

US EPA ARCHIVE DOCUMENT



# Using AOPs TO Quantitatively Predict Chemical Impacts on Fish Reproduction

PPDC Workshop  
July 9, 2013

# Introduction



2

## The Problem

- Animals act in a complex manner when exposed to chemicals.
- Effects often determined on lower biological level data (in vivo and in vitro assays) but need to predict effects on populations
- Dose-response often nonlinear.

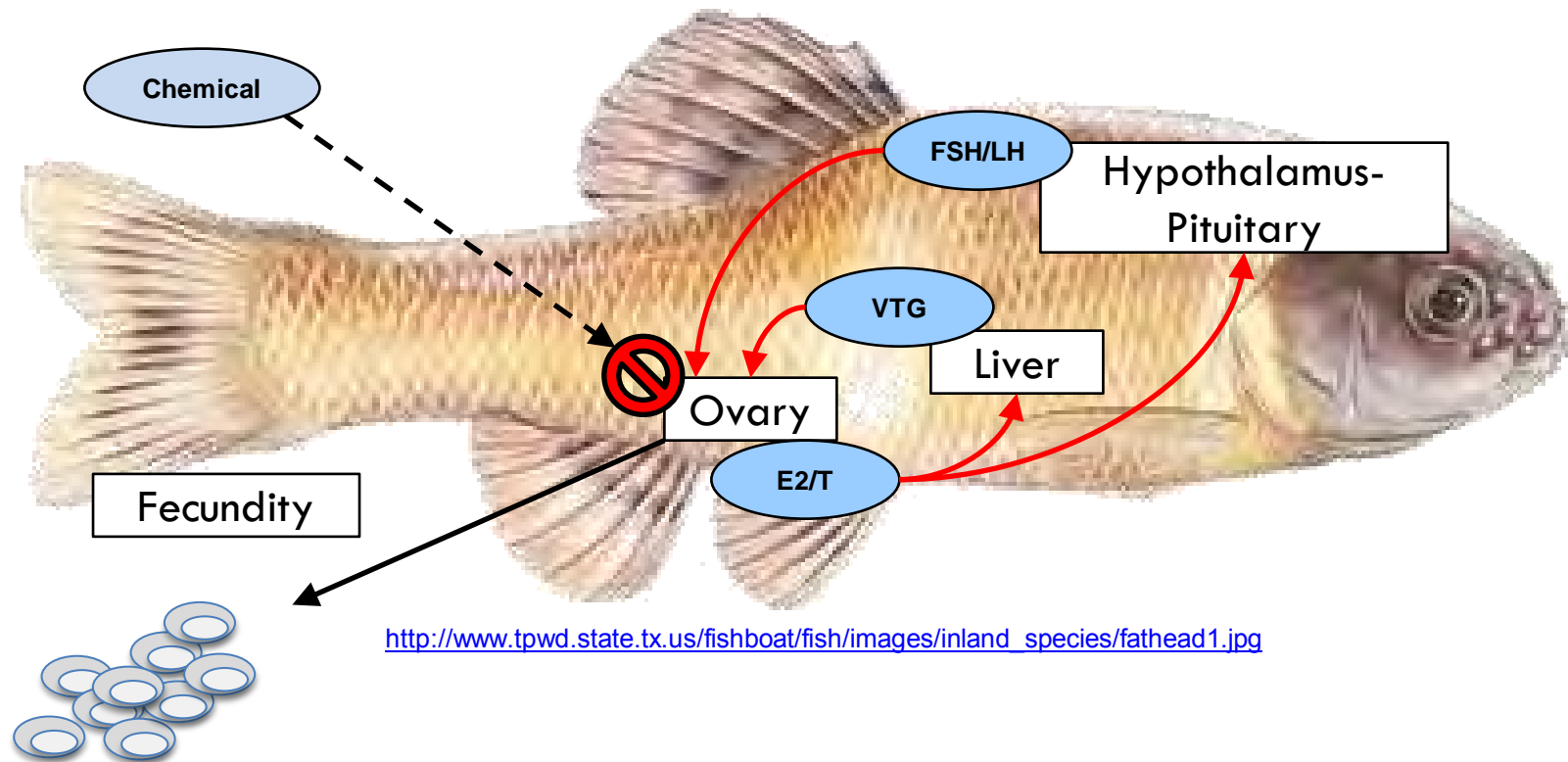
## Solutions

- Extrapolation across biological scales to predict population impacts
- Systems biology models representing a dynamic, quantitative model of the fish reproductive AOP
- Parameterization of a fish reproductive AOP model with in vitro/in vivo data

# Hypothalamus-Pituitary-Gonadal (HPG) axis controls reproduction



3



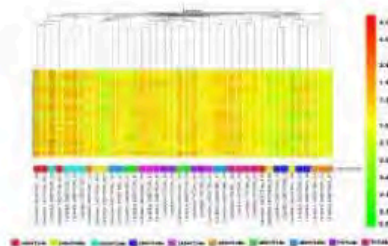
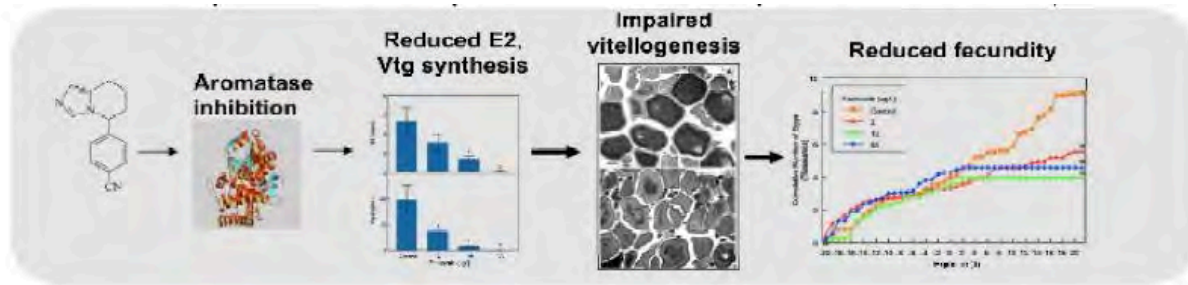
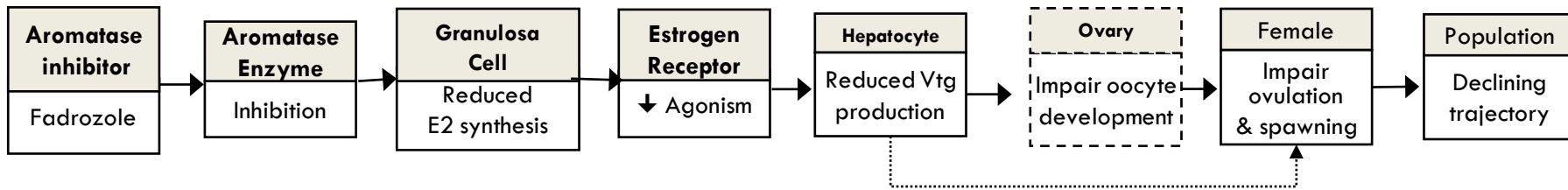
[http://www.tpwd.state.tx.us/fishboat/fish/images/inland\\_species/fathead1.jpg](http://www.tpwd.state.tx.us/fishboat/fish/images/inland_species/fathead1.jpg)

synthesis and regulation of reproductive hormones  
17 $\beta$ -estradiol (E2) and testosterone (T) which controls  
egg protein production, leading to egg production

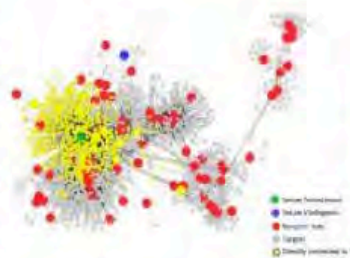
# Reproduction-related Adverse Outcome Pathway (AOP): aromatase inhibition



4



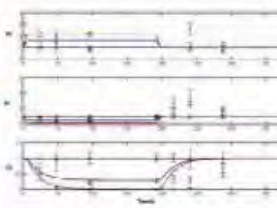
Screening for toxicological effects and chemicals



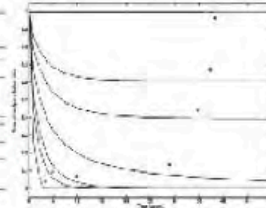
Pathway and network impacts



Mechanistic modeling



Predicted effect

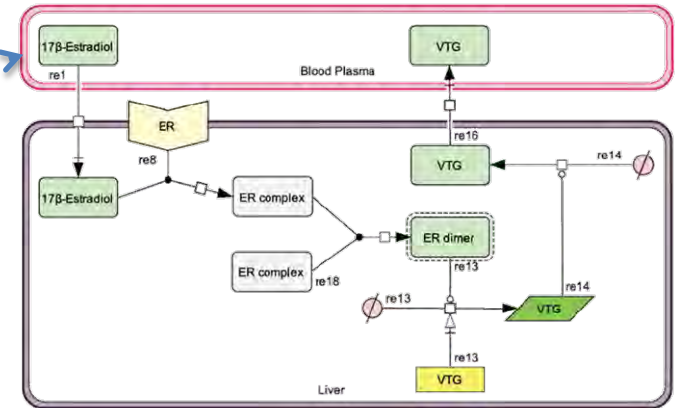
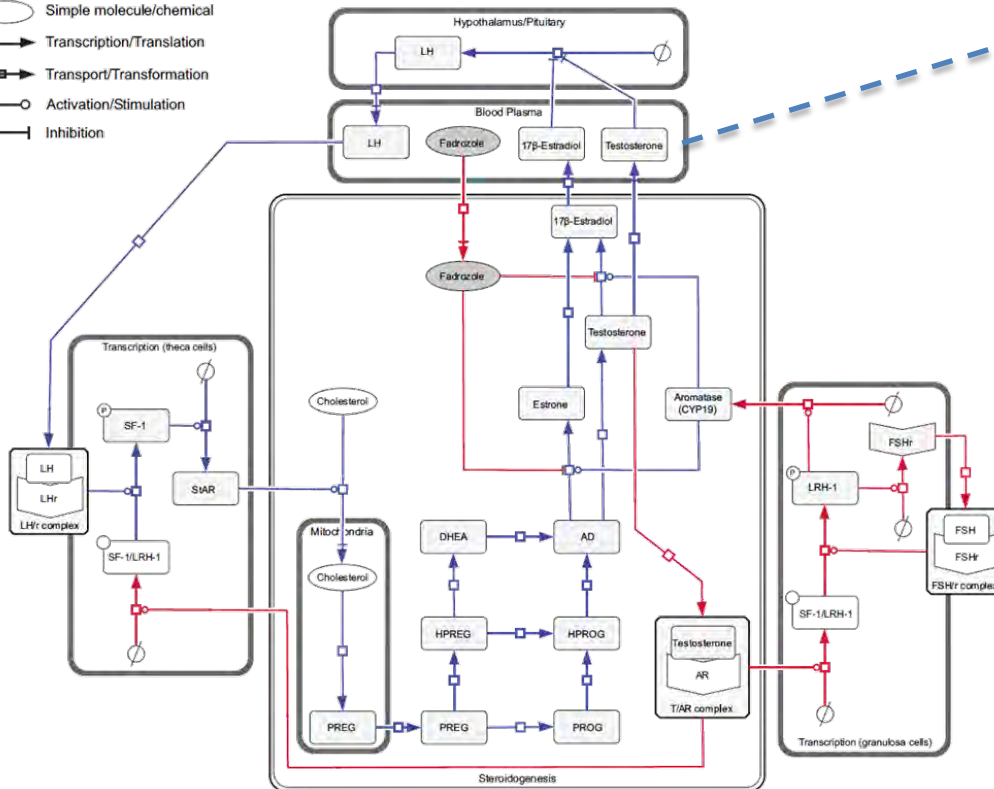
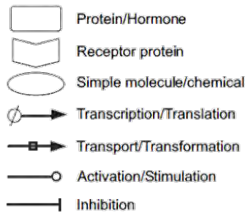


Population impact

# Conceptual mathematical model



5



- Liver compartment (vitellogenesis)
- Takes plasma E2 input and models:
  - ER binding E2
  - ER complex homodimerization
  - ER complex transactivation of vitellogenin

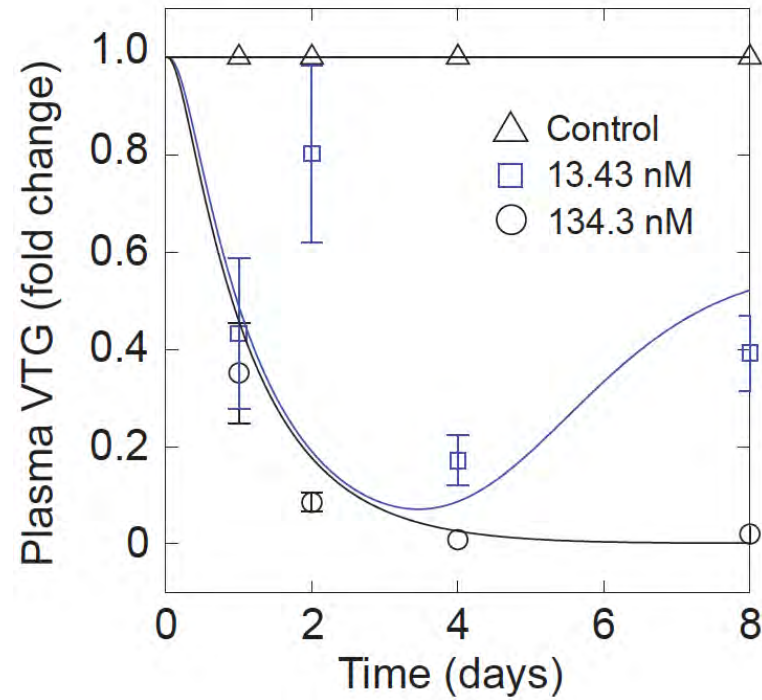
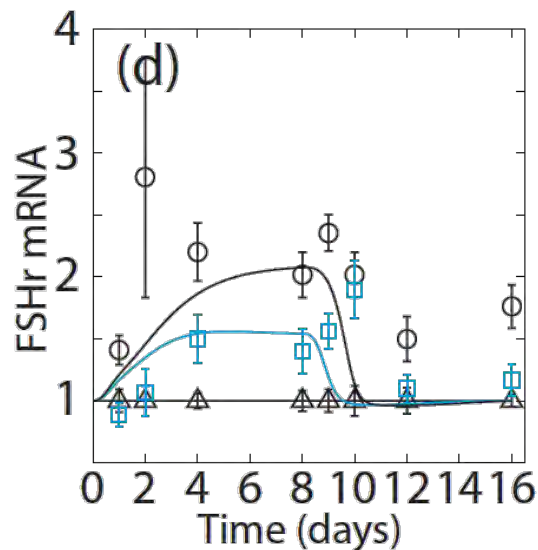
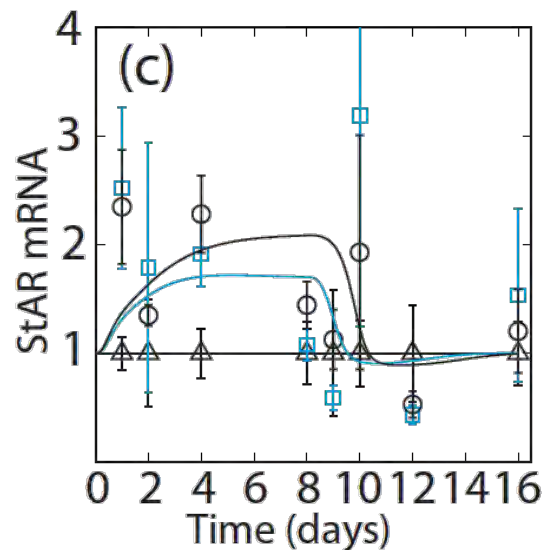
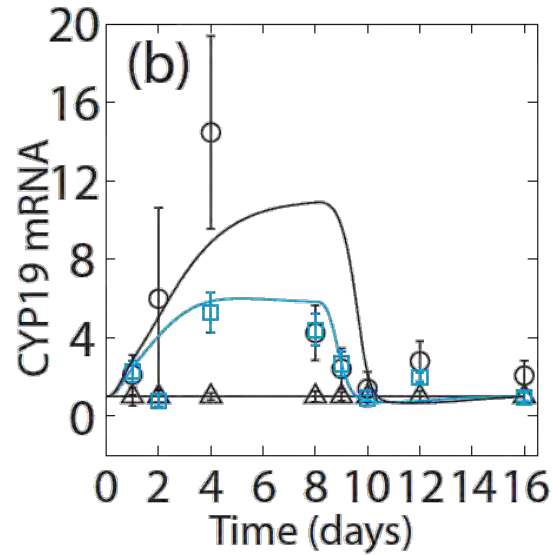
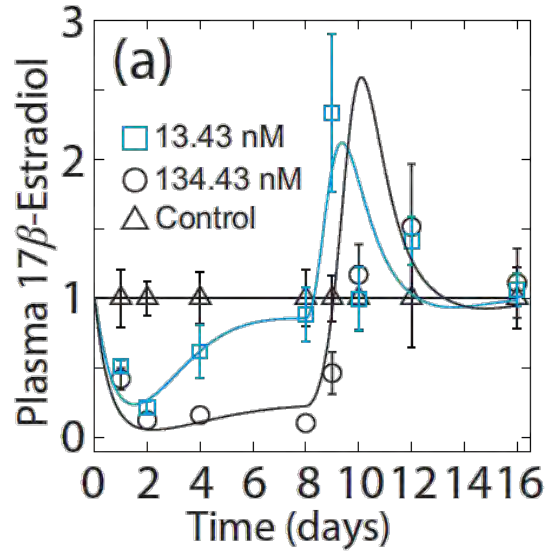
M. Mayo et al., (in preparation)

- Androgen receptor signaling initiates transcription, translation, and phosphorylation events
- Varying Fadrozole exposures cause varying plasma E2 response

# Predicting hormone and egg protein concentration profiles



6

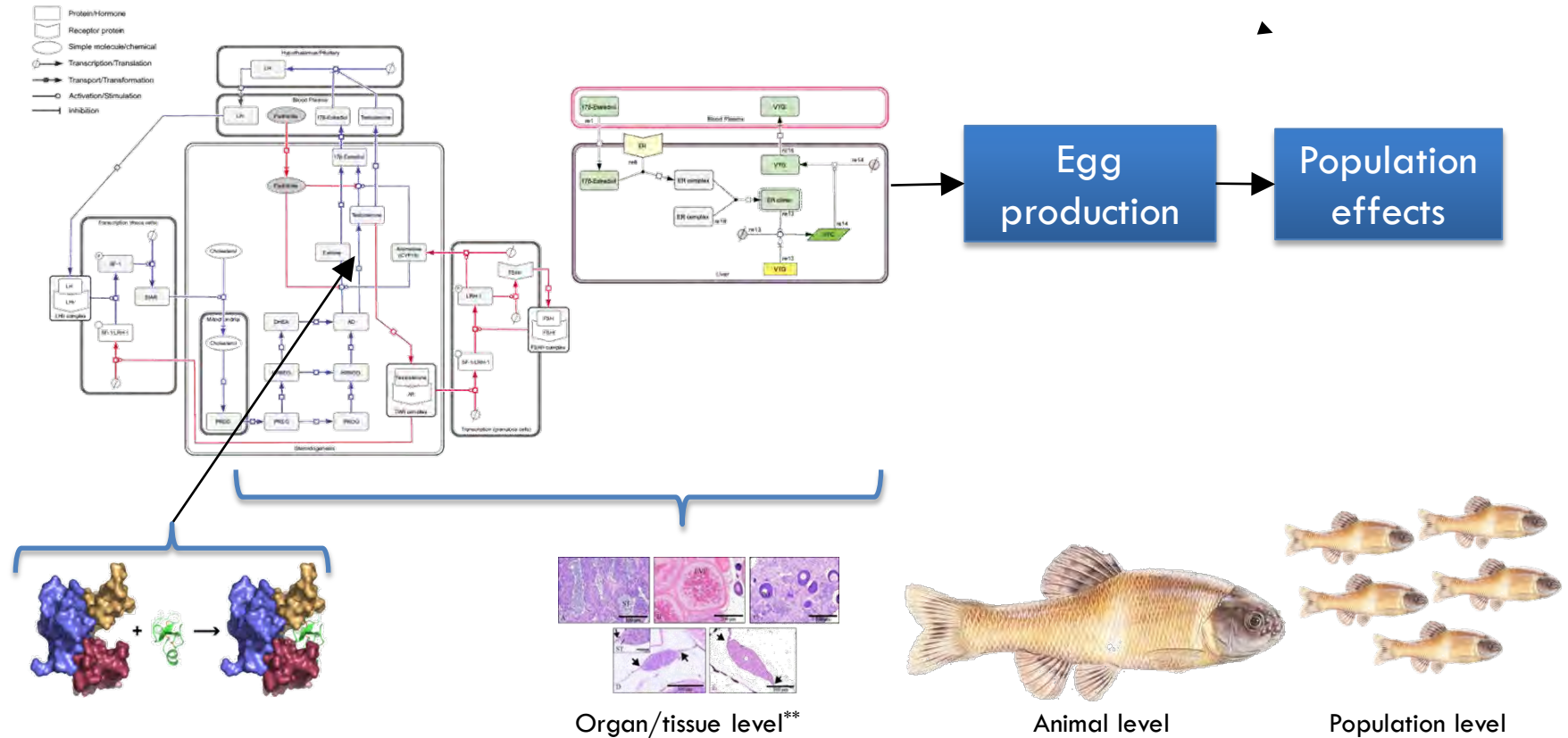
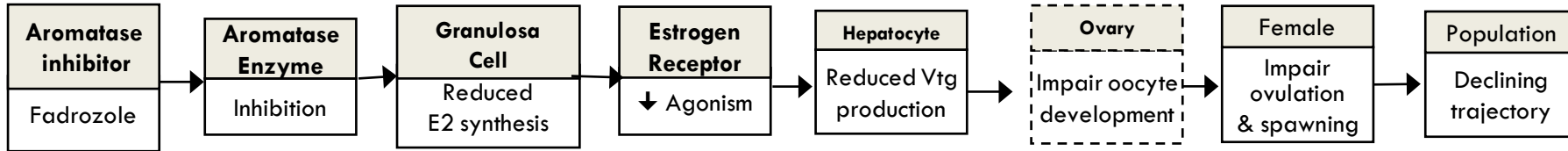




# Quantitative AOP



7



Egg production

Population effects

Organ/tissue level\*\*

Animal level

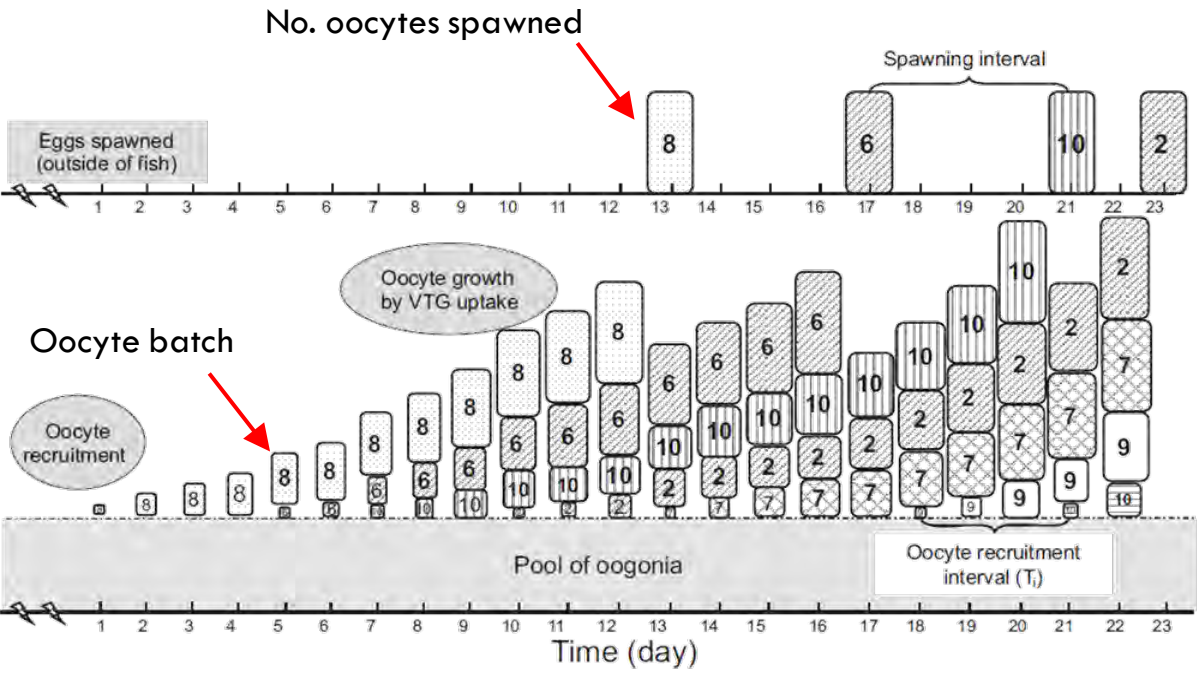
Population level



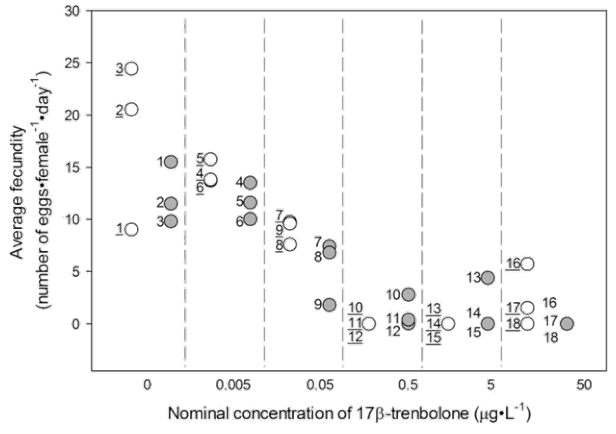
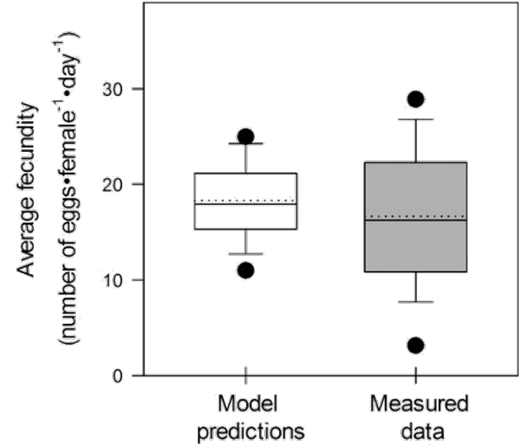
# Linking VTG to fish fecundity

8

- Leverage an existing mathematical model of FHM oogenesis
- Takes (dynamic) VTG concentration as input



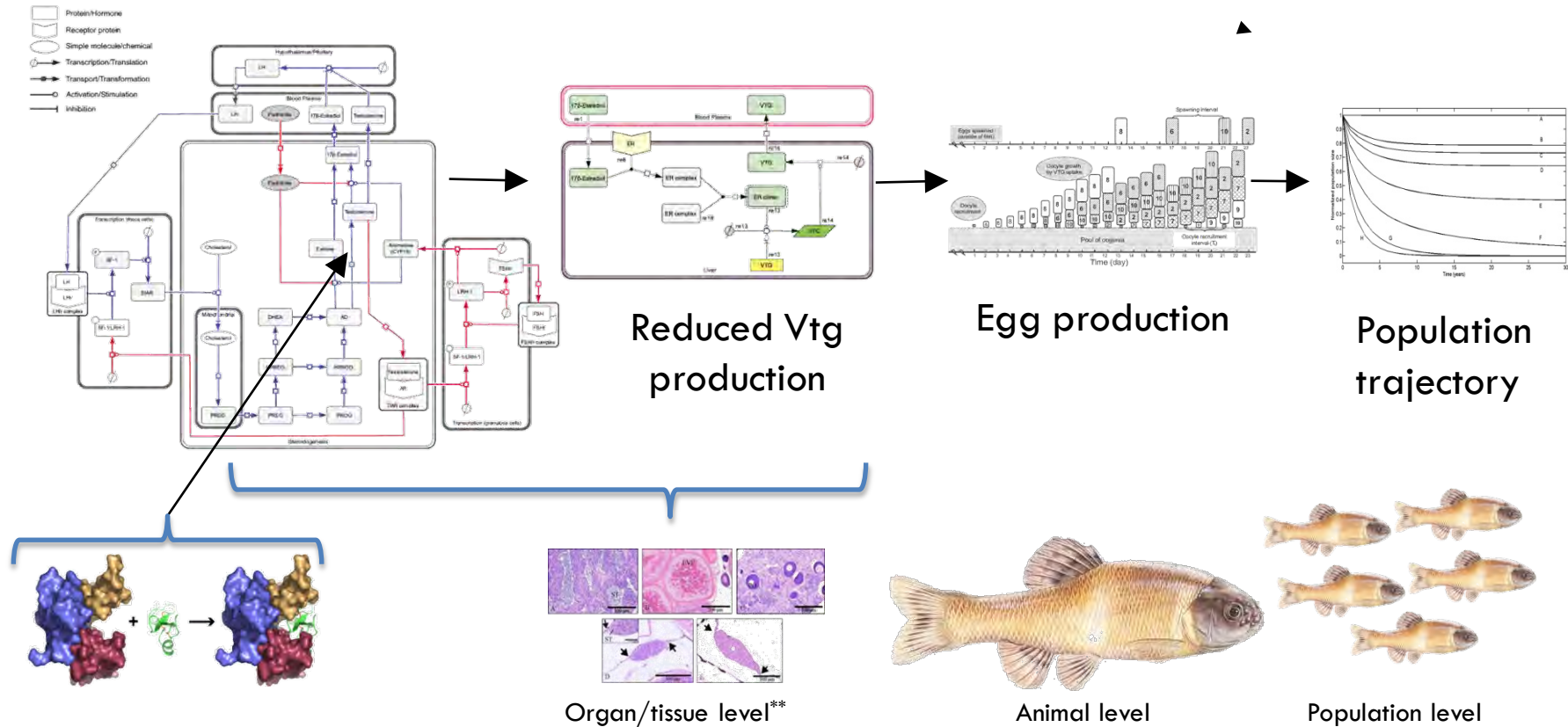
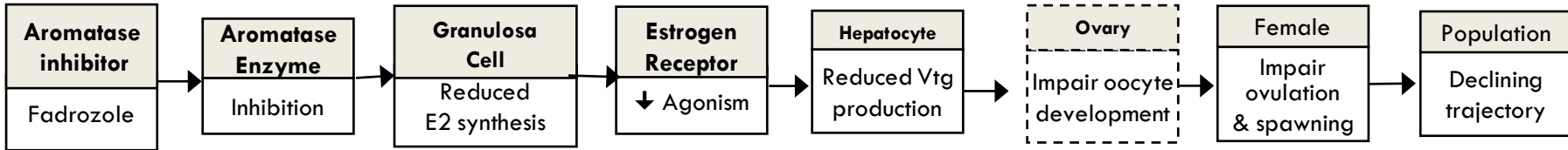
Z. Li et al., Can. J. Fish Aquat. Sci. **68**, 1528 (2011).



# Quantitative AOP



9



# Continuing work/applications



10

- Include the effects of other endocrine disrupting compounds
- Design front-end toolkit to carry out “virtual assays” through scenario playing with dynamic exposure conditions
- Use VTG measurements to parameterize model
- Use steroidogenesis/Aromatase inhibition factor to translate in vitro effects to population effects
- Include time dependent exposure
- Permit scenario playing with different effluent exposures to assess impact

Oocyte Growth Dynamics Model – Welcome

The Oocyte Growth Dynamics Model (OGDM) for Matlab is based on Li et al. (2011), Canadian Journal of Fisheries and Aquatic Sciences 68 (9), pp. 1528-1538. It simulates spawning for adult female fathead minnows over a user-specified time period.

Please note: spawning interval and clutch size data were obtained from 13 laboratory studies with observation periods ranging from 3 to 7 weeks (see Li et al. for study citations).

**Input Data**  
The OGDM requires a user-specified simulation period, fish identification (ID) and corresponding plasma vitellogenin concentration for each fish being simulated. The OGDM assumes that ovary vitellogenin concentration equals plasma vitellogenin concentration.

**Model Output**  
For each fish simulated, the OGDM will output a table (see Table 1) with a column of days for the simulation period specified, and subsequent columns representing each fish and their clutch sizes on each day of the simulation period.

Additional model results include cumulative fecundity and the total number of clutches produced in the simulation period per fish.

Table 1: Clutch size per day for three fish and a simulation period of 10 days.

Time (hr)	Fish ID 1	Fish ID 2	Fish ID 3
24	0	75	0
48	0	0	200
72	150	0	0
96	0	0	0
120	0	95	0
144	0	0	0
168	123	0	0
192	0	0	0
216	20	0	165
240	117	85	0

Continue Exit

Coded for Matlab by Karen H. Watanabe, August 2011  
Disclaimer: The OGDM is provided "as is." Clicking on the Continue button implies acceptance of the End User License Agreement (EULA). [View EULA](#)

- GUI-based software tool in development
  - Directed toward risk & hazard assessors

# Acknowledgements



11

## USACE-ERDC

### **Mathematical modeling**

Dr. Michael Mayo—US Army ERDC

Dr. Karen Watanabe—Oregon Health & Science University

### **Experimental data and analysis**

Dr. Natalia Garcia-Reyero

Dr. Tanwir Habib

### **With the assistance of US EPA Duluth**

G. Ankley, J. Berninger, J. Cavallin, E. Durhan, K. Jensen, M. Kahl, C. LaLone, E. Makynen, D. Miller, M. Severson, K. Stevens, D. Villeneuve

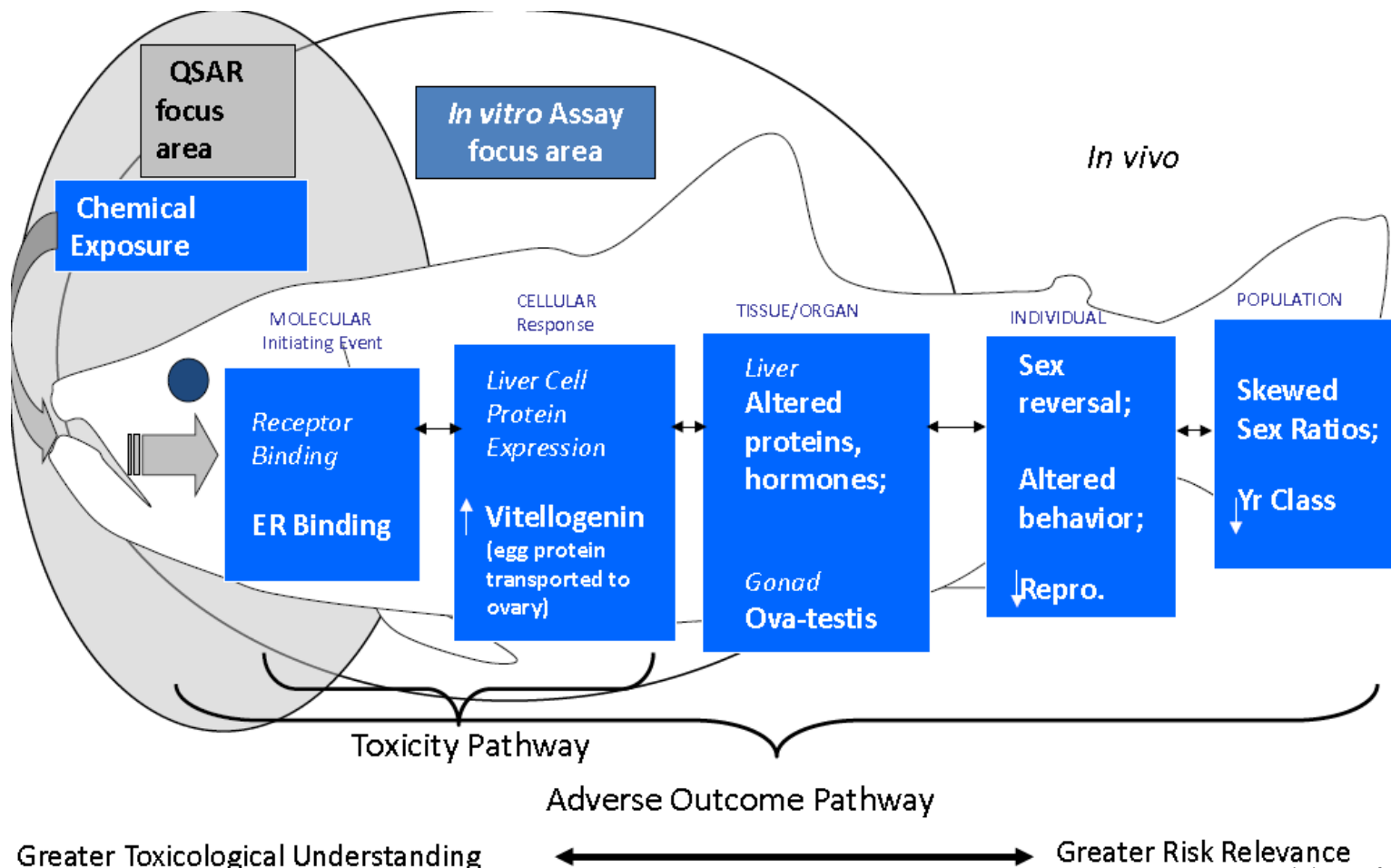
### **Small Fish Comp-Tox Group**



# OPP Ecological Risk Assessment Perspective – Fish Reproduction AOP



12



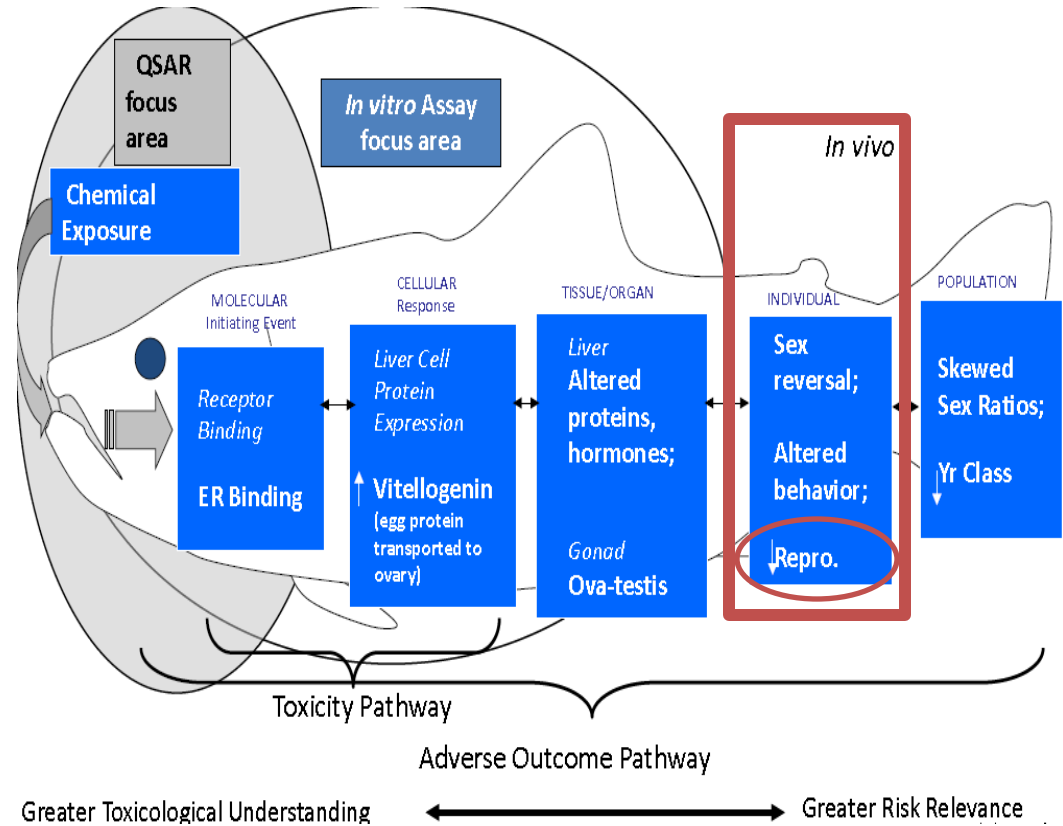


# OPP Ecological Risk Assessment Perspective – Fish Reproduction AOP



13

- ❑ Currently, screening level (apical) endpoints: survival, growth, and **reproduction**
- ❑ Small number of fish *in vivo* studies surrogates for all fish species
- ❑ Compare most sensitive apical endpoint effect to relevant exposure (quantitative) to evaluate potential risk concerns





# OPP Ecological Risk Assessment Perspective – Fish Reproduction AOP



14

## □ Current Utility of AOPs

- (Q)SAR and *in vitro* data used to determine whether additional *in vivo* fish toxicity data is needed to support risk assessment
  - Typically for degradates of concern
- (Q)SAR and *in vitro* data used to determine if bridging fish data across chemicals is appropriate
  - Provides information where a data-gap may exist
- Provide mechanistic information for weight-of-evidence decisions (e.g., Tier I EDSP battery)
  - *In vitro* assays (e.g., estrogen and androgen receptor binding assays) can provide mechanistic information when evaluating *in vivo* assays including the fish assay
- Sublethal effects (e.g., clinical toxicity signs including biochemical alterations) can be used to qualitatively characterize risk to fish

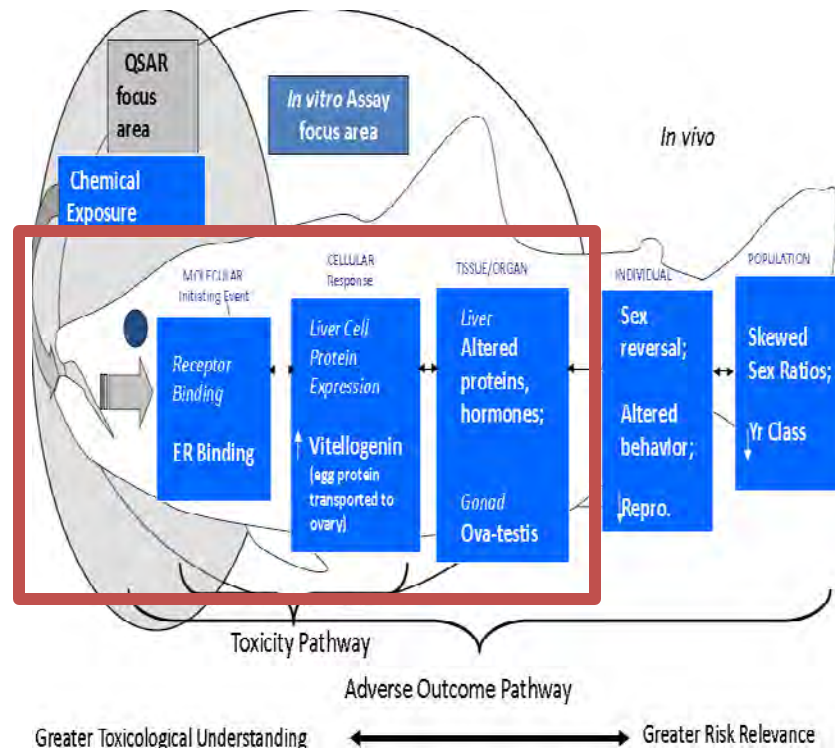
# OPP Ecological Risk Assessment Perspective – Fish Reproduction AOP



15

## Challenges with using AOPs:

- Quantitative linkages to apical endpoint
  - ? Change in Vtg  $\rightarrow$   $\downarrow$  Reproduction
- Accounting for competitive AOPs and/or compensatory mechanisms in whole animals
- Capturing potential adverse effects from a non-defined AOPs
  - Reproduction can be affected by many different factors
- Uncertainty in if quantitative linkages are reflective for broad range of fish species



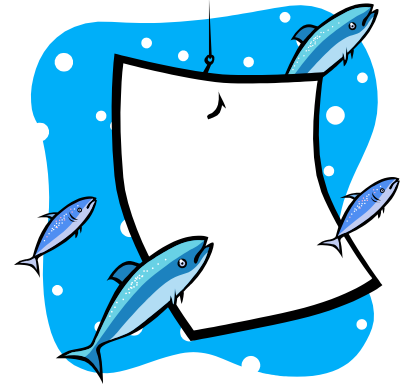
# OPP Ecological Risk Assessment Perspective – Fish Reproduction AOP



16

## □ Things to Consider for Risk Assessment

- Was the model used to generate data appropriate?
- Do effects occur at environmentally-relevant concentrations/conditions?
- Do reproductive effects occur in the presence of potential overt toxicity (e.g., death, erratic swimming, hemorrhaging)?
- Where are the effects in relationship to other existing toxicity data (dose-response curve and time course)?

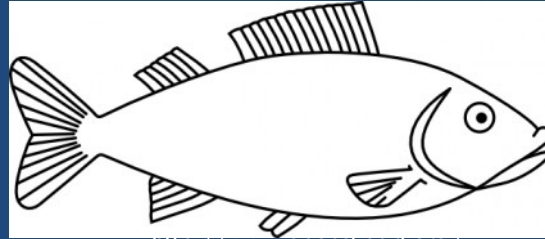


# OPP Ecological Risk Assessment Perspective – Fish Reproduction AOP



17

- **Future Development of AOPs that will inform potential effects on fish reproduction**
  - ▣ Continue to develop linkages with current AOPs
    - Provide quantifiable measures (linkages) from low to high biological levels of organization (including fecundity)
  - ▣ Develop new AOPs
  - ▣ Develop network of cumulative impacts from potentially perturbations from multiple AOPs



<http://www.opencart.org/>



# Thank You! Questions?