

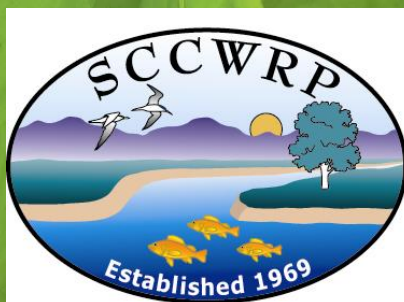
US EPA ARCHIVE DOCUMENT

For assistance in accessing this document please send  
an email to [EPACyanoHABs@epa.gov](mailto:EPACyanoHABs@epa.gov)

# New HAB Monitoring and Assessment Techniques and Tools

Meredith Howard

Southern California Coastal Water Research Project



# Measurements to Meet Monitoring Objectives

- Which potential toxin producing organisms/species are in the waterbody?
  - Microscopy, molecular tools (DNA Barcoding, qPCR), imaging instruments
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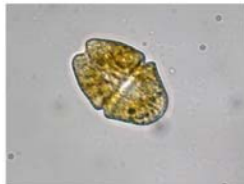
# Microscopy

- Traditional 'gold standard' method for quantification
- Cell counts are time consuming and expensive
- Cryptic or rare species can be missed
- Alternative to formal cell counts: Relative abundance

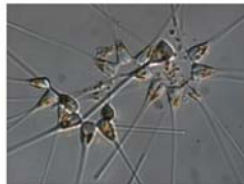
## Phytoplankton Identification

HOME | PHYTOPLANKTON | GLOSSARY

Search by image



Marine Diatoms



Marine Dinoflagellates



Freshwater

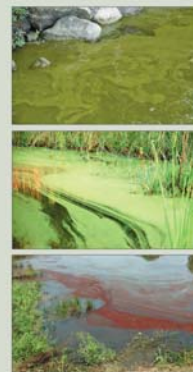
Phytoplankton ID Gallery

<http://oceandatacenter.ucsc.edu/home/>

Kudela Lab, UC Santa Cruz



## Field and Laboratory Guide to Freshwater Cyanobacteria Harmful Algal Blooms for Native American and Alaska Native Communities



Open-File Report 2015-1164

U.S. Department of the Interior  
U.S. Geological Survey

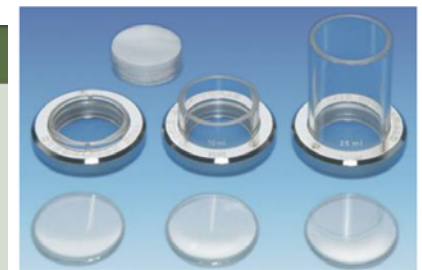






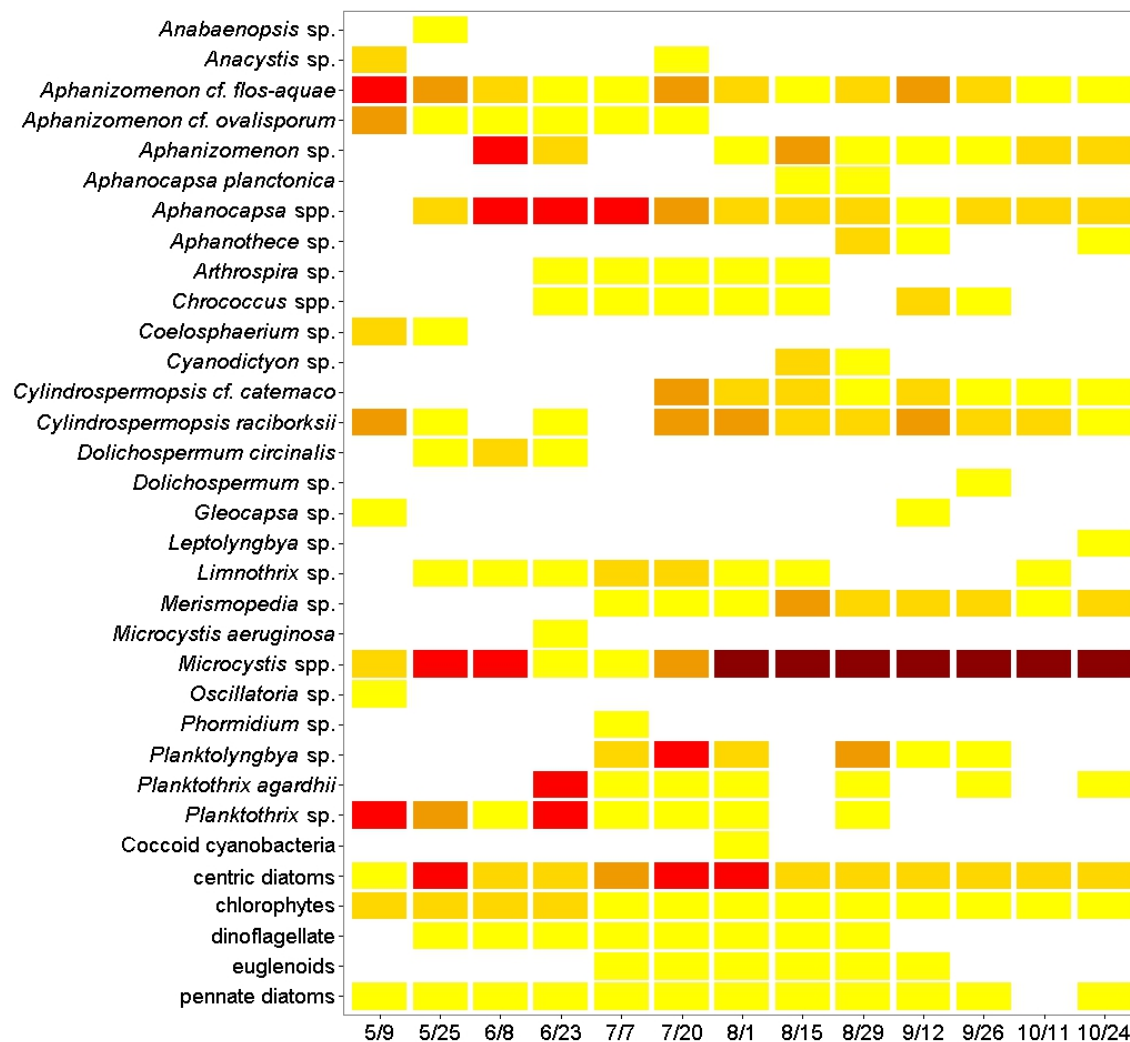


Figure 12: Utermöhl counting chamber



Tatters et al., 2017

	Rare	<1%
	Present	1-<10%
	Common	10-<25%
	Abundant	25-<50%
	Dominant	50-100%
	None Observed	






Nostoc - 200x — iPhone  
Field Scope

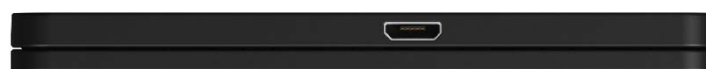


# Field Microscopes

 **μPeek**



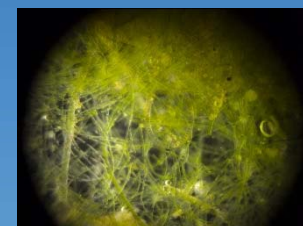
**microPeek:** The innovative mobile technology that turns every smartphone into a professional microscope.



μPeek + Slide Holder



Apple iPhone 5



**SCRONA**



brightfield microscopy



darkfield microscopy



fluorescence microscopy<sup>3</sup>



# FlowCAM

- Semi-automated imaging flow cytometer
- Identification and biovolume estimation



# Imaging FlowCytobot (IFCB): *in situ* Automated Microscope

- *In situ* submersible flow cytometer that generates images of particles
  - Images classified to identification, abundance, biovolume

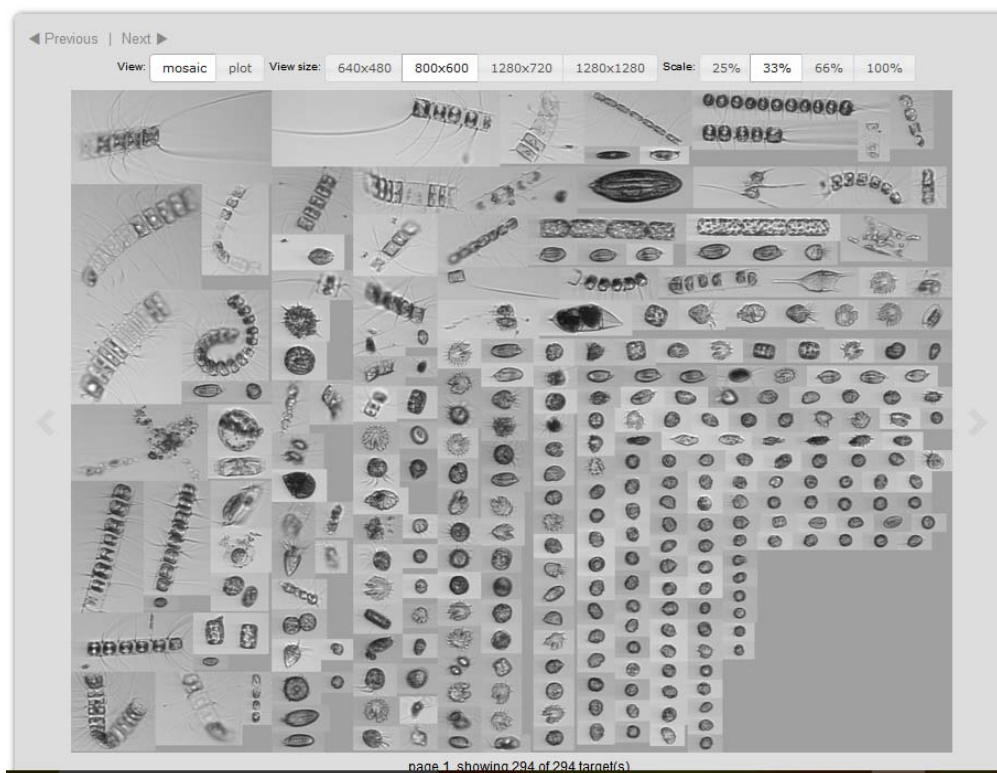


Image courtesy of Raphael Kudela, Santa Cruz Wharf  
<http://oceandatacenter.ucsc.edu/home/>



# Quantitative Polymerase Chain Reaction (qPCR)

- Identify and quantify taxa
  - Rapid, sensitive and specific approach for taxa identification
- Detection and quantification of toxin producing genes

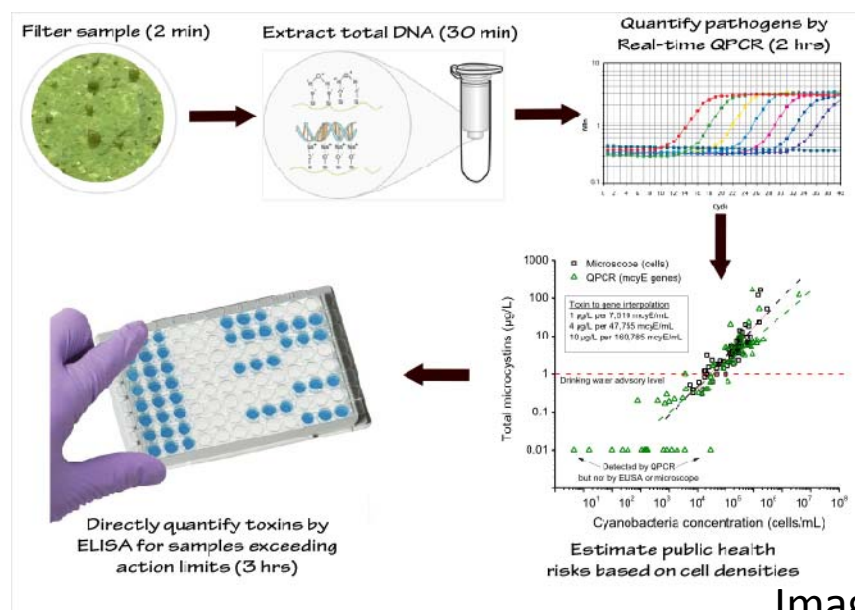
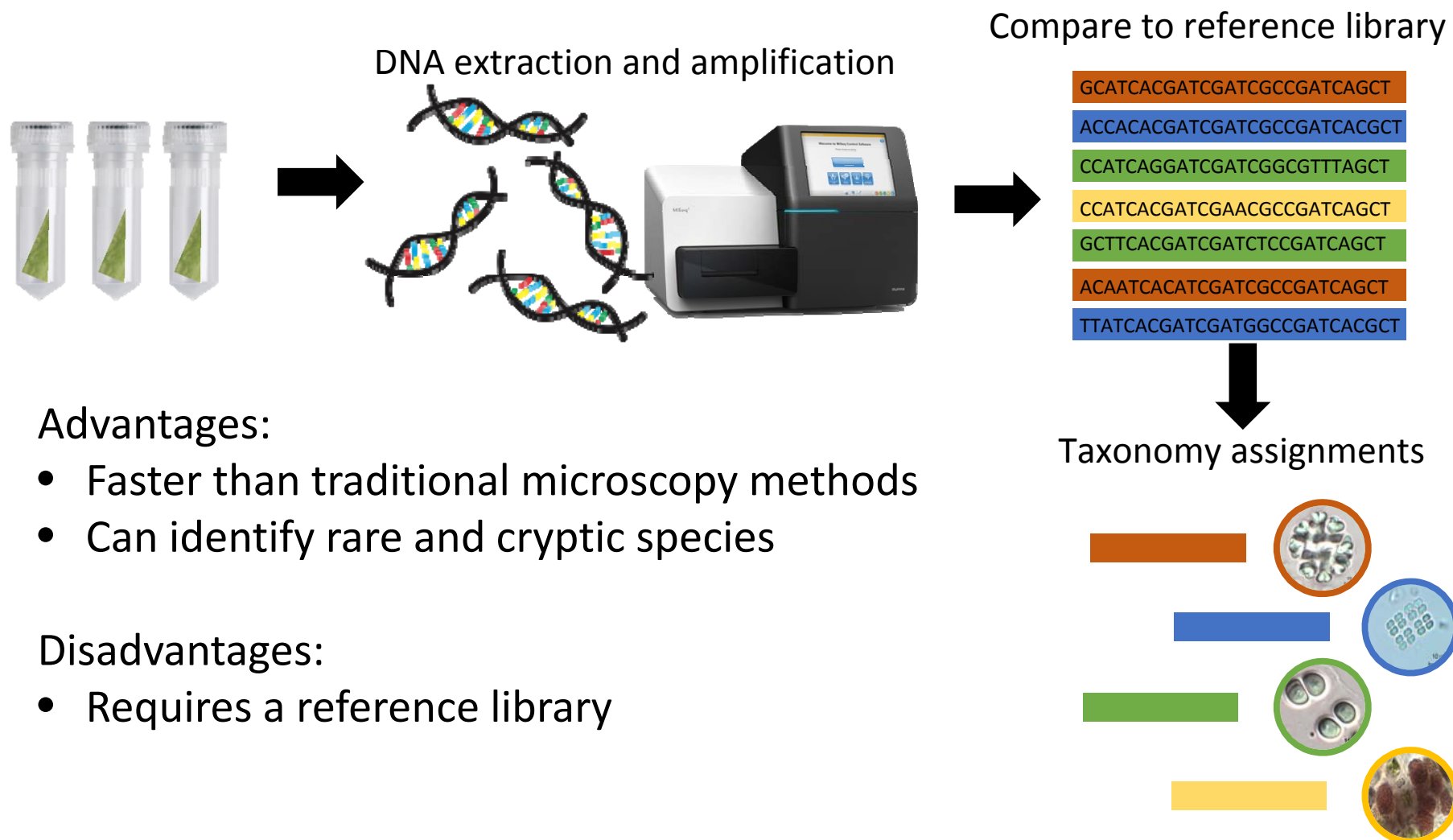
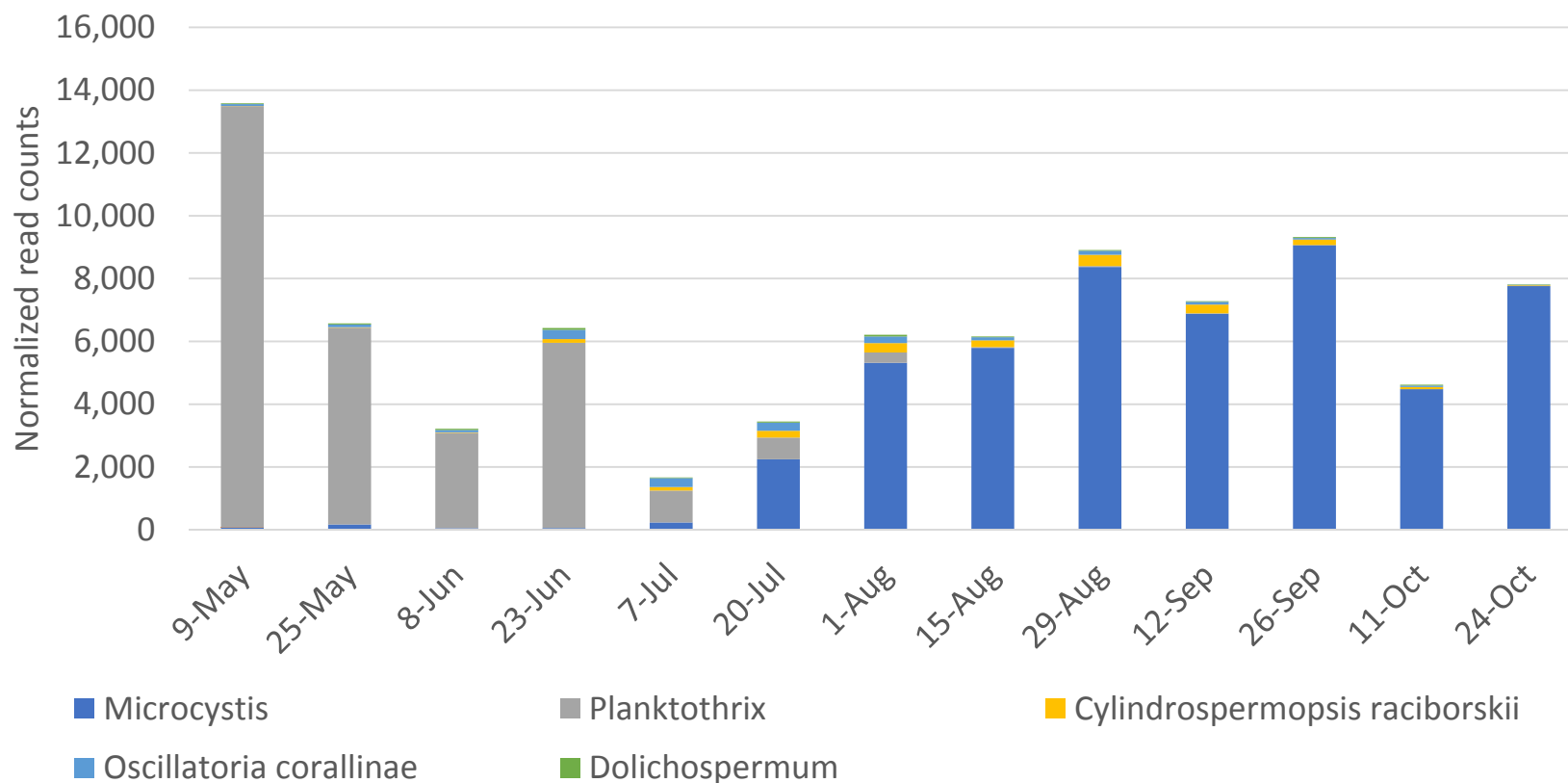


Image courtesy of Bend Genetics

# DNA Barcoding: DNA sequencing to identify algal species



# Potential toxin producers from Lake Elsinore



Theroux and Howard, unpublished data

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# Field Tests for Cyanotoxins



- Envirologix QualiTube
  - Microcystins



- Beacon Analytical Systems Tube Kit
  - Microcystins



- Abraxis strip tests ('dipsticks')
  - Anatoxin-a, cylindrospermopsin, microcystins



# Monitoring Tool: Solid Phase Adsorption Toxin Tracking (SPATT)

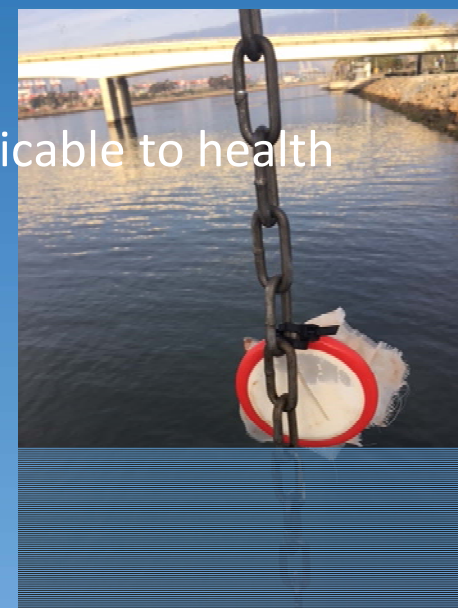
*Passive sampler that is time-integrative*

- Applicable in all waterbody types (marine, brackish, freshwater)
- Provides continuous toxin detection to capture ephemeral events that discrete samples can miss
- Low cost, simple and easy to deploy/recover



*Disadvantage:*

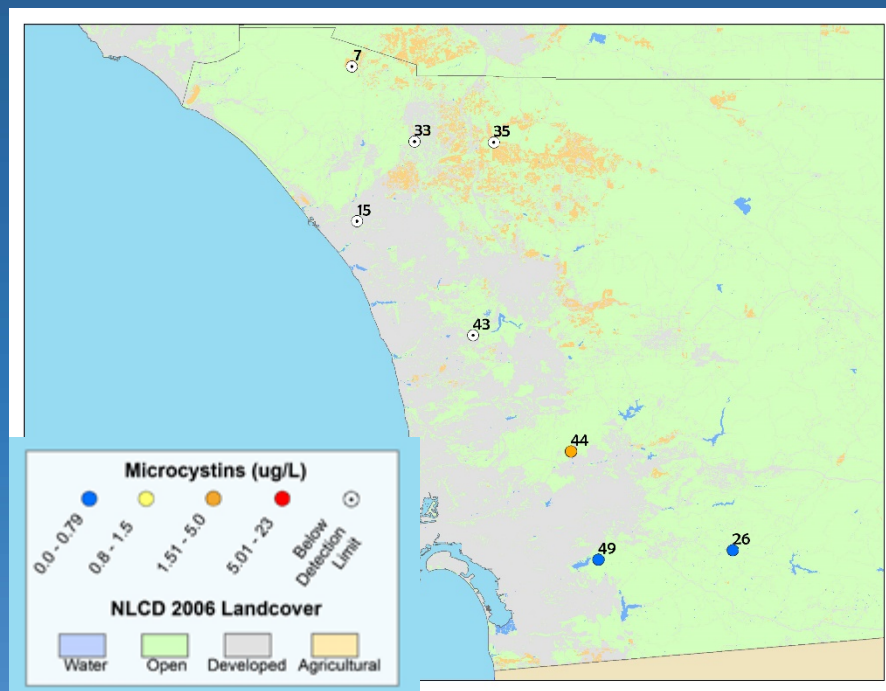
- SPATT will not provide a concentration of toxin that is applicable to health advisory thresholds (ng/g)
- Only measures dissolved toxins not total toxins



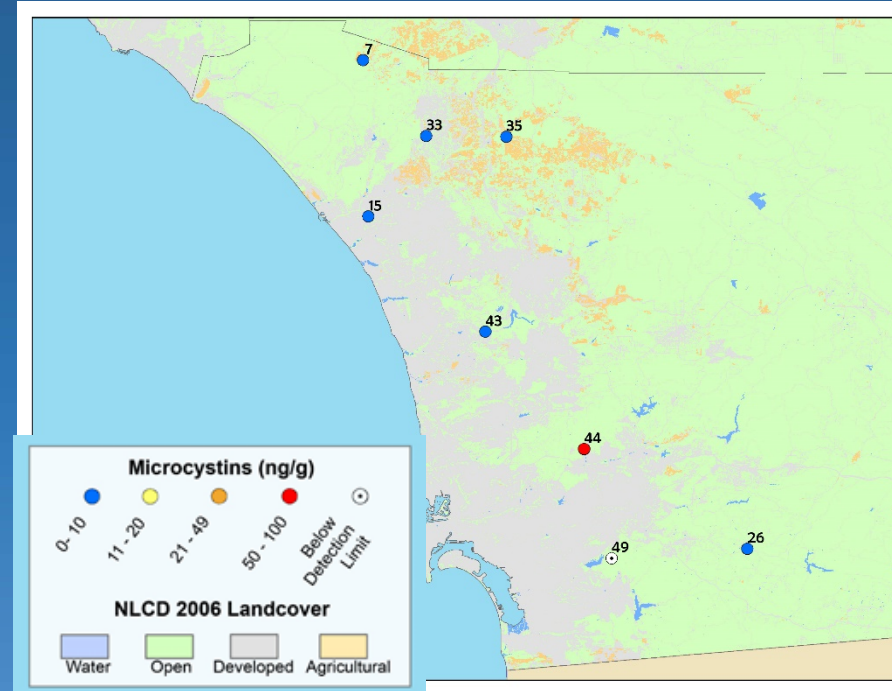
MacKenzie, 2010, Lane et al., 2010, Kudela, 2011

# Microcystin Prevalence Underestimated From Grab Samples By ~50%

Grab Samples



SPATT Samples



## % of Toxic Sites: Depressional Wetlands

Grab Samples	29%
SPATT Samples	83%

Howard et al., submitted







# Common Toxin Analytical Techniques

**Table 3.** Relative advantages and disadvantages of common analytical techniques utilized for analysis of cyanobacterial toxins and taste-and-odor compounds.

Analytical techniques	Advantages	Disadvantages
<b>Bioassays</b>		
Enzyme-Linked Immunosorbent Assay (ELISA)	Relatively easy to use	Data interpretation can be difficult
Inhibition Assays	Cost per analyses lowest of all techniques	Inhibition assays and radioassays not always available
Radioassays	Can be useful as screening tools	Bioassays frequently possess some reactivity towards compounds other than the intended target
	Can indicate toxicity in some cases	Radioassays require permits to work with radioisotopes
		Research objectives may require a chromatographic technique for compound specific quantitation
<b>Gas Chromatography (GC)</b>		
Flame Ionization Detector (GC/FID)	Compound specific	Toxins will most likely require derivitization
Mass Spectrometry (GC/MS)	Cost per analyses intermediate	Not all compounds amenable to derivitization
	Compound identification by GC/MS is superior to GC/FID	GC/FID may require further confirmation
		Sample concentration techniques may be necessary
<b>Liquid Chromatography (LC)</b>		
Ultraviolet-Visible (LC/UV-Vis)	Derivitization typically not necessary	Matrix effects can be substantial
Fluorescence (LC/Fluorescence)	Compound specific	Cost per sample most expensive
Mass Spectrometry (LC/MS)	Greatest number of toxins are amenable to LC techniques	Spectroscopic techniques may require further confirmation
Tandem Mass Spectrometry (LC/MS/MS)	Cost per analyte can be lowest in a multi-analyte method	Sample concentration techniques may be necessary
Ion Trap Mass Spectrometry (LC/ITMS)	Compound identification is superior by LC/MS/MS or LC/ITMS	



## Guidelines for Design and Sampling for Cyanobacterial Toxin and Taste-and-Odor Studies in Lakes and Reservoirs



Scientific Investigations Report 2008-5038

U.S. Department of the Interior  
U.S. Geological Survey

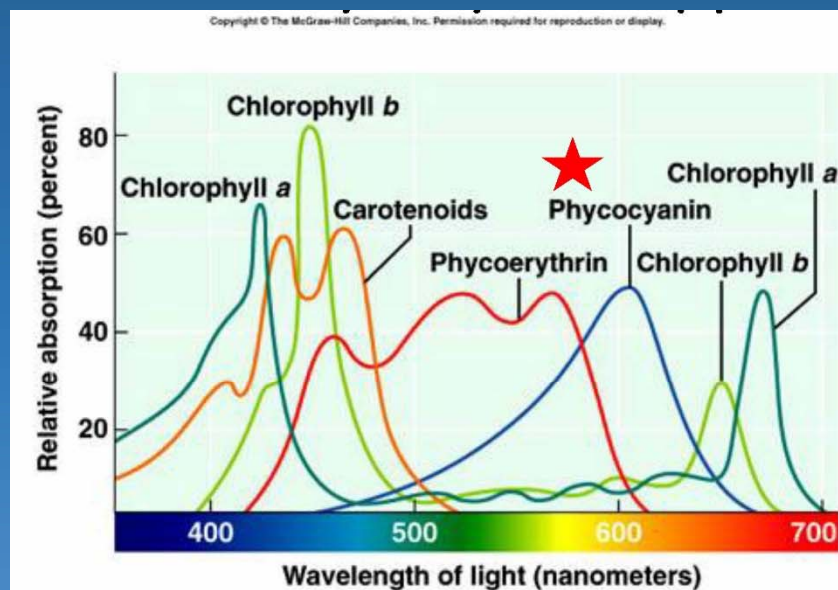
Graham et al. 2008 USGS report

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# Common techniques for chlorophyll and phycocyanin quantification



Turner Designs Trilogy



Orange module  
630nm excitation  
660nm emission



Field sonde  
Hydrolab minisonde



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# Aerial Cameras and Drones



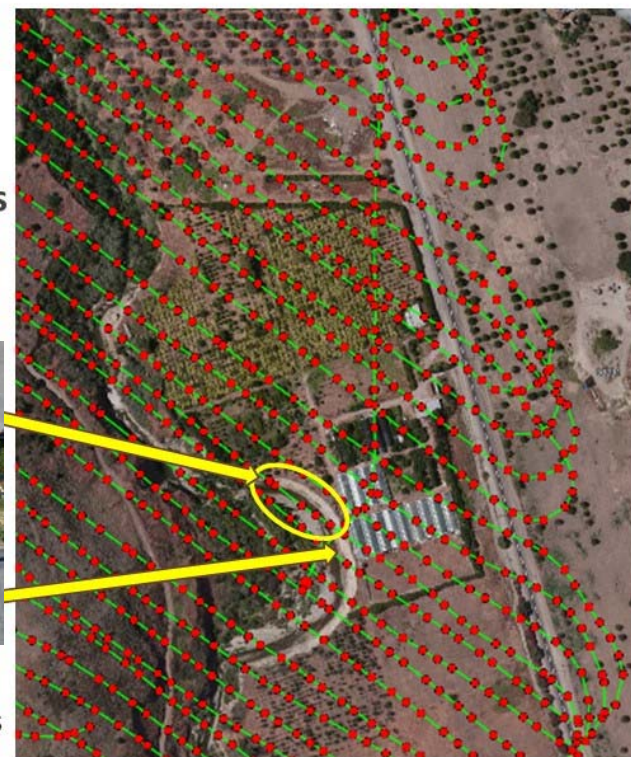
Iron Gate Reservoir, October 2016  
Photo courtesy of Susan Fricke



**Traditional camera sensors**  
worked well (RGB, RGB-IR)



Flight path with images centers



Steinberg, unpublished data<sup>9</sup>





# ECOMAPPER™

Autonomous Underwater Vehicle



The data provide a three-dimensional map of the various water quality parameters.

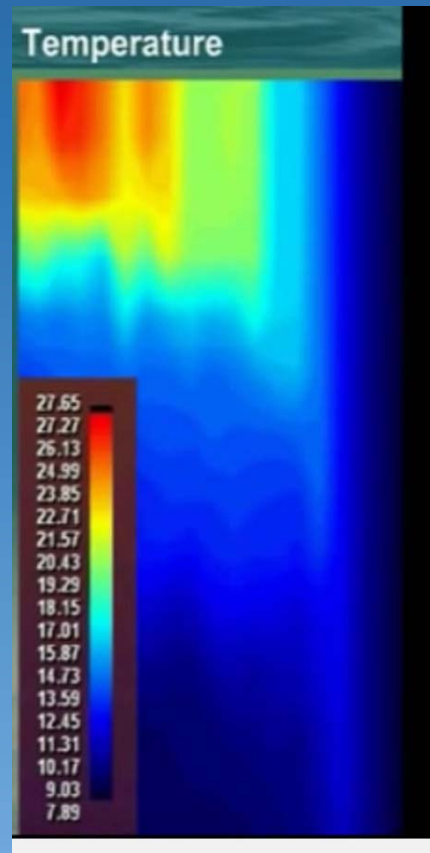
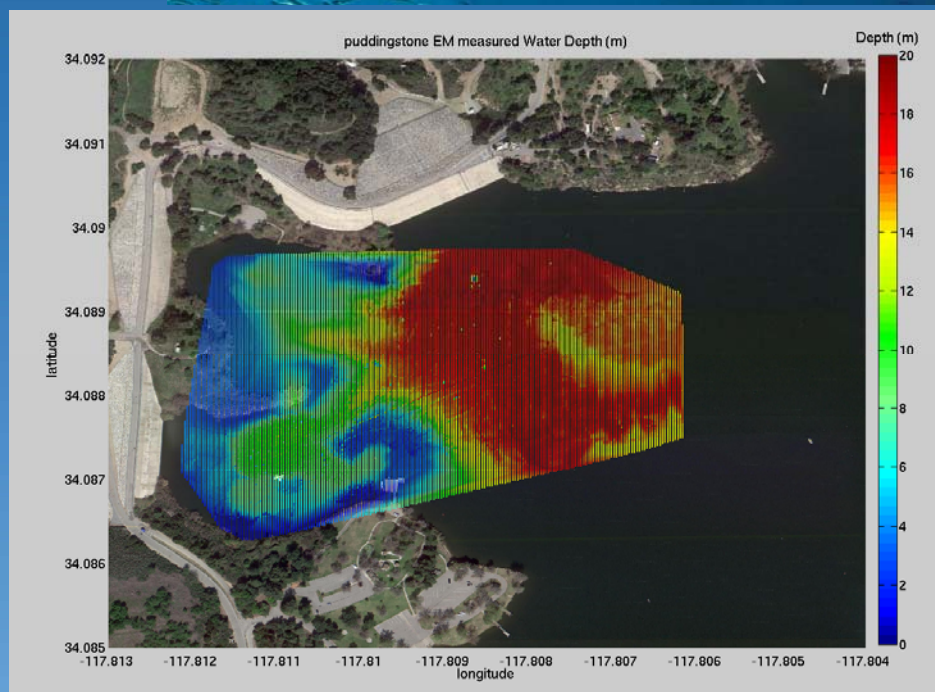


Figure courtesy of Stephanie Kemna, D. Caron & RESL at University of Southern CA

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# Models to Forecast Freshwater Blooms and Risk Levels

**USGS**  
science for a changing world

**Kansas Real-Time Water Quality**

Home View Data Methods Constituents Models Bibliography Links

NRTWQ Home >> Kansas >> View Data >> 07144790

Plot Site Info **Model Info**

USGS station: 07144790 Cheney Reservoir near Cheney, KS

Go to NWISWeb

Constituent: Computed probability of microcystin concentration hourly < Go >

Model Form: 1. 2013-01-01 - Present

Computed probability of microcystin based on total chlorophyll

Stone, M.L., Graham, J.L., and Gatotho, J.W., 2013.  
<http://pubs.usgs.gov/of/2013/1123/>.

Model Developed By Alan Wilson  
 Auburn University, Alabama

<http://wilsonlab.com/forecasting.html>

SECCHI DEPTH (meters) 0.15  
(1 ft = 0.305m)

ALGAL BLOOM TYPES

	Phytoplankton chlorophyll (µg/L)	Cyanobacteria phycocyanin (µg/L)	Toxic cyanobacteria microcystin (µg/L)
PREDICTED CONCENTRATIONS	99	328	0.01864
RISK LEVEL	High Risk	High Risk	High Risk

**PHYTOPLANKTON AND CYANOBACTERIAL FORECASTING MODELS**

ALGAL BLOOM TYPES

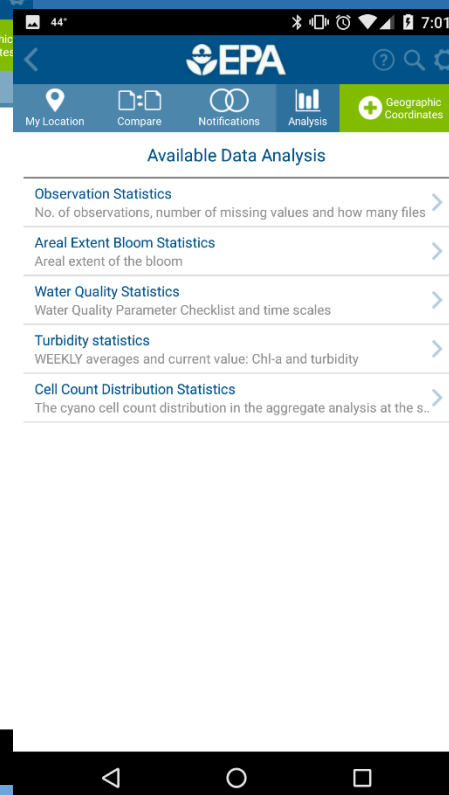
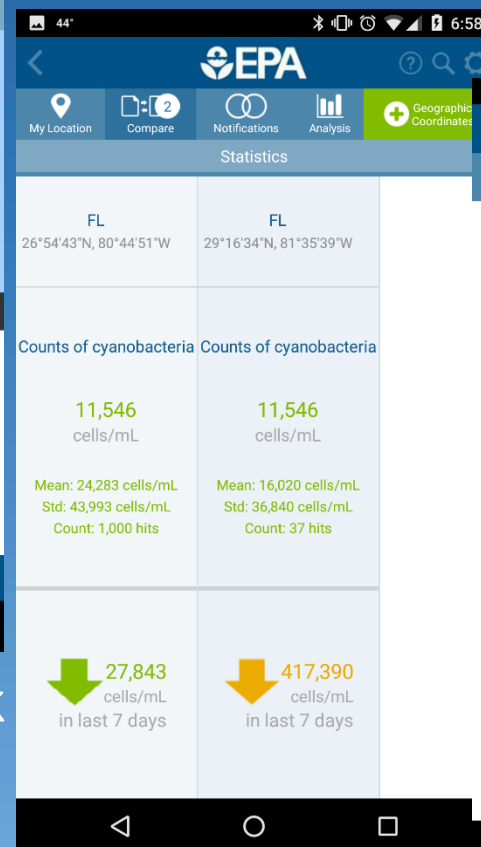
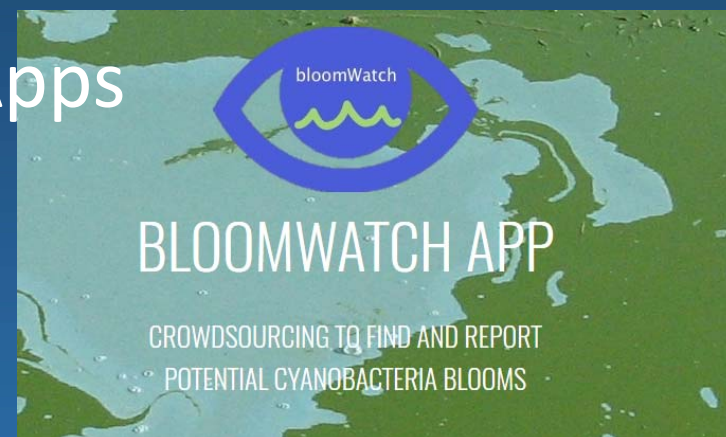
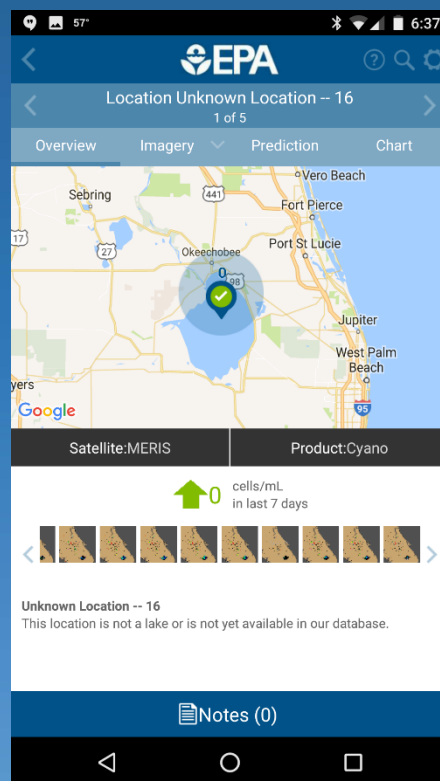
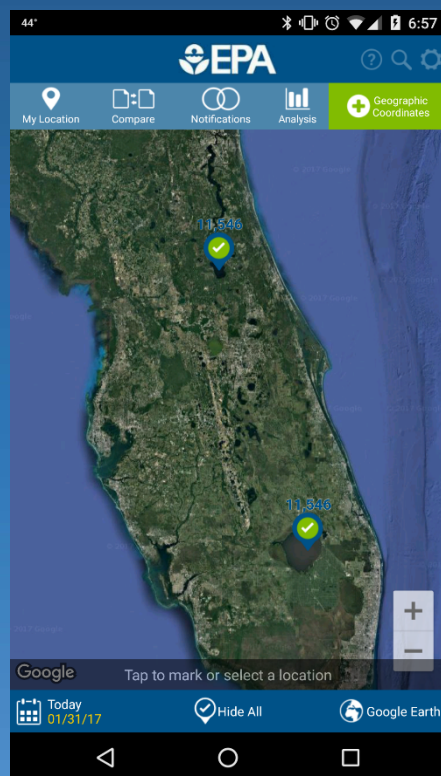
WATER QUALITY DATA	Phytoplankton	Cyanobacteria	Toxic cyanobacteria
Chlorophyll (CHL) µg/L		200	
Total Phosphorus (TP) µg/L			
Soluble Reactive Phosphorus (SRP) µg/L			
Total Nitrogen (TN) µg/L			
Nitrogen:Phosphorus (N:P) molar			

chlorophyll (µg/L) phycocyanin (µg/L) microcystin (µg/L)

PREDICTED CONCENTRATIONS

RISK LEVEL

# Smartphone Bloom Reporting Apps



Cyanobacteria Assessment Network (CyAN) Mobile App

Questions?

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