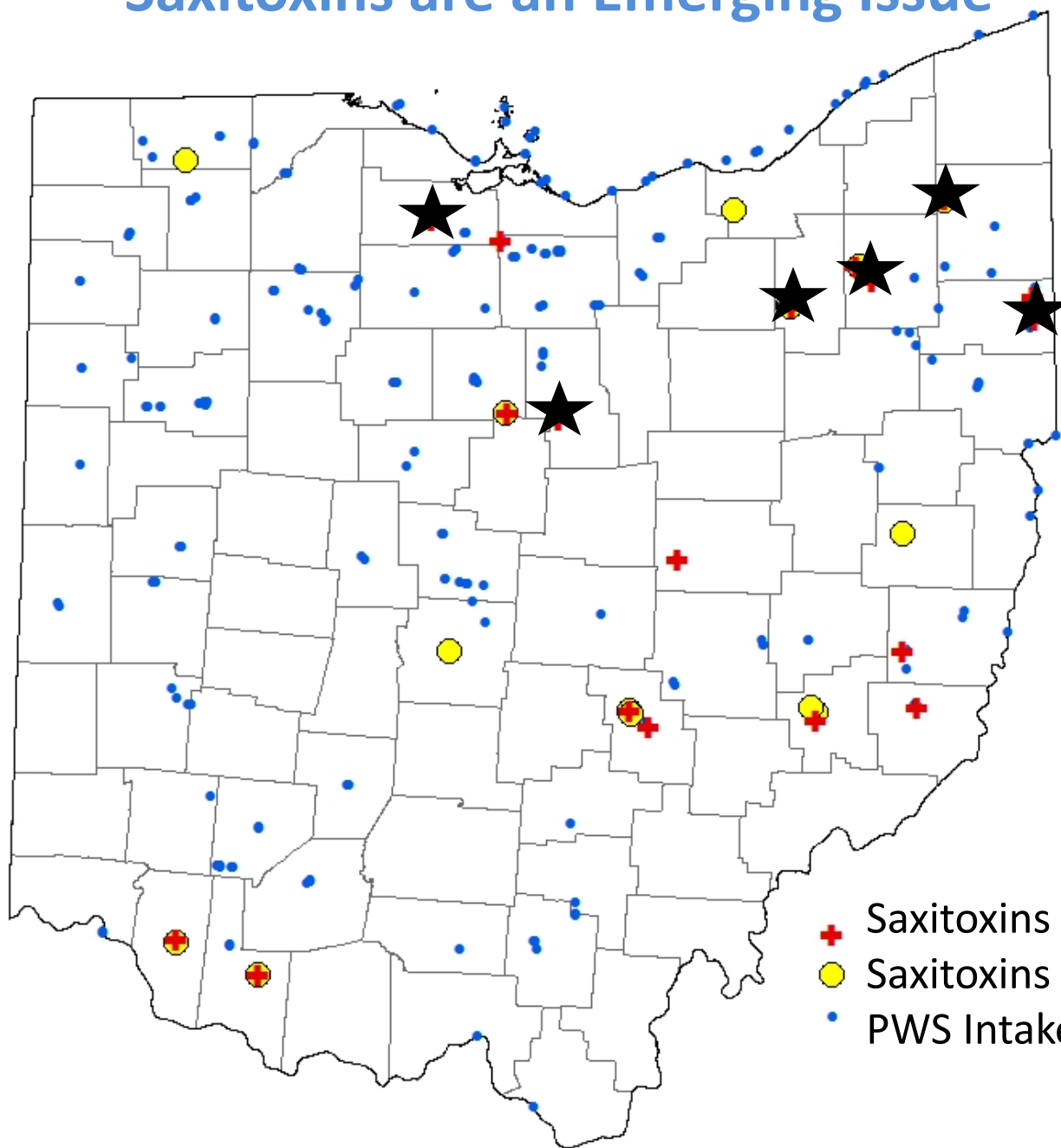
Was 2016 a good year to benchmark?

Grand Lake St. Marys Microcystin Concentrations at City of Celina Intake (Raw Water)



Data Source: Celina PWS

Saxitoxins are an Emerging Issue

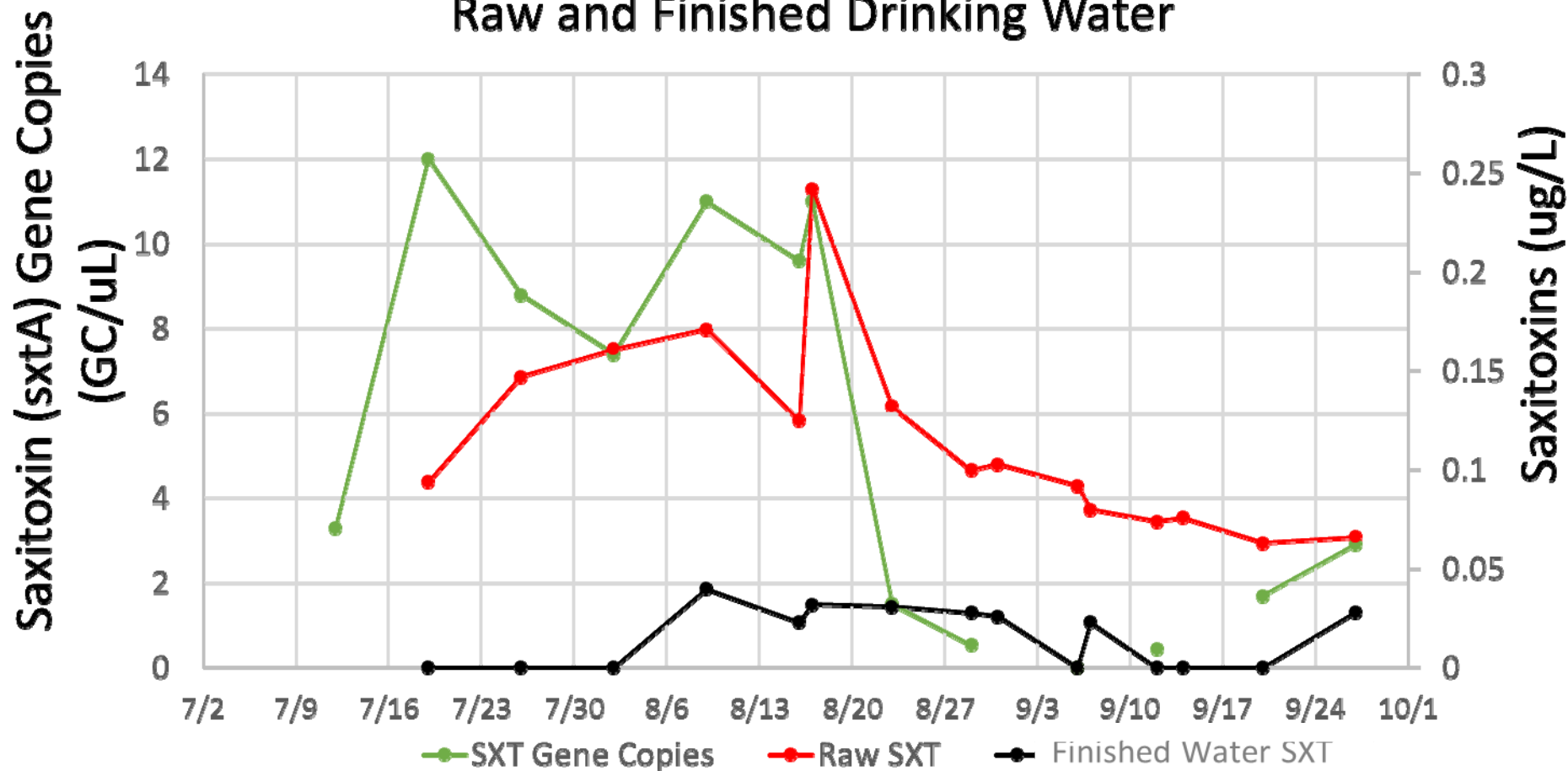


★ 2016 Finished
Water Saxitoxins
Detections
(below threshold)

Benthic cyanobacteria
may be a source of
freshwater saxitoxins.

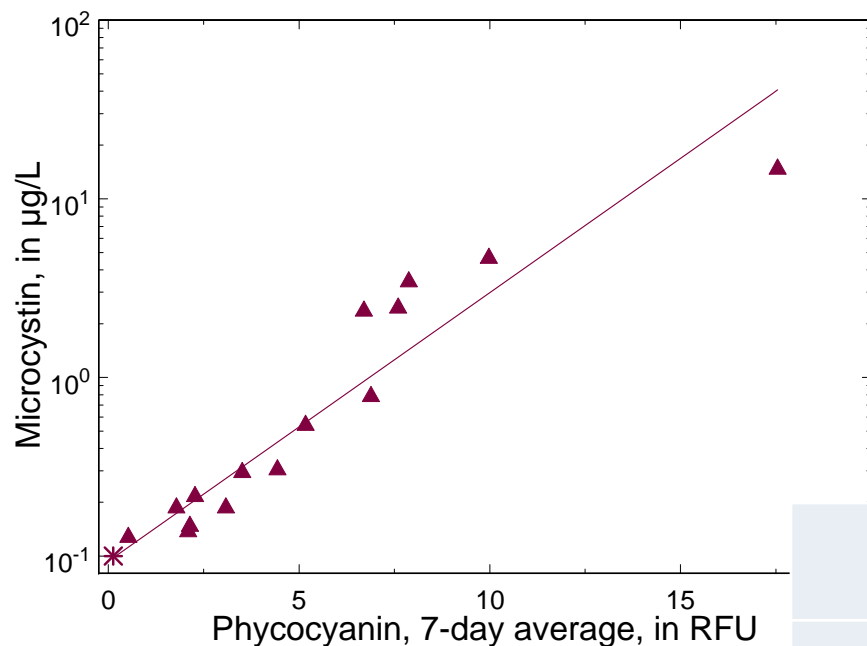
+ Saxitoxins Detections 2016
● Saxitoxins 2010-2015
● PWS Intakes

Saxitoxin Gene and Saxitoxins Detections in Raw and Finished Drinking Water



Multiplex Assay Works: Microcystin (mcyE) genes and microcystins also detected; Cylindrospermopsin (cyrA) genes also detected (cylindrospermopsin was not detected)

Use of Multi-Parameter Datasondes for Continuous Monitoring

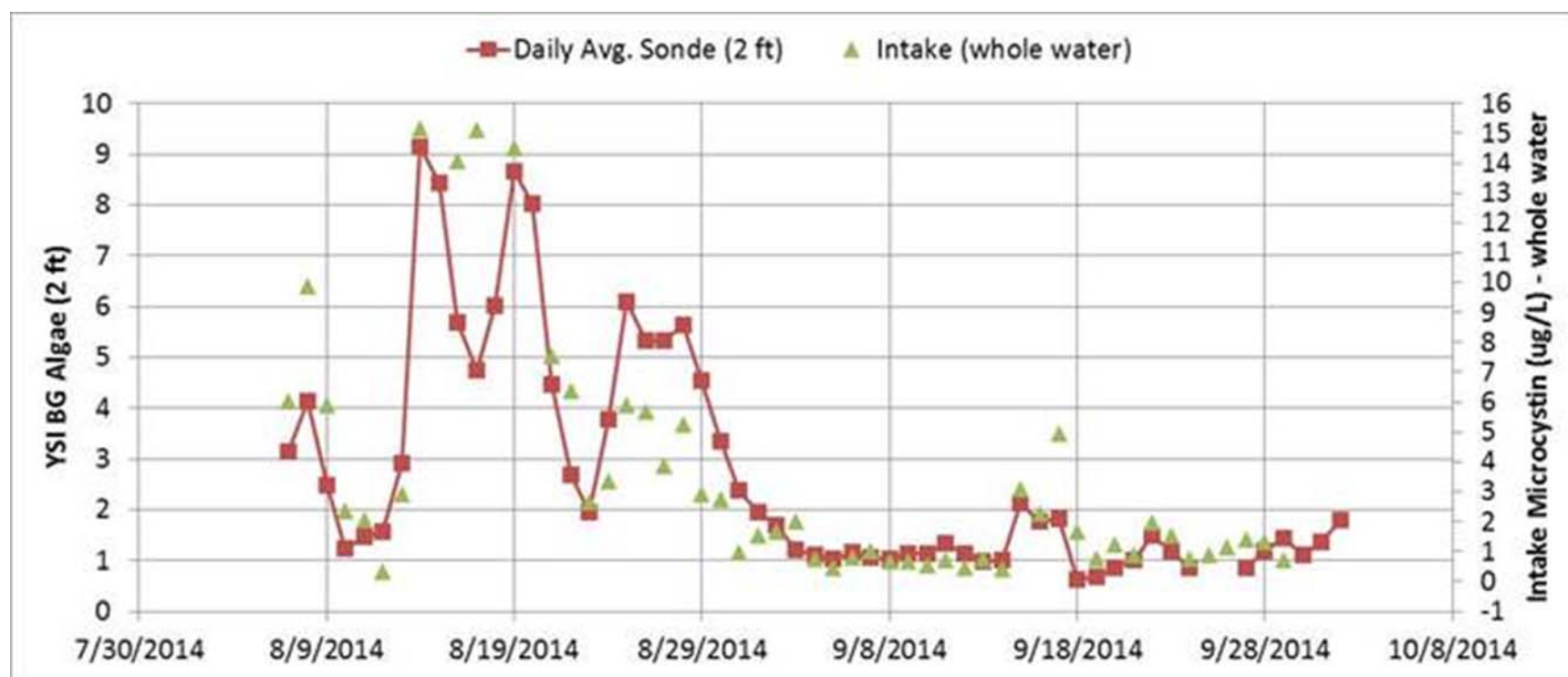


Spearman's correlation to microcystin concentrations

	rho	p
Phycocyanin, 7-day average	0.98	<0.0001
Dissolved oxygen, 14-day average	0.88	<0.0001
pH, 7-day average	0.83	<0.0001
Temperature, instantaneous 10 a.m.	0.73	0.0031
Chlorophyll, 24-hour average	0.53	0.0358
Specific conductance, 3-day average	-0.20	0.4473

Data Courtesy Donna Francy, USGS

Microcystin Concentration and Phycocyanin Fluorescence at Toledo's Intake



-Graph provided to Ohio EPA by Ed Verhamme, Limnotech.

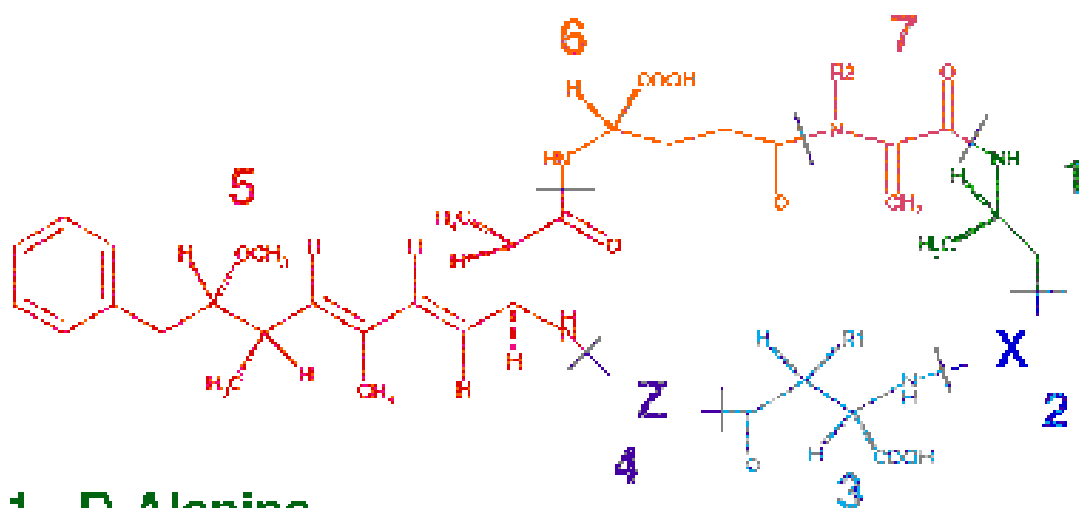
Ohio EPA provided funding to 36 PWSs for purchase of phycocyanin sondes or readers

Phycocyanin Data Interpretation

- Phycocyanin Concentrations vary based on type of cyanobacteria present, turbidity of the water and other factors.
- Relative/Raw Fluorescence Units (RFUs) better than Cell Counts.
 - Can calibrate to cell counts in source water, but this can change if cyanobacteria genera shift or turbidity changes.
- Evaluate trends, not absolute values.

Lessons Learned – Analytical Methods Matter

No “Perfect” Analytical Method for Detecting TOTAL Microcystins



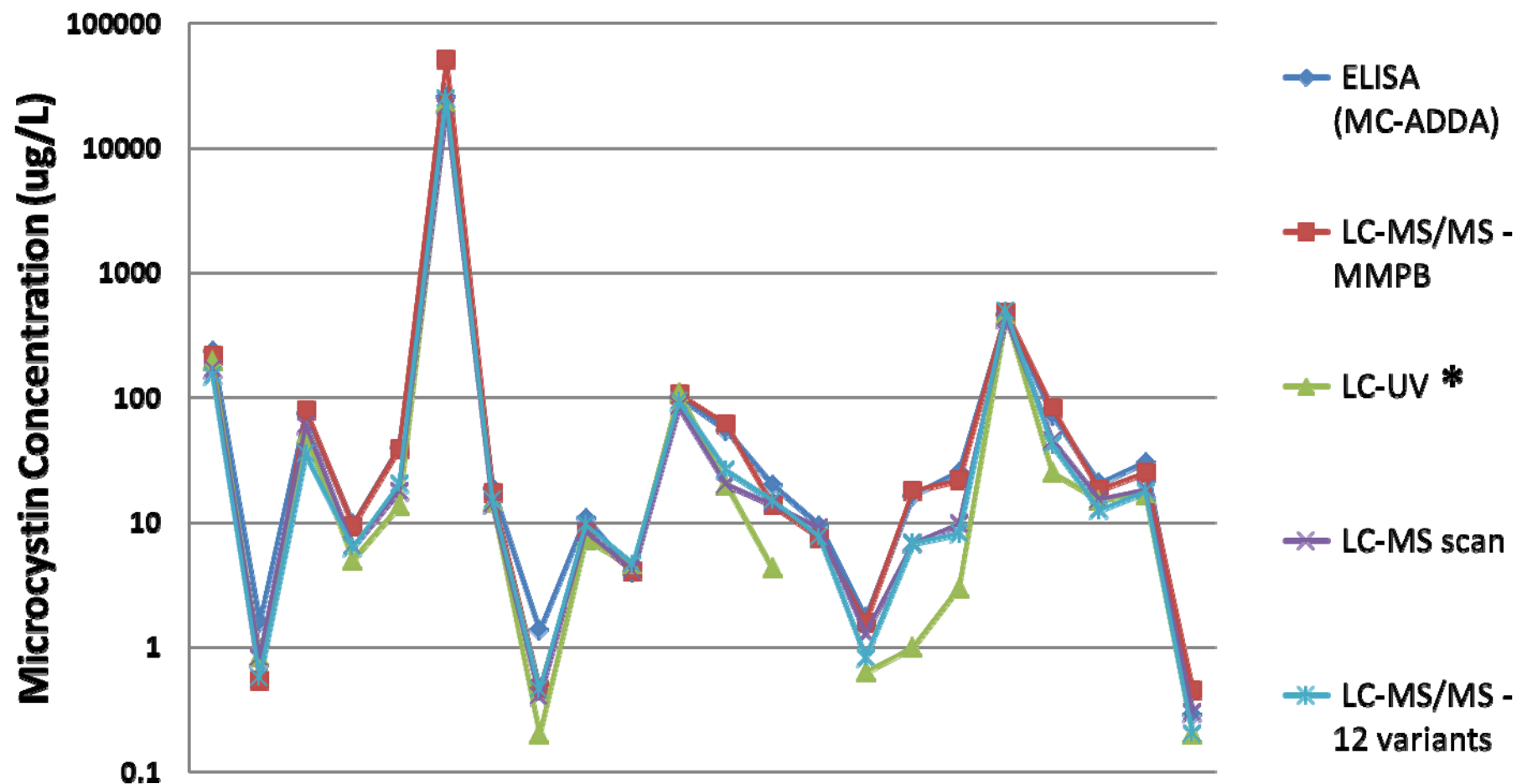
- 1 - D-Alanine
- 2 - Variable L-amino acid
- 3 - D-Methylaspartic acid
- 4 - Variable L-amino acid
- 5 - 3-amino-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoic acid (Adda)
- 6 - D-Glutamic acid
- 7 - N-Methyldehydroalanine

- Over 140 Microcystin Variants
- Standards Not Available for Majority

Analytical Method Comparison & Microcystin Variant Evaluation

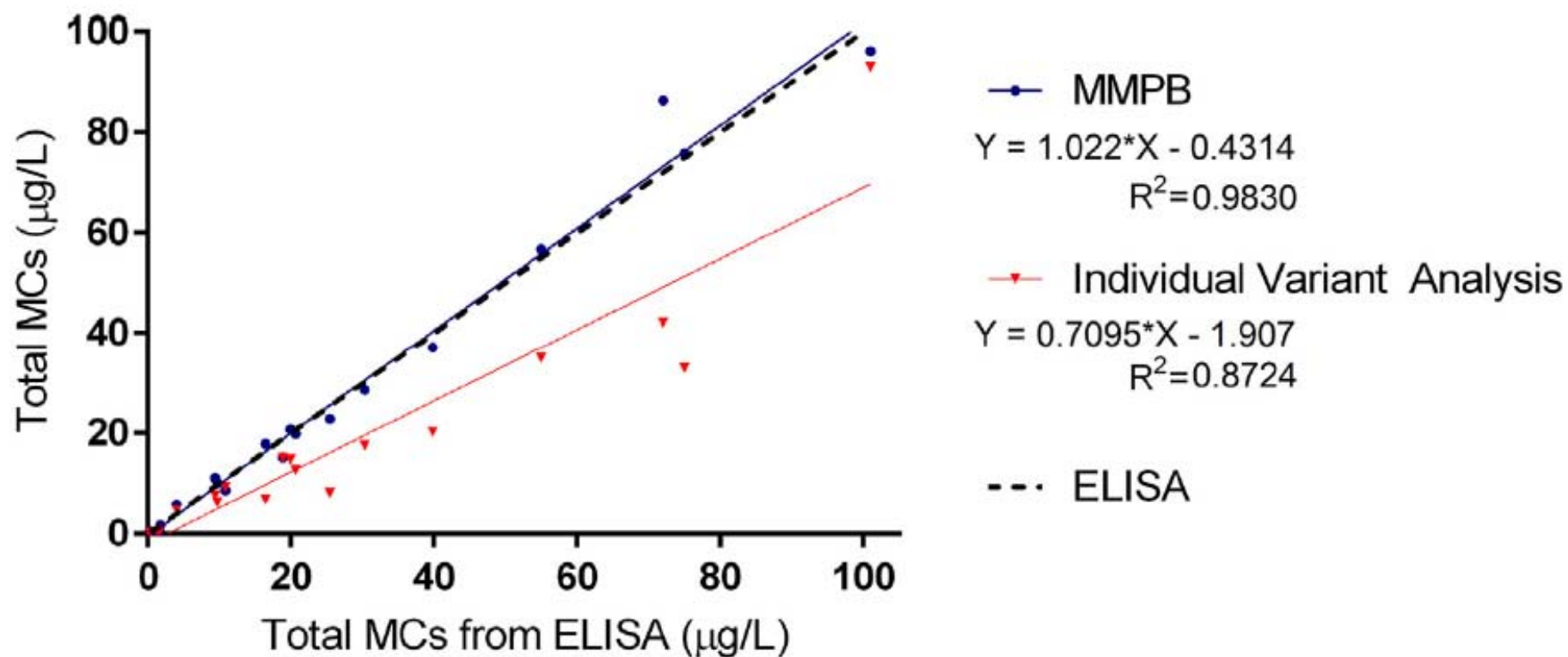
- 11 Sites: 4 up-ground reservoirs, 2 in-stream reservoirs, 2 Lake Erie locations, 2 canal-feeder lakes, and 1 river source
- 22 samples from 2014 selected to help evaluate spatial and temporal variability within source waters
- Variety of cyanobacteria genera represented
- Each sample analyzed using 5 separate analytical methods ELISA-MC-ADDA, LC-UV/PDA, LC-MS/MS – individual variant (12), LC-MS/MS-MMPB, LC-MS scan

Results of Method Comparison



* LC-UV data presented do not include false-positives that were eliminated from total (Based on lack of confirmation with LC-MS methods). Sample #14 was non-detect for MCs using LC-UV.

Results of LC-MS/MS MMPB and Individual Variant Analysis Compared to ELISA



Microcystins Methods - Key Findings

- 16 different MC-variants detected
- MC-LR was only detected at 5 of 11 sites (45%)
- Most commonly detected variants were: MC-YR, [Dha7] MC-LR and [DAsp3] MC-RR
- LC-PDA methods prone to interference, potential for false positives and false negatives
- LC-MS/MS MMPB method confirmed ELISA results (raw water)
- 91%-100% of samples had MC-variants not detectable by U.S. EPA Method 544 (including dominant MC-variant in some samples)
- LC-MS/MS individual variant analysis under-reported total microcystins, based on MMPB, LC-UV/MS scans, and ELISA data
- No perfect method for TOTAL Microcystins

Analytical Methods Utilized by Ohio EPA

	Microcystins ($\mu\text{g/L}$)	Cylindro- spermopsin ($\mu\text{g/L}$)	Saxitoxins ($\mu\text{g/L}$)	Anatoxin-a ($\mu\text{g/L}$)
Surveillance sampling	ELISA (MC-ADDA)	ELISA	ELISA	LC-MS/MS
Repeat sampling in response to a finished water detection	ELISA (MC-ADDA)	LC-MS/MS	LC-MS/MS	LC-MS/MS

ELISA: Enzyme-Linked Immunosorbent Assay

LC-MS/MS: Liquid Chromatography followed by tandem
Mass Spectrometry

Ohio HAB Initiatives & Resources

PWS Funding: \$1.5 million in grants for analytical kits, microscopes, and raw water quality sensors and \$100 million in zero-interest loans for drinking water infrastructure improvements including additional treatment, alternative sources and storage.

Technical Assistance: Site visits with all susceptible surface water systems on treatment optimization and contingency planning. Numerous presentations, webinars and workshops have been held with PWSs, stakeholders, and partners.

Guidance: Developed Cyanotoxin Treatment White Paper, Cyanotoxin Treatment Optimization Protocol, Cyanotoxin General Plan, Algaecide Application Fact Sheet, Distribution Cyanotoxin Monitoring; ELISA SOPs, Response Strategies; Website.

Emergency Response: Established PWS-specific HAB incident action plans and coordinated with partner agencies during multiple table top exercises and drills.

Clean Water Act Integration: Established cyanotoxin indicators of impairment for determining public drinking water supply beneficial use attainment

Research: Partnered with university and government researchers to address data gaps.

www.epa.ohio.gov/ddagw/HAB.aspx



Summary

- HABs are Not Always Visually Apparent
- HABs Can Occur Year-Round, in a Variety of Source Waters, and can be Different (or similar) Every Year
- Early Warning Monitoring Helps
- Methods Matter
- Saxitoxins are an Emerging Issue
- Plan Ahead!

Questions?

www.epa.ohio.gov/ddagw/HAB.aspx

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