

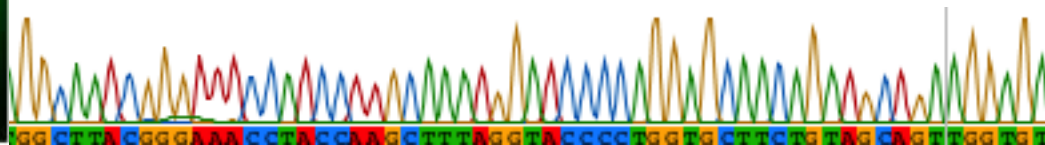
US EPA ARCHIVE DOCUMENT

A vertical strip on the left side of the slide shows several bright green, spiral-shaped structures, likely cyanobacteria, against a black background. These structures are coiled and have a glowing, filamentous appearance.

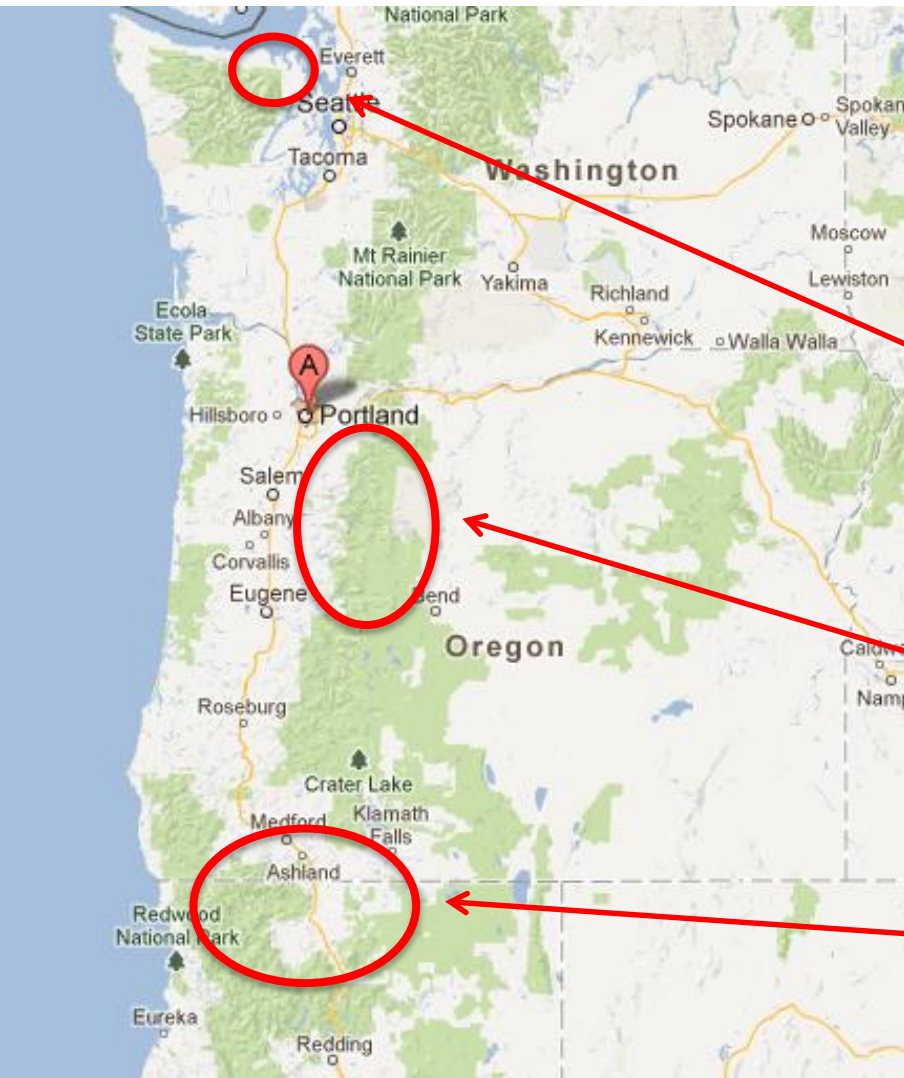
Cyanobacterial occurrence in lakes and rivers in the PNW and the role of eutrophication: Progress through genetic technologies

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Cyanobacterial blooms in the PNW and Northern California



Anderson Lake, Washington

Unusually high levels of anatoxin-a
Toxin producer identified

Willamette Valley reservoirs, Oregon

Potential toxicity in drinking water sources
Toxicity and annual strain successions

Klamath Valley, Oregon/California

Toxic *Microcystis* in reservoirs and river
Genotype trends over 9 years
Long-distance river transport

Anabaena/Aphanizomenon/Dolichospermum with varied toxigenicities: null, *ana+*, *cyr+*, *mcy+*

Anderson Lake, Washington

Anatoxin-a up to 1090 µg/L

Anabaena sp. WA102 *Ana+* culture



↑
Anabaena flos-aquae/
lemmermannii morphology



Cascade Range reservoirs & lakes, Oregon

Potential toxicity in drinking water sources

Dexter Res: Non-toxic

Detroit Res: 7-epi-Cylindrospermopsin @ 195 µg/L

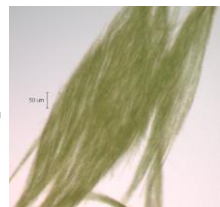
Odell Lake: Microcystin @ 675 µg/L

Upper Klamath Lake, Oregon

Aphanizomenon flos-aquae (AFA)

Harvested for “nutritional supplements”

AFA UKL13 culture



Microcystis and microcystin in the PNW

Washington State

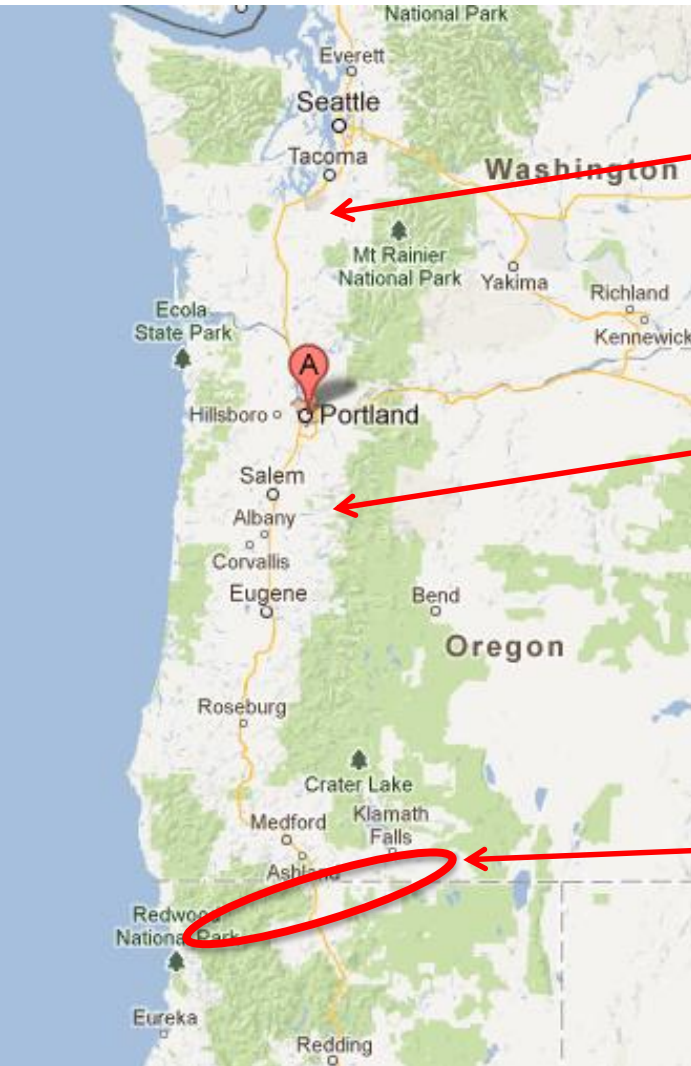
Microcystis and microcystin common in Washington State lakes

Oregon

Toxic *Microcystis* at low altitude W. Oregon, E. Oregon, and Willamette River in downtown Portland

Klamath Valley, Oregon/California

Microcystis in Upper Klamath Lake, Klamath River reservoirs and 300 km of Klamath River



Eutrophication: Trophic State Index and PNW cyanobacterial blooms

Trophic Class	Trophic State Index	Chl-a (µg/L)	TP (µg/L)	Secchi Depth (m)
Oligotrophic	<40	<2.6	<12	>4
Mesotrophic	40 - 50	2.6 - 20	12 - 24	2 - 4
Eutrophic	50 - 70	20 - 56	24 - 96	0.5 - 2
Hypertrophic	>70	>56	>96	<0.5

Lake	Chl-a (µg/L)	TP (µg/L)	TN (µg/L)	Trophic Class
Anderson, WA		40 - 80 (150)	1000 - 1300	Eutrophic
Dexter, OR	20 - 30	50 - 80	500 - 750	Eutrophic
Copco, CA	40 - 200	200	800 - 1500	Hypertrophic
Taihu, China	40 - 80	100 - 250	2000 - 4000	Hypertrophic

Sources: Jefferson Co. Report 2013 (And); Dreher unpub (Dex); Asarian et al. 2009 (Copco); Xu et al. 2015 (Taihu)

CyanoHAB challenges in the PNW

Abatement through reduced nutrient inputs

- But we face elevated P levels from volcanic geology

Abatement by other means,

e.g., flow control, temperature regulation, lower fish stocking

Assessment & monitoring of threats to public & ecological health:

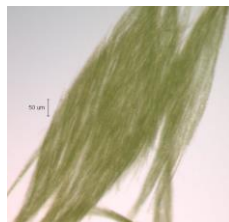
- Drinking water: toxins, taste & odor compounds
 - Recreational exposure, incl. pets (esp. dogs)
 - Food contamination through irrigation
- Toxin transfer to remote sites downriver & bioaccumulation in shellfish
 - Benthic sources of toxins
- Food-web and ecological disruptions; e.g., role as threat to endangered sucker fish in Upper Klamath Lake (OR)
 - Impacts of climate change, esp. increasingly dry summers

Genetic-inspired research is essential in crafting the best management & public health strategies to these challenges

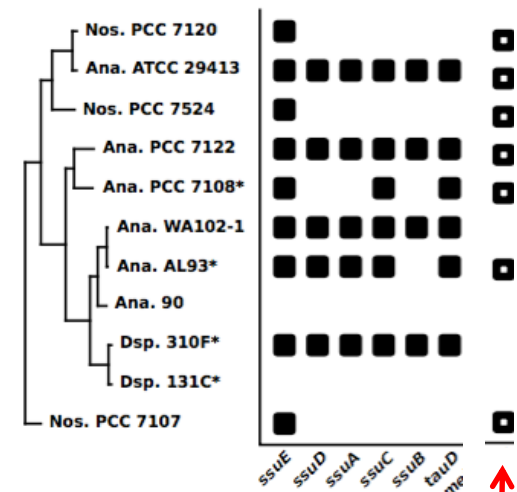
Genome sequencing of CyanoHABs

Genome sequencing (human and gut microbes) has revolutionized medicine
Genome sequencing of freshwater cyanobacteria has similar potential

- ❖ Clarify evolutionary and taxonomic relationships between isolates



- ❖ Reveal pathways relevant to ecological fitness & differences between isolates



Genes for utilization of reduced forms of S



Phycocyanin synthesis

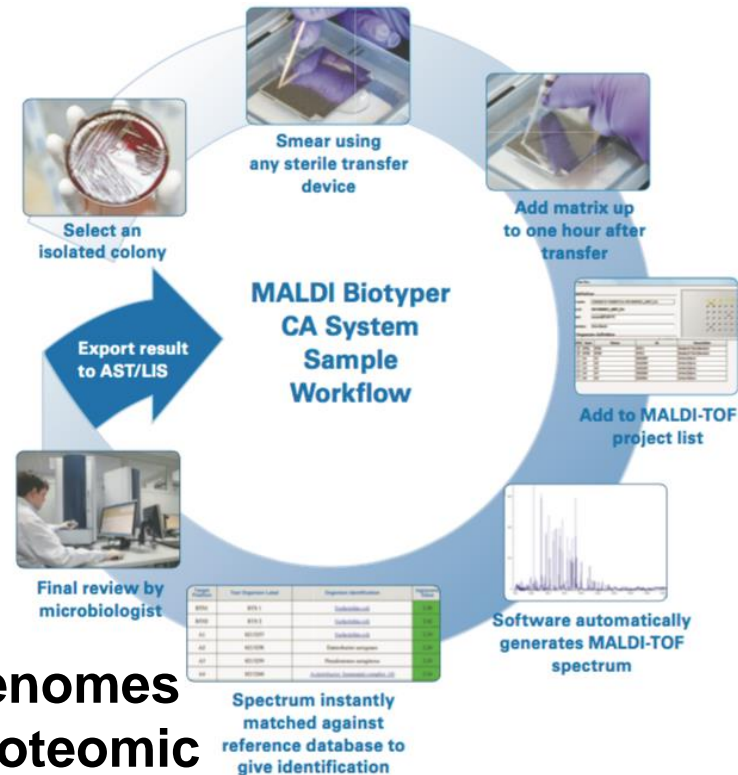
Genome sequencing of CyanoHABs

Genome sequencing (human and gut microbes) has revolutionized medicine
Genome sequencing of freshwater cyanobacteria has similar potential

- ❖ Allow optimal design of primers for PCR-based high-throughput monitoring

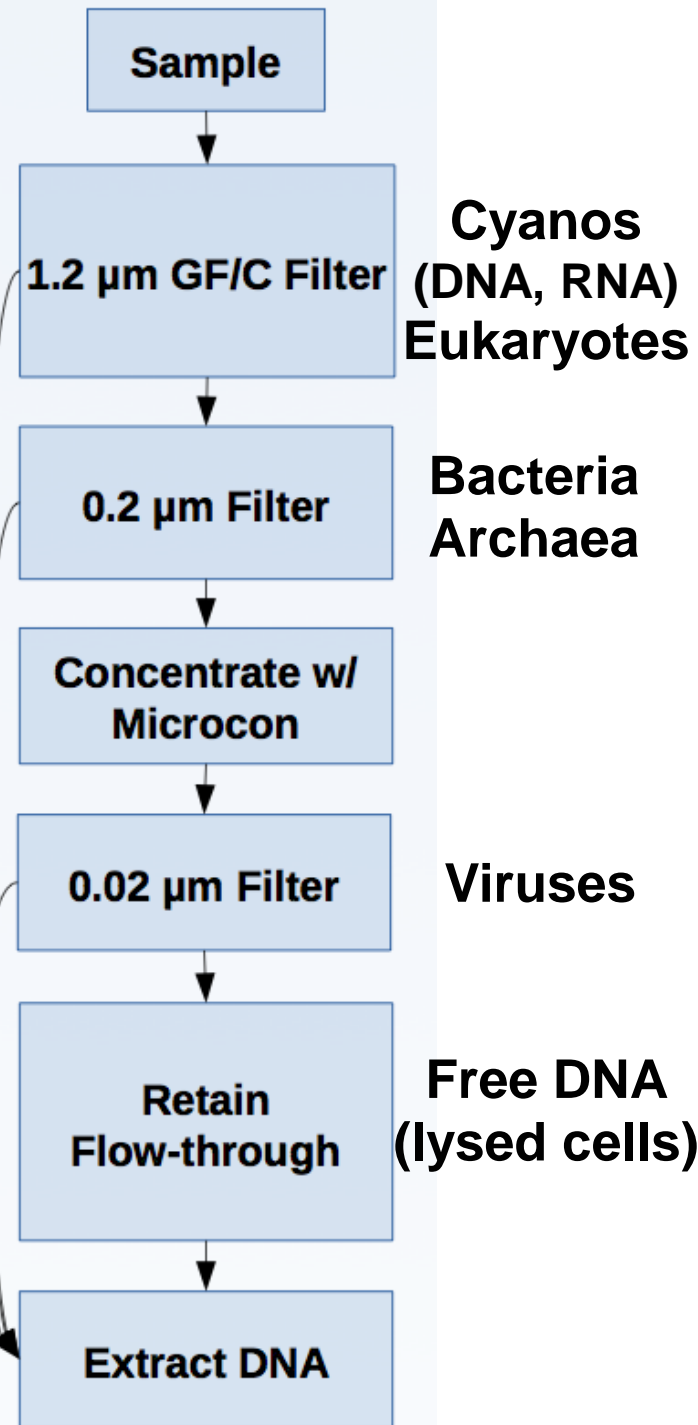
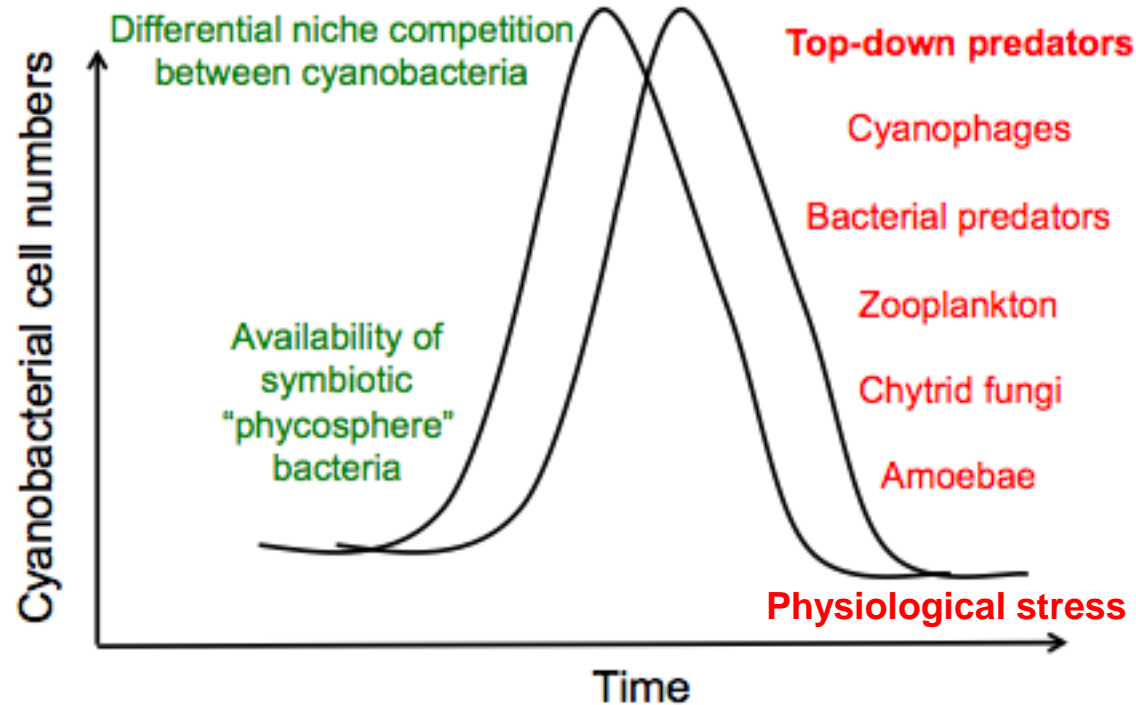
Forward primer (NB78)

<i>Aphanizomenon</i> sp. WA102	GCTTAAATGGTTTGCGCGAA	
Mismatches	A T T	
Primers	GCTTGAACGGCTTGCGCGAA	
Mismatches	A T	
<i>Anabaena</i> sp. WA102	GCTTAAATGGCTTGCGCGAA	



- ❖ Provide reference genomes for possible future proteomic strain identification, as widely used in clinical bacterial ID

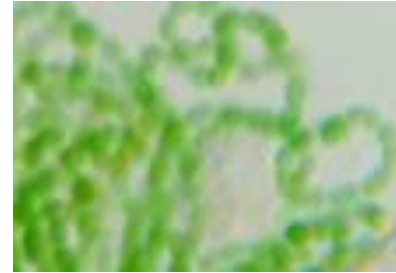
DNA and RNA analysis for determining the biological influences of population dynamics



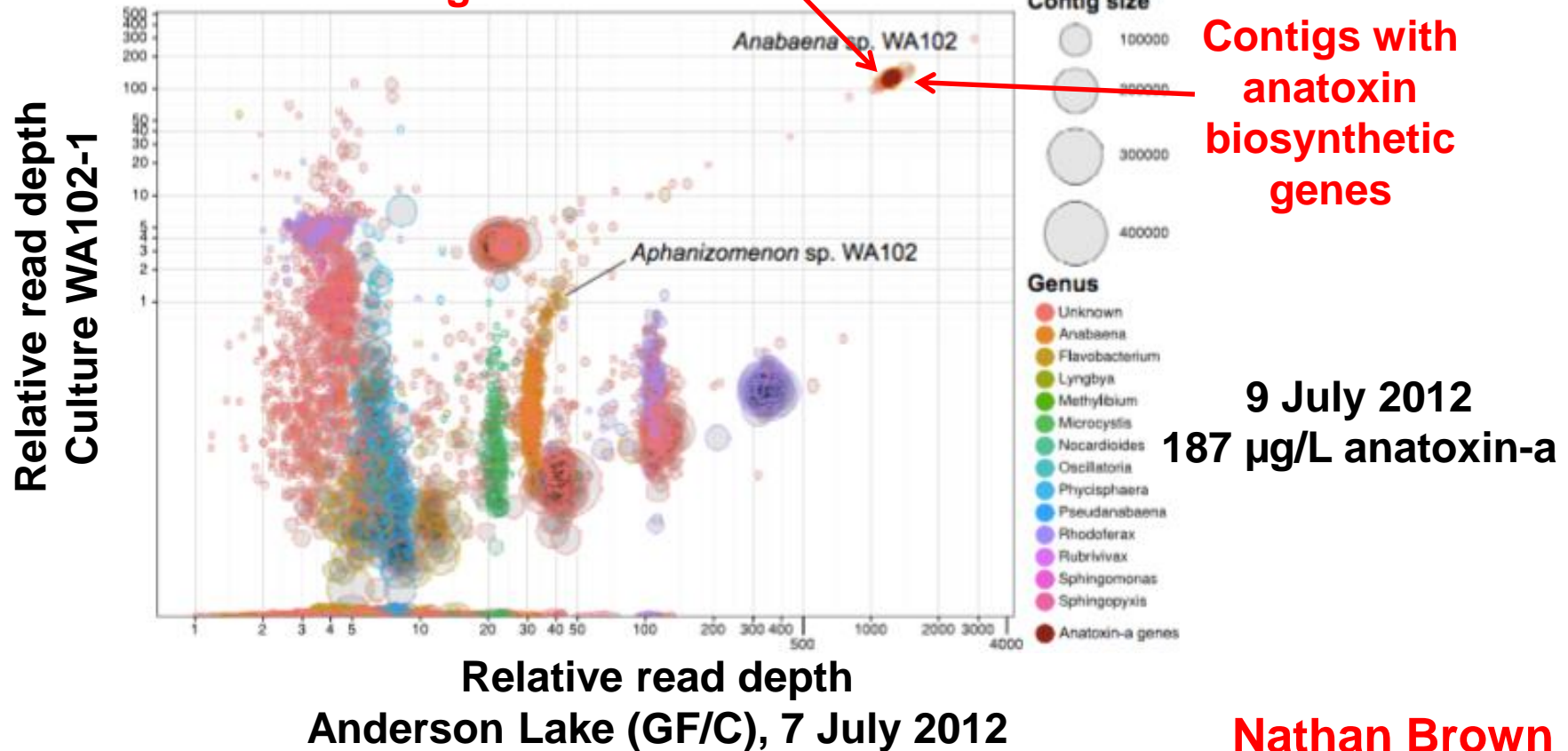
Anderson Lake, WA: Anatoxin-a up to 1090 µg/L

Anabaena sp. WA102 culture; *ana+*
Major anatoxin producer in Anderson Lake

Shotgun metagenome analysis to study all
DNA present in a sample

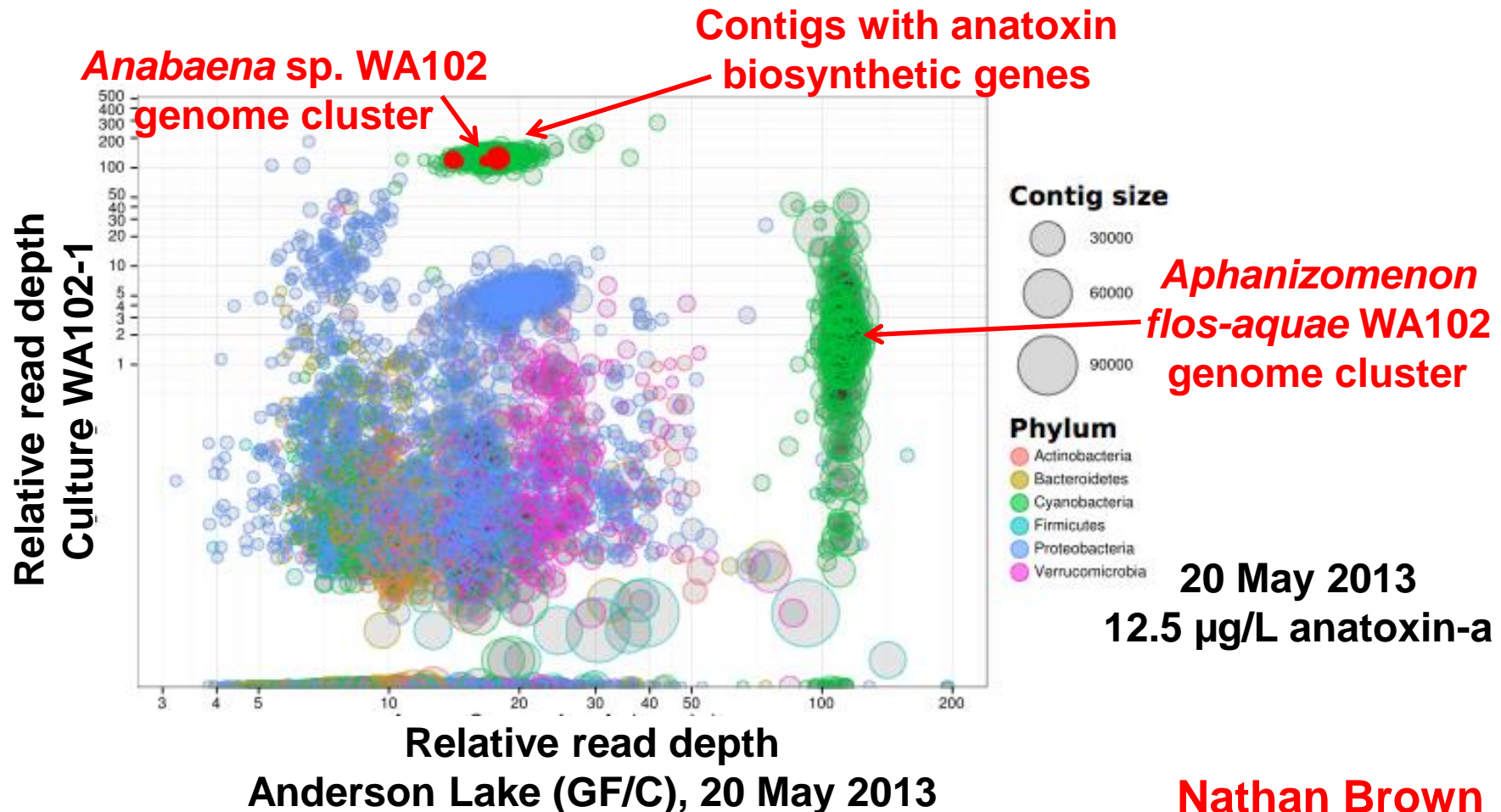


Anabaena sp. WA102
genome cluster

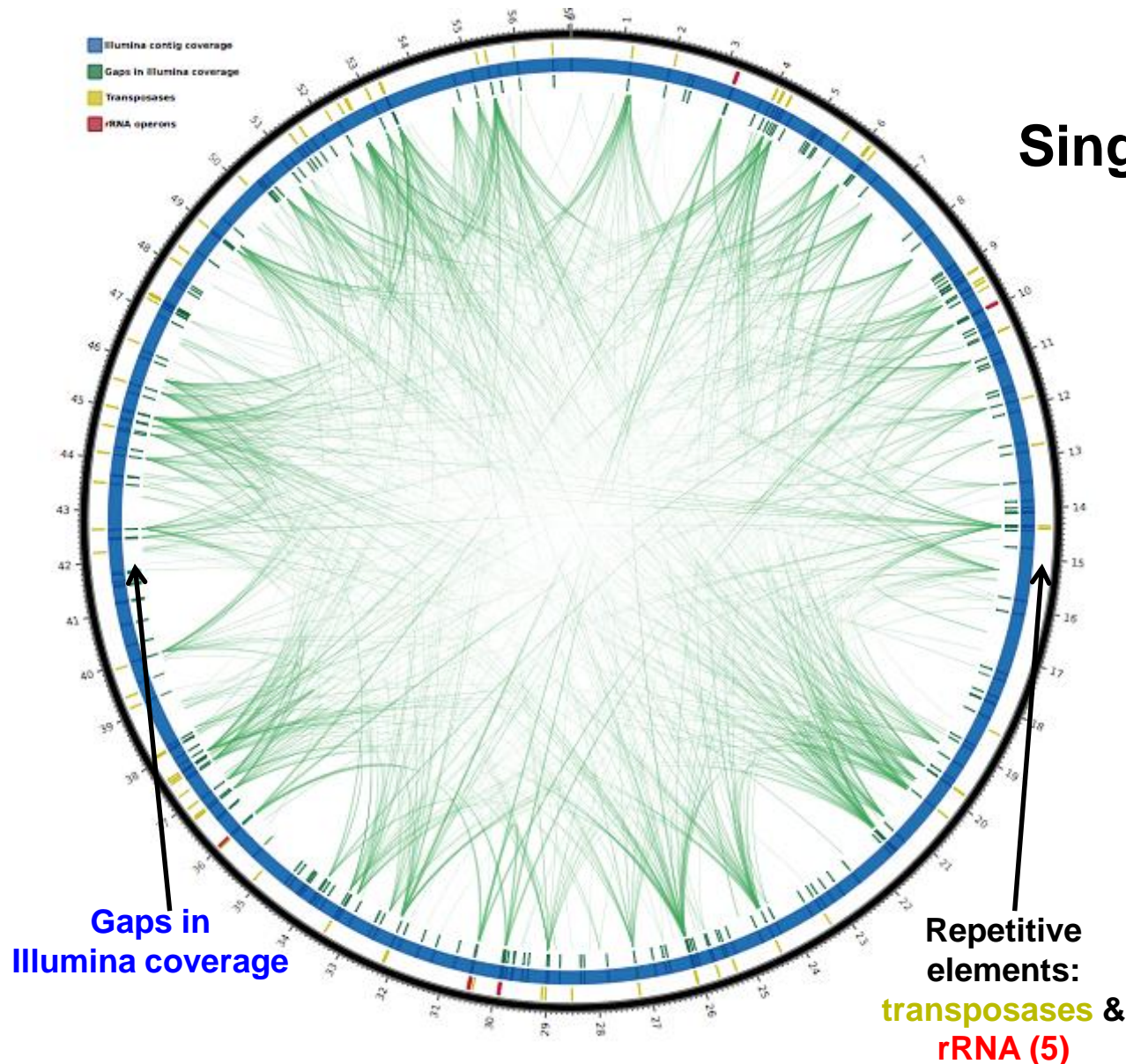


Anderson Lake, WA: Anatoxin-a up to 1090 µg/L

Shotgun metagenome analysis to study all
DNA present in a sample



Anabaena sp. WA102



5.7 Mbp
Single chromosome

Completely assembled
genome PacBio
2 SMRT cells
72x av. Coverage
Av Phred score = 82

Illumina: ~700 contigs

Nathan Brown

Extreme gene order randomization between *Anabaena* sp. WA102 and sp. 90

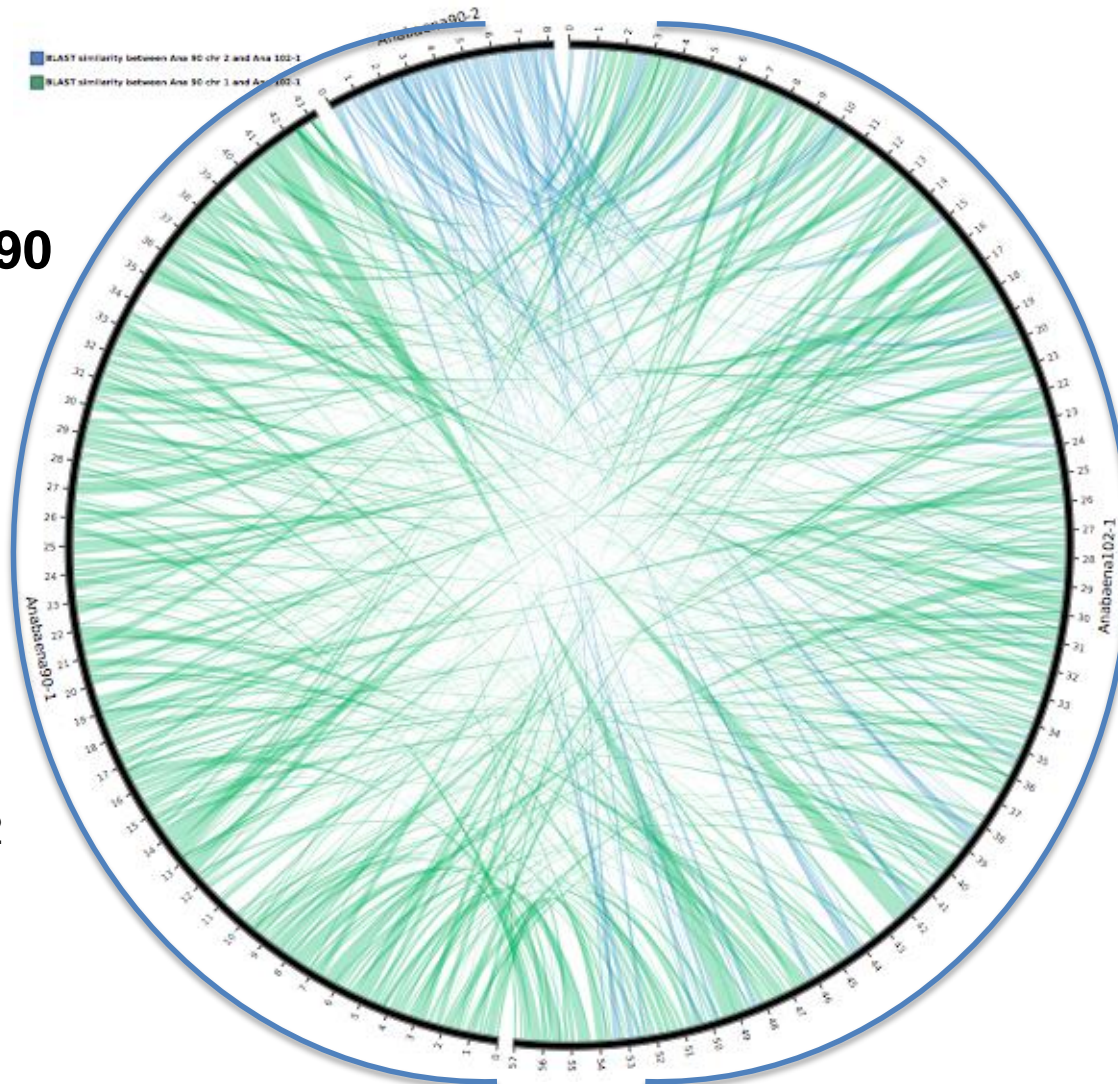
Av. nucleotide identity = 92%

***Anabaena* sp. 90**
Isolated 1986
Baltic Sea

5.1 Mbp
Two
chromosomes
(4.3 + 0.8)
mcy+
Wang et al., 2012

***Anabaena* sp.
WA102**
Isolated 2013

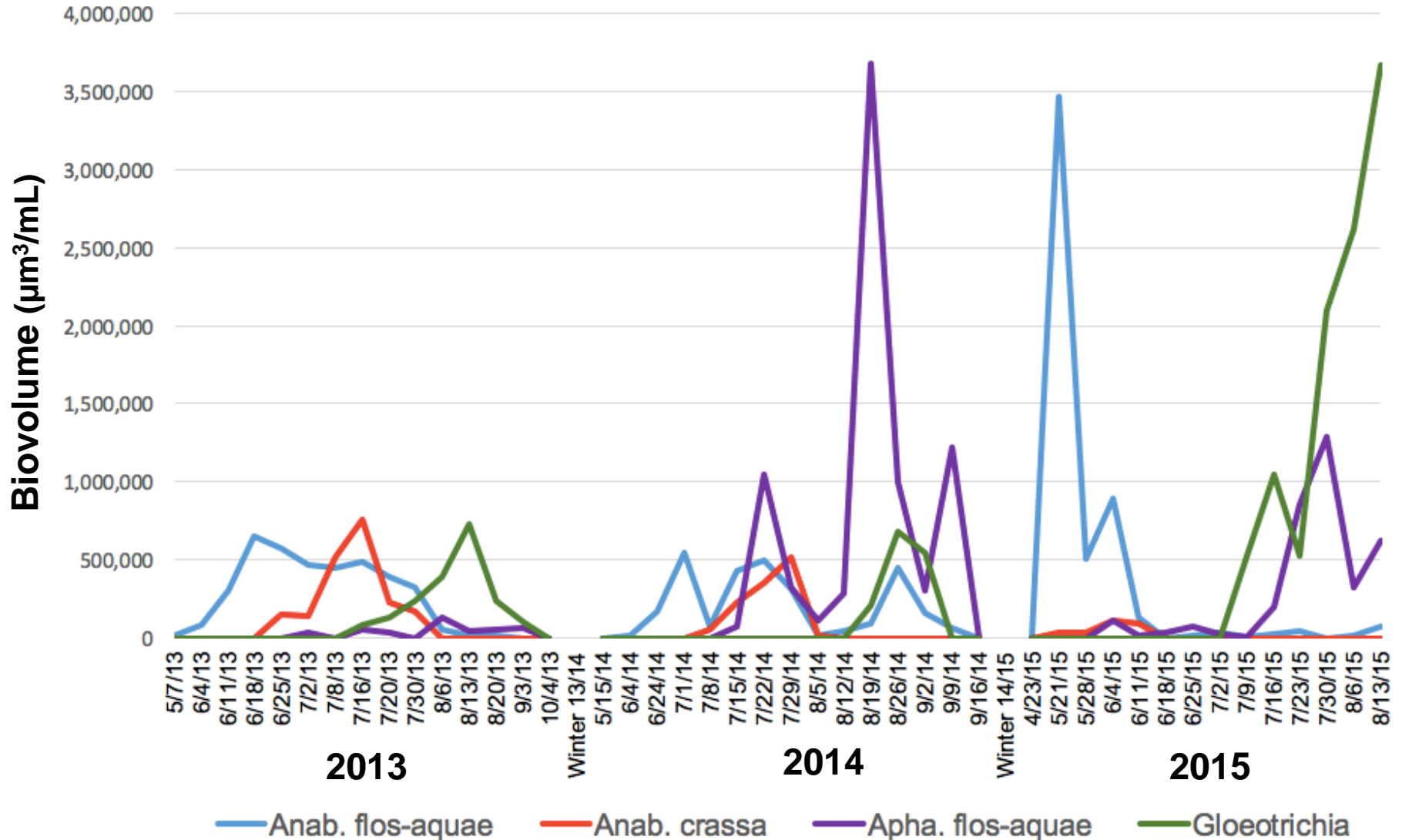
5.7 Mbp
Single
chromosome
ana+



Nathan Brown

Cyanobacterial successions in Dexter Reservoir

**** All nontoxic ****

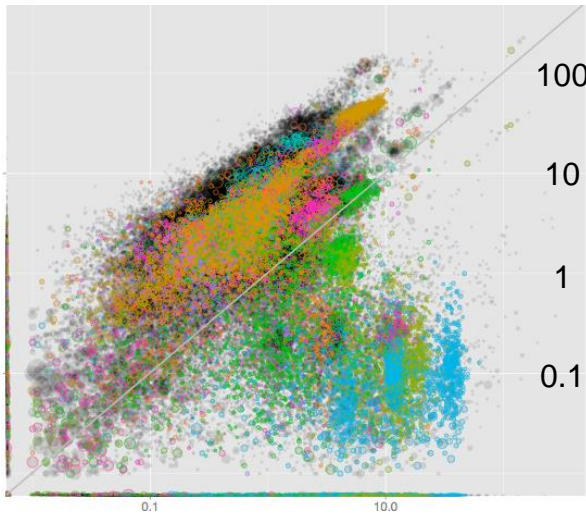


Shotgun metagenomic analysis

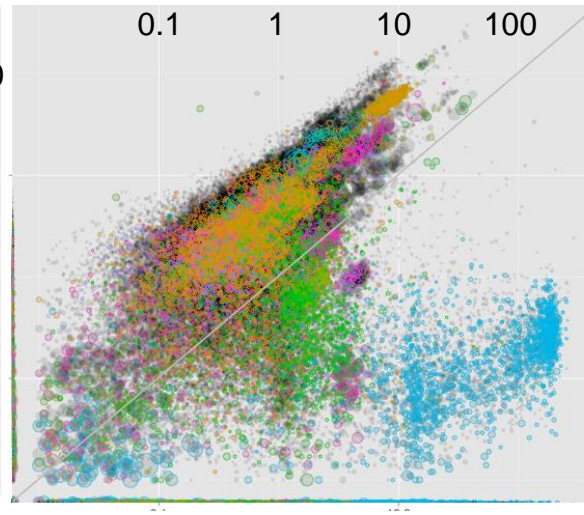
Dexter Reservoir 2014

Relative read depth
0.2 - 1.2 μm fraction

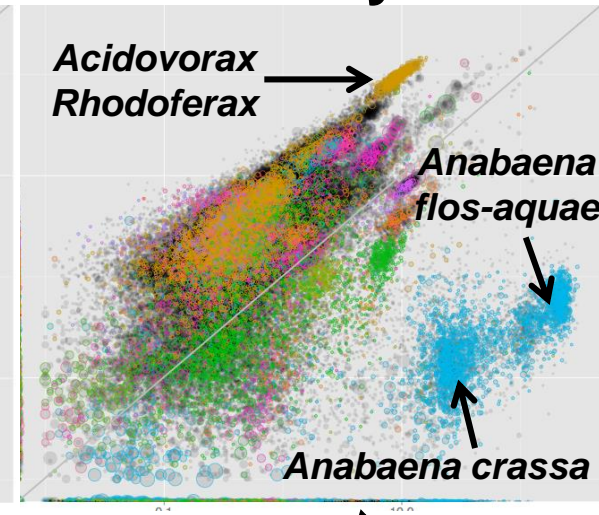
17 June



24 June

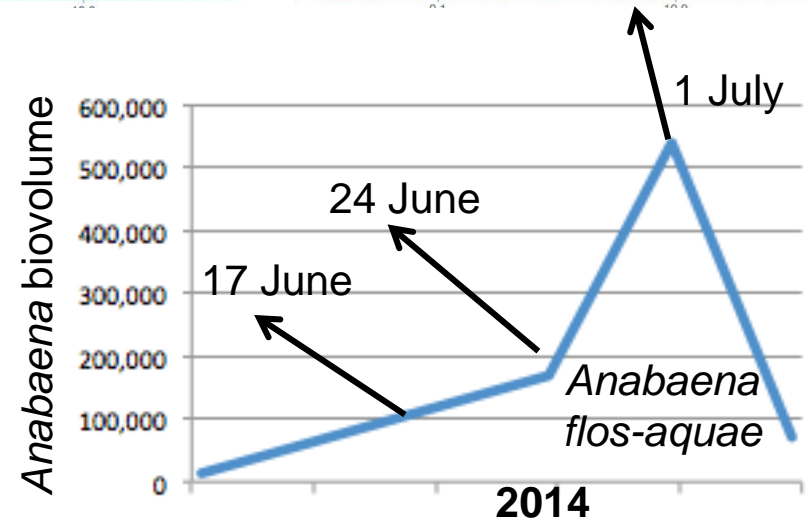


1 July



Relative read depth
>1.2 μm (GF/C) fraction

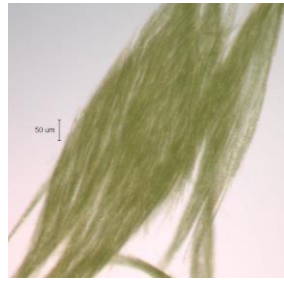
Illumina HiSeq metagenomes
assembled using IDBA, prokaryotic
contigs taxonomically sorted with
PhylopythiaS+, and clustered with
mmgenome



Tim Otten



Upper Klamath Lake *Aphanizomenon flos-aquae* non-axenic culture community genome sequencing



Bacterium	Genome size (Mbp)	Genome completion %
<i>Aphanizomenon flos-aquae</i>	4.25	97
<i>Alphaproteobacterium</i>	3.5	100
<i>Betaproteobacterium</i>	3.4	100
<i>Bacteroidetes</i>	3.2	100

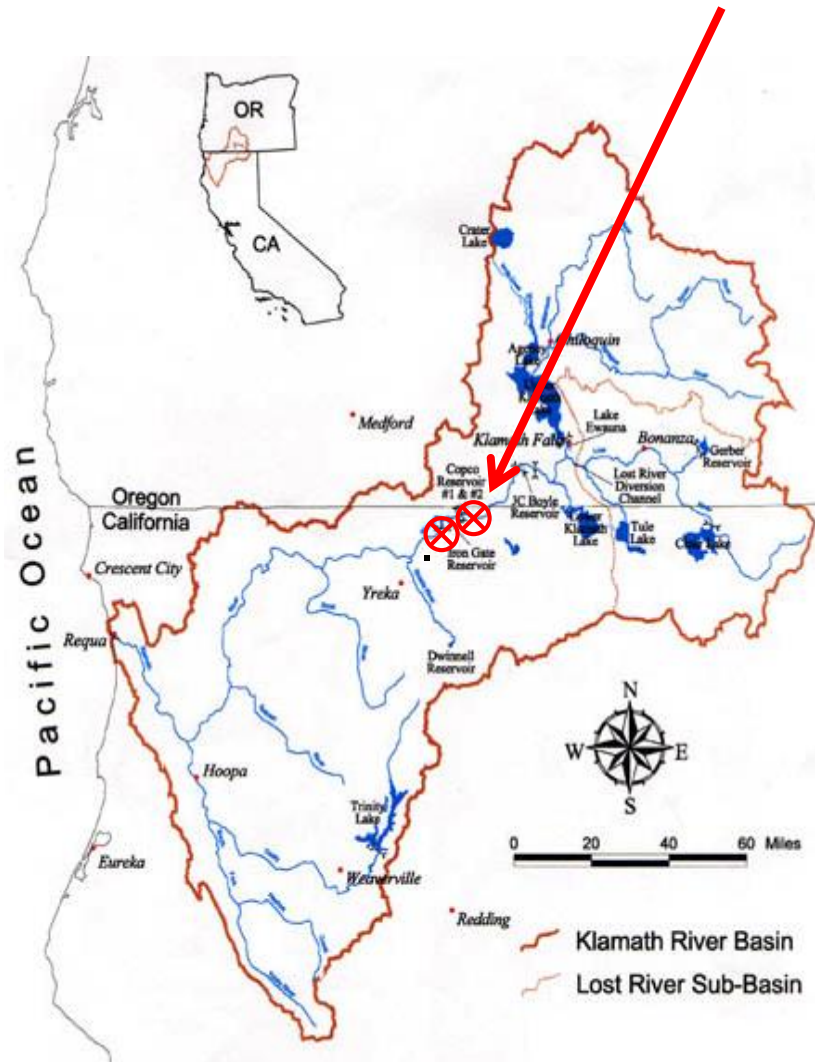
PacBio SMRT cell sequencing

- The **3 novel bacterial genomes** indicate heterotrophs dependent on *Aphanizomenon* for N (probably provided as NH_3), C and maybe even reduced forms of S.
- *Bacteroidetes* seems to encode a secreted peroxidase, which AFA lacks; may provide protection against extracellular reactive oxygen species (ROS)

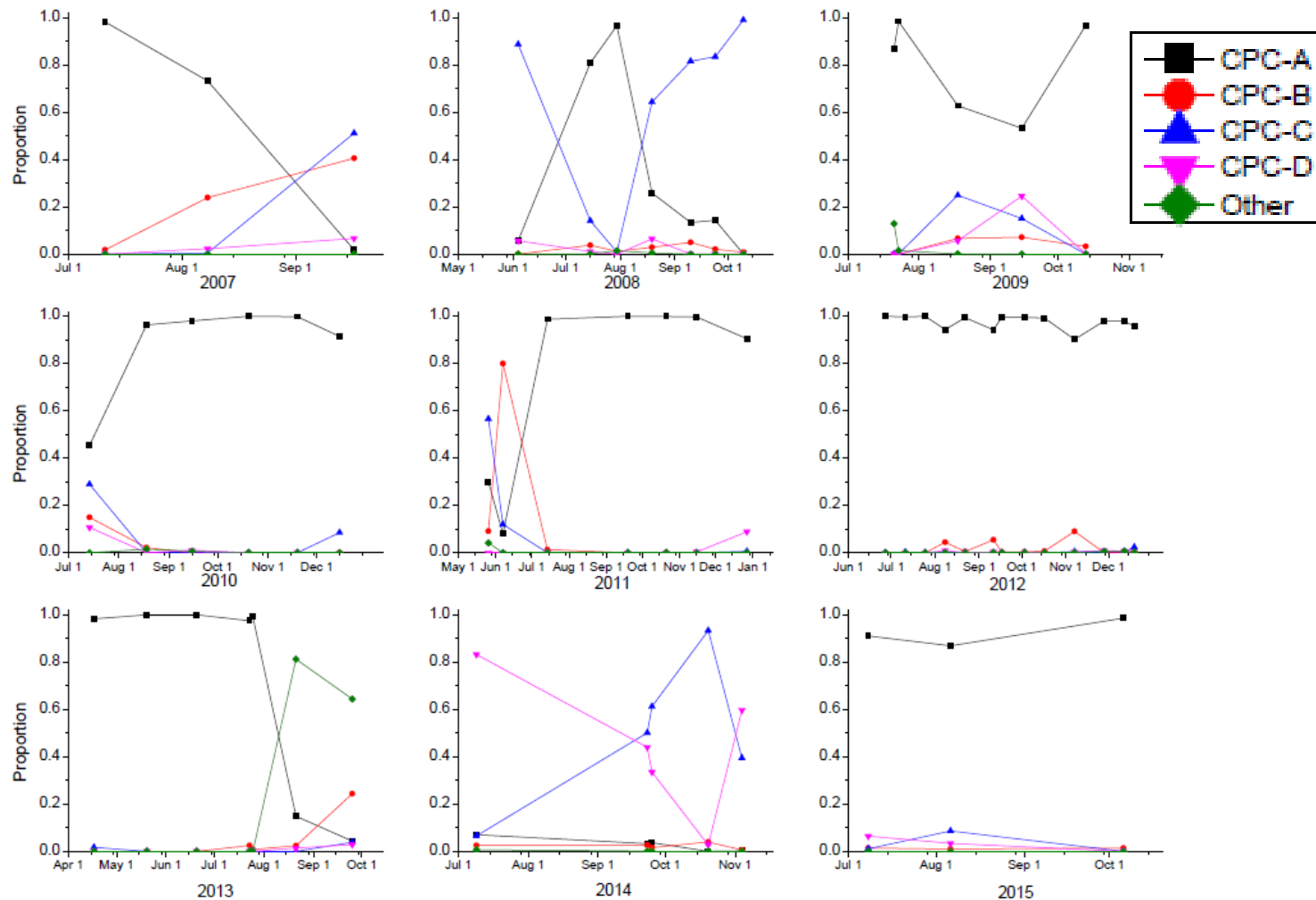
Connor Driscoll

Klamath River Basin

Microcystis aeruginosa blooms and microcystin levels exceed WHO guidelines annually in Copco and Iron Gate Reservoirs
and frequently in 300 km of river downstream



Copco Reservoir *Microcystis* strain dynamics 2007-2015



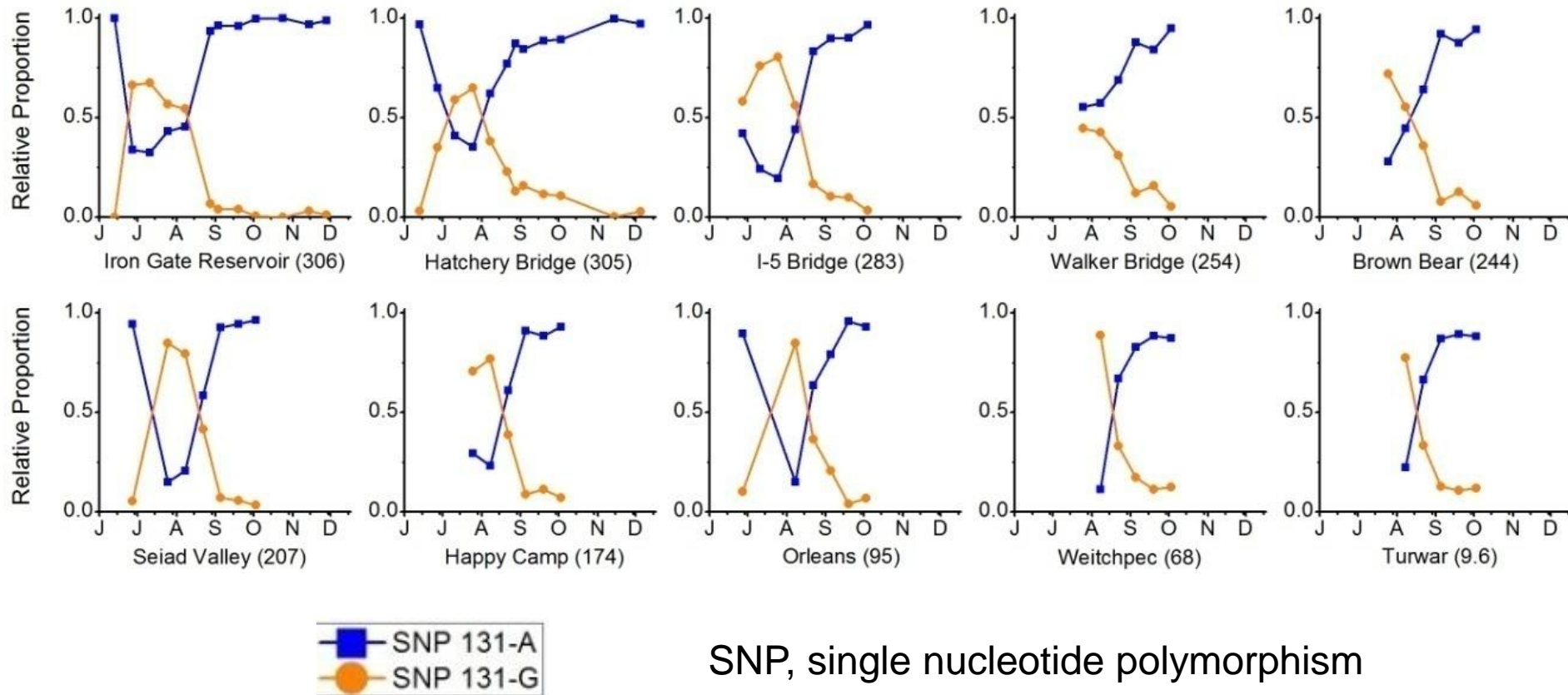
Phycocyanin gene *cpcBA* Illumina amplicon analysis

CPC-A encompasses the major **toxic strain**

CPC-B,C,D are correlated with low toxicity

Tim Otten

SNP DNA fingerprinting connects *Microcystis* populations in Iron Gate Reservoir and 300 km of downstream Klamath River (2012)



454 pyrosequencing ($n = 98,029$ *cpcBA* amplicon reads)

Tim Otten

Genetic technologies enable

- Identification of genetically distinct cyanobacteria present in a bloom; development of strain-specific PCR-based primers for high-throughput monitoring
- Inventory of toxin and T&O genes present; in which cyanos?
- Deduction of physiological optima for different strains from genome content; prediction and experimental study of strain competition and succession scenarios
 - Development of strain-specific predictive models for competitiveness under nutrient reduction or climate change scenarios
- Use of gene tracking to study spread and emergence of CyanoHABs

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