

US EPA ARCHIVE DOCUMENT

**Coastal 2000
Northeast Component**

Information Management Plan

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1. Introduction

Information management for the Coastal 2000 project will occur on three distinct levels: national, regional, and state. At the national level, a central database will contain summarized data from all regions of the coastal United States. The National Coastal Database will contain only well documented data that has passed strict QA review, and will be made available to a wide audience. At the next level down, regional information management centers will maintain detailed databases to be used for regional assessments, containing data from several states. At the state level, even more detailed data may be stored. Each state participating in the Coastal 2000 effort will use their own information management systems to store data collected at Coastal 2000 sampling sites. The states will provide core indicator data required by Coastal 2000 to the regional information centers, where it will be merged with data from other states in the region. The regional data centers will provide merged data from several states to the national data center.

This document describes information management procedures for the Northeast Information Management Center (NEIMC) of Coastal 2000- one of the four regional IM centers for the project for the year 2000.

First, the planned flow of data from the state level to the regional level is described. Options for state level information management are discussed. Recommended field datasheets and a computerized data entry system, which will be provided to all participating states by the NEIMC, are described. Procedures for tracking samples shipped to analytical laboratories are described. Procedures to be used by analytic laboratories to submit data to the NEIMC are also described.

Next, the Northeast Region SAS Database is described in detail. This description defines all the core parameters that are required from the nine participating states.

Finally, specific data transfer and formatting procedures are suggested for the state agencies and analytic laboratories submitting data to the regional database.

The purpose of this document is to present data formatting and transfer protocols that will allow a great deal of data collected by many different participants to be loaded into a uniform database in a timely manner. These protocols were first developed for the Mid-Atlantic Integrated Assessment (MAIA) Estuaries project, and proved to be effective in managing data from diverse sources. The Coastal 2000 project recognizes that individual states may have other protocols in place for managing data. This is acceptable, providing that the state makes a detailed proposal on how their data will be submitted to the NEIMC, and get prior approval from their cooperative agreement's project officer.

2. Data Flow in the Coastal 2000 Northeast Region

The NEIMC will store data collected by nine states in one central database. Data supplied by states to the NEIMC will be stored in two ways. The core indicator data required by the Coastal 2000, will be loaded into a database made up of SAS datasets. The purpose SAS database is to store all required Coastal 2000 field and lab data in a consistent format, and is designed to facilitate regional assessments. Additional non-core data collected by different states will be stored as individual ASCII text files, with accompanying documentation files.

The SAS database design is described in detail in Section 3 of this document. Individual states are expected to format core indicator data in accordance with this design before submitting them to the NEIMC. Specific guidelines for formatting data to be loaded into the SAS database, as well as other non-core data files, are discussed in the Section 4.

The SAS database design allows certain data collected by states above and beyond the Coastal 2000 core requirements to be stored in the same database. These data include extra water chemistry analytes such as particulate nutrients, and additional sediment contaminant data. The design of the database allows these data to be stored in the same database as the core indicator data if analyte codes and adequate documentation are provided by the states that collected them.

Additional data that will not fit in the SAS database design will be collected by many states. One example is bacteria counts, routinely collected by some states. These data are not part of the Coastal 2000 sampling plan, but are of interest to regional assessment groups. They will be stored at the NEIMC in ASCII text data files in formats submitted by the states. These data must contain valid Coastal 2000 Station Identifiers, and must be accompanied by detailed documentation. All data files should be reviewed by state QA personnel and deemed error free before being submitted to the NEIMC.

Data provided by the states will consist of measurements taken in the field during sampling (field data), and the results of laboratory analyses of field samples (lab data). Field data include locational information, date and time of sampling, and *in situ* measurements of water temperature, salinity, and dissolved oxygen level. Lab data include results of several analytic procedures measures such as of chemical analysis, toxicity testing, and benthic community measures. Lab data may also include QA information from lab replicate samples, blanks and standard reference materials.

All states will provide field data. Some states will provide lab data. However, most samples collected by for Coastal 2000 will by analyzed by national contract labs . The lab data generated under the national contracts will be sent directly from those labs to the NEIMC, so the states will not be responsible for formatting these data. Only states that chose to have

samples analyzed in their own labs will need to format lab data for the NEIMC.

2.1 Field Data

The NEIMC can provide states with datasheets that capture the all data required by the Coastal 2000 project (see Appendix C). These data sheets also capture information useful to project managers, such as the names of crew members, and some additional data useful for QA purposes. The datasheets are designed to accommodate bar-coded sample id. labels, as an alternative to hand-written sample id's. The use of bar-coded sample id. labels has been found to greatly reduce transcription errors.

The states are not required to use these datasheets. If state monitoring programs use other means for recording field data, those systems may be used. Some states may wish to use some of the datasheets provided by the NEIMC, but record other information (such fish survey data) on their own datasheets. Only the station setup sheet is needed for all sampling events by all states. The Coastal 2000 Quality Assurance Project Plan (Heitmuller, 2000) calls for all field data to be recorded on some form of hard-copy data sheet. The NEIMC should be provided with copies of all completed data sheets.

The NEIMC can also provide states with a computerized data entry program to enter data from the standard datasheets into an electronic files. This system is designed for Windows 95 (or higher) operating systems, and requires the proprietary software product FormFlow Filler. The states are not required to use this system, but they are expected to supply the NEIMC with electronic data files containing all relevant field data in a consistent format.

If the states chose to use the data sheets and computer system supplied by the NEIMC, they only need to supply the NEIMC with copies of the completed datasheets and the output database from the computer system, which consists of three Dbase files and their indexes. The NEIMC staff will process the Dbase data files and add the data to the regional database, and will also assist states in adding these data to their own information systems. The advantage of this option for states is simplicity- they need only enter data in an easy to use interface, and mail a diskette to the NEIMC. The advantage of this approach for the NEIMC is in its accuracy and reliability. The field computer system has already been used by EPA field crews for three years of MAIA field sampling with successful results.

States may chose to use their own data management systems for recording and entering field data. It is the state's responsibility to format their data into files that can be added to the regional database. In these cases, the format of the electronic data files supplied to the NEIMC should match the SAS tables described in Section 3.2 as closely as possible. In addition, a list of all samples shipped to national contract laboratories must be supplied to the NEIMC. Data

files should be in ASCII format and be accompanied by detailed documentation. Specific guidelines for formatting ASCII data files are provided in Section 4.

An important aspect of field data management is sample tracking. Many physical samples will be collected in the field and shipped to analytic laboratories. Samples must be clearly labeled with identifiers that laboratories will use when reporting results data. These sample identifiers must be recorded in the field. The use of bar coded sample labels is highly recommended by the NEIMC. These labels can be scanned into the computer system with a bar code wand, which greatly reduces the chance of data entry errors.

The sample identifier consists of a two letter state code, a four digit station number, and a three to four letter code that identifies the sample type. It is very important that the correct sample identifiers are used on samples. These identifiers will be used to associate laboratory results with sampling events.

Field crews should record all samples on a shipment tracking datasheet as they are packed for shipping to laboratories. These data must be forwarded to the NEIMC for all samples shipped to national contract labs.

2.2 Laboratory Data

Most samples collected in the field for Coastal 2000 will be analyzed at national contract laboratories selected by EPA. Results of these analyses will be sent directly to the NEIMC in a predefined format. These formats are described in detail in the Lab Formatting and Transfer Guidance Document (Buffum, 2000). It will be the NEIMC's responsibility to process these raw data, check them for errors, and add them to the database. The states will have access to the data as soon as they are in the database.

States may chose to have some samples analyzed by their own laboratories. In these cases, state IM personnel will be responsible for formatting the data before sending it to the NEIMC. This formatting includes matching the lab results with the correct sampling station and sampling date, and in some cases with the replicate sample number. For example, the may be three sets of nutrients samples collected at three different water depths at each station. The results of the analyses of these samples must be tied to the correct water layer. Accurate sample tracking is crucial to accomplishing this.

The lab results sent to the NEIMC should be formatted in a manner consistent with the SAS database design, and must be clearly documented. State IM personnel should check the data for errors, and verify that lab QA procedures were followed. Data files must be in ASCII format and be accompanied by detailed documentation. Specific guidelines for formatting ASCII data

files are provided in Section 4..

2.3 Metadata

An important part of any scientific database is the metadata, i.e. information about the data. Metadata files for the Coastal 2000 data will be written in the format required by MAP (Frighten and Steel, 1995). Each SAS data table in the central database will have an accompanying metadata file. These metadata files will be compiled by the NEIMC, using information supplied by the states. The information needed for the metadata files includes detailed descriptions of sampling methods, sampling gear, laboratory methods and QA protocols used to produce the data. The states will be provided short forms to fill out for the data they contribute (see Appendix B). These forms capture the required metadata text for all the core indicators.

The states will be required to supply full metadata text for any data file contributed to the NEIMC is not consistent with the format of the SAS database. The metadata should be aimed at a wide audience of users who may not be familiar with the data, and should provide all information needed to analyze the data meaningfully.

3. The Coastal 2000 Northeast Regional SAS Database design

The SAS database consists of twelve primary data tables, one table containing QA information, and three code tables. These data will be stored initially as SAS datasets, and will be made available to end-users as comma separated value (CSV) ASCII files from a web page. Each data table will have an accompanying metadata file which will provide detailed descriptions of all data elements. The metadata files will provide information on sampling methods, laboratory methods, and QA protocols, and references to other related documents.

3.1 Overview of the SAS Database

The thirteen data tables that make up the database are:

Table Name	Description
STATIONS	Sampling Station Location Data
EVENTS	Station Visit Data
WATRPROF	Water Column Profile Data-Physical Measurements
FTRAWL	Fish Trawl Data
FISHSPEC	Fish Count by Species per Trawl
FISHPATH	Fish Length and Pathology Data
NUTRNTS	Water Quality-Nutrients Data
SEDGRAIN	Sediment Grain Size Data
TOXICITY	Sediment Toxicity and Microtox Test Data
SEDCHEM	Sediment Chemistry Data
CHEM_QA	Chemistry QA Sample Data
BEN_ABUN	Benthic Abundance Data
TISSCHEM	Tissue Chemistry

Three additional lookup tables provide information on coded variables:

BEN_TAXA	Benthic Taxonomy Table
ANALYTES	Chemical Analyte Code Table
FTAXON	Fish Species Names

The following section provides detailed descriptions of the individual data tables.

STATIONS - Sampling Station Location Data

This Stations data table will contain one record for each planned sampling station in the

Northeast Region. This data table will be maintained at the NEIMC and will define planned latitudes and longitudes of sites. States will be provided with these data for stations in their waters, and should not update them. The exact sampling location recorded at each visit to a station is recorded in the EVENTS data table, which may differ slightly from the planned locations. The data table contains the following elements:

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
STAT_ALT	Station Alternate Site Code (A,B, or C)
STATE	State station is located in
STA_TYPE	Station Type
ESTUARY	Estuary Name
STA_LAT	Latitude (decimal degrees)
STA_LNG	Longitude (decimal degrees)
ST_COOP	State Cooperative Agreement Station sampled under
LOCAL_ID	Alternate Station Identifier used by State

The data table will provide three possible sampling locations for each station, identified by the STAT_ALT code. All sampling will occur at only one of these locations- alternatives are provided in case the primary proves to be unsafe or impossible to sample.

3.2 Field Data Tables:

These data tables contain data collected in the field by sampling crews.

EVENTS - Station Visit Data

This data table will contain one record for each sampling visit to a station. If there are multiple visits to a station, each visit must have a unique record in this data tables. The two variables STATION and EVNTDATE make up the unique identifier for visits. The STAT_ALT indicates which alternate site was used for the sampling. EVENT_ID is an extra event identifier which may be used by states in their own databases. It is included here only for documentation purposes. The actual latitudes and longitudes (recorded upon arrival on station) may differ slightly from the planned latitude and longitude included in the STATIONS data table.

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event
STAT_ALT	Station Alternate Site Code (A,B or C)
EVENT_ID	Partner's Event Identifier

CREW_ID	Sampling Crew Identifier
EVNT_LAT	Latitude (decimal degrees)
EVNT_LNG	Longitude (decimal degrees)
DEPTH	Water Depth (meters)
SECCHI_D	Secchi Depth (meters)
SAV	Submerged Aquatic Vegetation Present at site (Y/N)
TRASH	Trash Present at site (Y/N)
MACROALG	Macro Algae present at site (Y/N)

WATRPROF - Water Profile Data-Physical Measurements

This data table contains measurements of temperature, salinity, dissolved oxygen, pH and light transmittance, at different depths, measured in the field during sampling. There may be several records for each event, containing measurements taken at different depths.

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event
W_DEPTH	Water Depth of measurements
TEMP	Temperature (Deg C)
SAL	Salinity (ppt)
OXY	Diss Oxygen (mg/l)
PH	pH (pH units)
PAR_SRF	PAR at Surface (mE/m2/s)
PAR_DPTH	PAR at Depth (mE/m2/s)

FTRAWL - Standard Trawl Data

This data table contains one record for each standard or non-standard fish trawl. By EMAP conventions, a standard trawl is defined as approximately 10 minutes in duration at 1 to 3 knots. The results of standard trawls are used for measures of fish community structure. Only one standard trawl is performed per station visit, but additional, non-standard trawls may be performed to catch fish for chemical samples and pathology exams. For Coastal 2000, some states may use entirely different trawling methods. The variable FTRAWLID is an identifier used to distinguish between different trawls at the same station. Trawling methods must be described in metadata. The variable GEARCODE is used to identify trawling gear unique to a state.

The data table contains the following elements:

Variable Name	Variable Label
STATION	Station Identifier
EVNTDATE	Date of Sampling Event
FTRAWLID	Trawl Identifier
TRLTYPE	Standard or Non Standard Fish Trawl? (S/N)
BEG_LAT	Trawl Beginning Latitude (decimal degrees)
BEG_LNG	Trawl Beginning Longitude (decimal degrees)
END_LAT	Trawl End Latitude (decimal degrees)
END_LNG	Trawl End Longitude (decimal degrees)
FTRL_DUR	Duration of Fish Trawl (minutes:seconds)
FTRL_SPD	Average Speed over Bottom (knots)
FSPECCNT	Number of Unique Species in Trawl
FISHCNT	Number of Individual Fish in Trawl
GEARCODE	Gear Code

FISHSPEC - Fish Counts by Species per Trawl

This data table contains one record for each species of fish caught in a standard trawl, and provides a count of individual fish, and their mean length. The data table contains the following elements:

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event
FTRAWLID	Trawl Identifier
FSCINAME	Fish Species-Scientific Name
TAX_CNT	Number of Fish of this species

FISHPATH - Fish Length and Pathology Data

This data table contains one record for each fish caught and measured. By EMAP conventions, the first 30 individual fish of each species caught in each standard trawl are always measured and examined for gross pathologies. Certain fish caught in non-standard trawls are also measured, if they are included in chemistry samples. Fish from non-standard trawls are saved for specific analytical purposes and not for the purpose of comparing fish community structures.

The FISHPATH data table contains the following elements:

Variable Name	Variable Label
STATION	Station Identifier
EVNTDATE	Date of Sampling Event
FTRAWLID	Trawl Identifier
FSCINAME	Fish Species-Scientific Name
FSEQNUM	Fish Sequence Number
F_LENGTH	Fish Length (mm)
LUMPS	Fish Pathology: Lumps (Y/N)
LUMPLOC	Locations of Lumps
GROWTHS	Fish Pathology: Growths (Y/N)
GRTHLOC	Locations of Growths
ULCERS	Fish Pathology: Ulcers (Y/N)
ULC_LOC	Locations of Ulcers
FINROT	Fish Pathology: Fin Erosion (Y/N)
FROTLOC	Locations of Finrot
GILL_ERO	Fish Pathology: Gill Erosion (Y/N)
GERO_LOC	Gill Erosion Location
GILL_DC	Fish Pathology: Gill Discoloration (Y/N)
GDC_LOC	Gill Discoloration Location

3.3 Lab Data Tables

The following data tables will contain the results of analyses performed in laboratories of samples collected in the field. Lab results are generally reported by sample id. numbers. It is the responsibility of IM personnel to link the sample id's to the correct station, event date, depth, and replicate number wherever necessary.

NUTRNTS - Water Quality-Nutrients Data

This data table contains water quality measurements derived from laboratory analyses of surface, bottom, and mid-water samples collected in the field. There may be multiple records per sampling event for stations, depending on the number of samples collected.

Variable Name	Variable Label
STATION	Station Identifier
EVNTDATE	Date of sampling event
LAYER	Water Layer (SRF, BOT, MID)
REP_NUM	Replicate Number (>1 indicates replicate sample)
ANALTYE	Code to identify analyte measured

CONC	Concentration of analyte
UNIT	Unit of Measure
MDL	Method Detection Limit
QACODE	QA Qualifier
LABCODE	Laboratory Identification Code

The following analyte codes are used to define the measurements in this data table.

ANALYTE	Description	Core Indicator
SI	Dissolved Silica	Y
NH4	Dissolved Ammonia	Y
NO23	Dissolved Nitrite and Nitrate	Y
NO2	Dissolved Nitrite	Y
PO4F	Dissolved Orthophosphate	Y
TSS	Total Suspended Solids	Y
CHLA	Chlorophyll a	Y
PON	Particulate Organic Nitrogen	N
TDN	Total Dissolved Nitrogen	N
TDP	Total Dissolved Phosphorous	N
PHAE	Phaeophytin	N
PHOSP	Total Particulate Phosphorous	N
POC	Particulate Organic Carbon	N

More analyte codes may be added to this list if the need arises. Analytes must be clearly described in metadata text.

SEDGRAIN - Sediment Grain Size Data

This data table contains grain size data from the composite samples collected for sediment chemistry analysis. There is one record per event, for events where sediment samples were collected.

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event
REP_NUM	Replicate Number (>1 indicates replicate sample)
SAND	Sand Content (%)
SILTCLAY	Silt/Clay Content (%)
MOISTURE	Moisture Content (%)

TOC	Total Organic Carbon (%)
QACODE	QA Qualifier
LABCODE	Laboratory Identification Code

TOXICITY - Sediment Toxicity and Microtox Test Data

This data table contains summary results from two sediment toxicity tests: an *Ampelisca* survival test and a Microtox test. Tests are performed on extracts from one composite sediment sample

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event
REP_NUM	Replicate Number (>1 indicates replicate sample)
SRVPCCON	<i>Ampelisca</i> Survival as % of Control
SRVPC_SG	<i>Ampelisca</i> Survival- Stat. Significance
ATOX_SIG	<i>Ampelisca</i> Survival- Significance
EC50_MC	Microtox Moisture Corrected Mean EC50 (%)
MTOX_SIG	Microtox Test Significance
QACODE	QA Qualifier
LABCODE	Laboratory Identification Code

SEDCHEM - Sediment Chemistry Data

This data table contains sediment chemistry measures. There are multiple records for each event- one record for each concentration measured. Three variables make up the unique identifier for records in this data table: STATION, EVNTDATE, and ANALYTE. The BATCH_ID variable identifies the “batch” of samples that were analyzed in the laboratory together. This identifier is used to link sample data to QA data associated with the batch, that is found in the CHEM_QA table.

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event
BATCH_ID	Laboratory Batch Name
ANALYTE	Code for Analyte Measured
CONC	Concentration of Analyte in Sample
CHMUNITS	Concentration Units of Measure
MDL	Method Detection Limit

QACODE	QA Qualifier
LABCODE	Laboratory Identification Code

CHEM_QA - Chemistry QA Data

This data table contains QA sample data from chemistry analysis sets. These data are not associated with a specific station or event. Instead, they relate to batches of chemistry samples analyzed together in a laboratory. The batch is identified by the variable BATCH_ID. These data can be joined to the data in the SEDCHEM on the BATCH_ID, and are used to assess and document the accuracy of those data.

Variable Name	Variable Label
BATCH_ID	Laboratory Batch Name
SAMPTYPE	QA Sample Type
ANALYTE	Code for Analyte Measured
CONC	Concentration of Analyte in Sample
CHMUNITS	Concentration Units of Measure
MDL	Method Detection Limit
QACODE	QA Qualifier
LCMNAME	Lab Control Material Name
LABCODE	Laboratory Identification Code

The following SAMPTYPE codes will be used to define QA samples.

SAMPTYPE	Description	Unit of Measure
LRB	Lab Reagent Blank	varies
LCM	Lab Control Material	ug/g or ng/g Dry W't
LCMPR	Lab Control Material % Rec.	Percent Recovery
LF1	Lab Spiked Sample- 1st Member	ug/g or ng/g Dry W't
LF1PR	Lab Spiked Sample- 1st Mem. % Rec.	Percent Recovery
LF2	Lab Spiked Sample- 2nd Member	ug/g or ng/g Dry W't
LF2PR	Lab Spiked Sample- 2nd Mem. % Rec.	Percent Recovery
MSDRPD	Rel % Difference: LF1 to LF2	Percent
LFB	Lab Fortified Blank	Percent Recovery
LSFPR	Lab Spiked Sample % Rec.	Percent Recovery
LDRPD	Lab Duplicate Relative % Diff.	Percent

More codes may be added as the need arises.

BEN_ABUN - Benthic Abundance Data

This data table contains benthic abundance measurements from up to three grab samples per event.. Like the Sediment Chemistry data table, it is in a vertical format. It contains one record for each taxa found per grab. Four fields are needed to uniquely identify a record: STATION, EVNTDATE, BENGGRAB, and TAXNAME.

The TAXNAME identifies the benthic taxa- this name can be used to look up taxonomic information for each taxa in the BEN_TAXA table. States are required to supply full Scientific names to identify benthic taxa. ABUNDANC is expressed as number of organisms per square meter of sampled area. Note that different states may use different size grab samplers and may have to extrapolate to estimate this number.

The variable ID_LEVEL describes the level at which the organism was identified (SPECIES, GENUS, FAMILY, etc.)

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of sampling event
BENGGRAB	Grab Associate with Infauna Sample (# per sq. meter)
TAXNAME	Taxa Name
ABUNDANC	Species Abundance in Sample (#)
ID_LEVEL	Level of Taxonomic ID
QACODE	QA Qualifier
LABCODE	Laboratory Identification Code

TISSCHEM - Fish and crustacean tissue chemistry data.

The data table contains one record for each chemical measured in each tissue sample analyzed. Composite samples are made up of approximately five fish or crustaceans of the same species, caught at the same station.

TAXNAME identifies the species of the fish or crustacean analyzed. Tissue type codes identify the type of tissue analyzed. Possible codes are: FW (fish whole body) FE (edible fish fillet). More codes may be added if different tissue types are analyzed.

The TISSCHEM data table contains the following elements:

Variable Name	Variable Label
STATION	Coastal 2000 Station Name
EVNTDATE	Date of Sampling Event

SAMPTYPE	Sample Type (Identifies QA Samples)
BATCH_ID	Laboratory Batch Name
TAXNAME	Scientific name of organisms analyzed
TISUTYPE	Tissue Type Code
NUM_HOM	Number of Individuals in Homogenate
MN_WGHT	Mean Weight of Individuals in Homogenate
MN_SIZE	Mean length (fish) or width (Crabs) in Homogenate (mm)
WETWGHT	Wet Weight of Sample Analyzed (g)
PCTMOIST	Percent Moisture of sample Analyzed (%)
PCTLIPID	Percent Lipid Content of Tissue Examined
ANALYTE	Code for Analyte Measured
CONC	Concentration of Analyte in Sample
CHMUNITS	Concentration Units of Measure
MDL	Method Detection Limit
QACODE	QA Code
LABCODE	Laboratory Identification Code

3.4 Lookup Tables

These tables contain information for coded variables used in the data tables. The code lists developed for the MAIA project will be used as a basis for the Coastal 2000 tables. These tables are listed in Appedix A. New codes can be added to the tables as needed.

ANALYTES - Chemical Analyte Code Table

This look-up table provides chemical names and CAS numbers (where available) for chemicals identified in the TISSCHEM file by ANALYTE. It contains the following elements:

Variable Name	Variable Label
ANALYTE	Chemical Analyte Code
CHEMNAME	Full Chemical Name
CASNUM	CAS Number

FTAXON- Fish Taxon Code Table

This look-up table provides the common name and scientific name of fish species identified in the fish community datasets. It contains the following elements:

Variable Name	Variable Label
FSCINAME	Fish Species-Scientific Name

FCOMNAME	Fish Species-Common Name
TSN	ITIS Taxonomic Serial Number

BEN_TAXA- Benthic Taxonomy Code Table

This look-up table provides taxonomic information for benthic taxa identified in the BEN_ABUN dataset. It contains the following elements:

Variable Name	Variable Label
TAXNAME	Taxa Name
TSN	ITIS Taxonomic Serial Number
PHYLUM	Phylum Name
CLASS	Class Name
ORDER	Order Name
FAMILY	Family Name
GENUS	Genus Name
SPECIES	Species Name

3.5 Notes on the database design

These data tables can be merged on key fields. All except the QA and Code tables contain the field STATION, a station identifier which can be linked to the STATIONS table to find descriptive and locational data for the sampling station. All tables containing measurement data also contain the field EVNTDATE, which identifies the date the samples were collected. The two fields STATION and EVNTDATE should be used to merge tables.

Records in the SEDCHEM data table may be joined to the ANALYTES table on the field ANALYTE to get a descriptive name and the CAS number for analytes measured.

Records in BEN_ABUN can be joined to the BEN_TAXA table on the field TAXNAME to get taxonomic information on benthic organisms.

All tables with measurement data contain the field QACODE. This field is blank if there are no quality assurance issues associated with the data. The field may contain one or more codes to document QA issues. Data with significant QA problems will be excluded from this database. All data will be reviewed and deemed acceptable for Coastal 2000 analyses. QA qualifier codes are described in detail in the metadata files.

4. Formatting and Transfer of ASCII data files

States will supply several types of data to the NEIMC in ASCII data files. If the states do not choose to use the regional field computer system, they can supply field data in ASCII format. If they do not use the national contract laboratories to analyze samples, they will supply lab data in ASCII format. All data other than the core indicators that will be stored at the NEIMC should also be supplied in ASCII data files.

Field and Lab data that make up core indicators should be formatted to match the structure of the SAS data tables which they will be read into as closely as possible. Any additions to the predefined table structure should be discussed with NEIMC personnel prior to data submission.

The guiding principals of formatting ASCII data for submission to the NEIMC are:

- 1) data should be in fixed format or comma delimited ASCII files
- 1) data files should be rectangular in structure, ie. a simple structure of rows and columns.
- 3) data files should be checked for accuracy and completeness before sending to the NEIMC.
- 4) data will be accompanied by comprehensive documentation.
- 5) data files should have a header section before the data records begin that describes the file and all data elements.

An example of an ASCII data file is presented below. The first 18 lines make up a header that describe the data. The header records are followed by ASCII data in comma separated value format. These data can easily be read into SAS or Excel.

```
Dataset:      Sediment Characteristics Data
File Name:    sedgrain.dat
Date Created: 02/11/00
# Variables:  8
# Header Records: 18
# Data Records: 387
Delimiter:    ,
Missing Value Symbol: .
```

```
Variables: STATION  Station Name
           EVNTDATE Event Date
           SAND     Sand Content (%)
           SILTCLAY Silt/Clay Content (%)
           MOISTURE  Moisture Content (%)
           TOC      Total Organic Carbon (%)
           LABCODE  Contract/Lab Identifier
```

```
STATION ,EVNTDATE,SAND ,SILTCLAY,MOISTURE,TOC ,LABCODE
MA97-0001,08/25/97,83.55,16.45,33.4,0.69,GRN-1,
MA97-0003,08/26/97,96.76,3.24,25.2,0.13,GRN-1,
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Appendix A: Northeast Coastal 2000 Code lists.

Chemical Analyte Codes- this list provides chemical names and in most cases CAS numbers for chemical analyte codes. Not all of these analytes are required by Coastal 2000 program. More codes will be added to the list if more analytes are measured by different states.

ANALYTE	Chemical Name	CAS Number
ABHC	alpha-Hexachlorohexane	319846
ACENTHE	Acenaphthene	83329
ACENTHY	Acenaphthylene	208968
AG	Silver	7440224
AL	Aluminum	7429905
ALDRIN	Aldrin	309002
ANTHRA	Anthracene	120127
AS	Arsenic	7440382
AVS	Acid volatile sulfide	18496258
BBHC	beta-Hexachlorohexane	319857
BENANTH	Benz (a) anthracene	56553
BENAPY	Benz (a) pyrene	50328
BENEPY	Benz (e) pyrene	192972
BENZOBFL	Benzo (b) fluoranthene	205992
BENZOFL	Benzo (b+k) fluoranthene	56832736
BENZOKFL	Benzo (k) fluoranthene	207089
BENZOP	Benzo (g, h, i) perylene	191242
BIPHENYL	Biphenyl	92524
CD	Cadmium	7440439
CHRYSENE	Chrysene	218019
CISCHL	Alpha-Chlordane	5103719
CR	Chromium	7440473
CU	Copper	7440508
DBHC	delta-Hexachlorohexane	319868
DBT	Dibutyltin	683181
DIBENZ	Dibenz (a, h) anthracene	53703
DIELDRIN	Dieldrin	60571
DIMETH	2, 6-dimethylnaphthalene	581420
ENDOSUI	Endosulfan I	959988
ENDOSUII	Endosulfan II	33213659
ENDOSULF	Endosulfan Sulfate	1031078
ENDRIN	Endrin	72208
ENDRINA	Endrin Aldehyde	7421934
ENDRINK	Endrin Ketone	53494705
FE	Iron	7439896
FLUORANT	Fluoranthene	206440
FLUORENE	Fluorene	86737
HEPTACHL	Heptachlor	76448
HEPTAEPO	Heptachlor epoxide	1024573

HEXACHL	Hexachlorobenzene	118741
HG	Mercury	7439976
INDENO	Indeno (1,2,3-c,d)pyrene	193395
LINDANE	Lindane (gamma-BHC)	58899
MBT	Monobutyltin	1118463
MENAP1	1-methylnaphthalene	90120
MENAP2	2-methylnaphthalene	91576
MEPHEN1	1-methylphenanthrene	832699
MIREX	Mirex	2385855
MN	Manganese	7439965
NAPH	Naphthalene	91203
NI	Nickel	7440020
OPDDD	2,4'-DDD	53190
OPDDE	2,4'-DDE	3424826
OPDDT	2,4'-DDT	789026
OXYCHL	Oxychlorthane	26880488
PB	Lead	7439921
PCB101	2,2',4,5,5'-pentachlorobiphenyl	37680732
PCB105	2,3,3',4,4'-pentachlorobiphenyl	32598144
PCB118	2,3',4,4',5-pentachlorobiphenyl	31508006
PCB128	2,2',3,3',4,4'-hexachlorobiphenyl	38380073
PCB138	2,2',3,4,4',5'-hexachlorobiphenyl	35065282
PCB153	2,2',4,4',5,5'-hexachlorobiphenyl	35065271
PCB170	2,2',3,3',4,4',5-heptachlorobiphenyl	35065306
PCB18	2,2',5-trichlorobiphenyl	37680652
PCB180	2,2',3,4,4',5,5'-heptachlorobiphenyl	35065293
PCB187	2,2',3,4',5,5',6-heptachlorobiphenyl	52663680
PCB195	2,2',3,3',4,4',5,6-octachlorobiphenyl	52663782
PCB206	2,2',3,3',4,4',5,5',6-nonachlorobiphenyl	40186729
PCB209	decachlorobiphenyl	2051243
PCB28	2,4,4'-trichlorobiphenyl	7012375
PCB44	2,2',3,5'-tetrachlorobiphenyl	41464395
PCB52	2,2',5,5'-tetrachlorobiphenyl	35693993
PCB66	2,3',4,4'-tetrachlorobiphenyl	32598100
PCB8	2,4'-dichlorobiphenyl	34883437
PERYLENE	Perylene	198550
PHENANTH	Phenanthrene	85018
PPDDD	4,4'-DDD	72548
PPDDE	4,4'-DDE	72559
PPDDT	4,4'-DDT	50293
PYRENE	Pyrene	129000
SB	Antimony	7440360
SE	Selenium	7782492
SEM_CD	SEM- Cadmium	
SEM_CU	SEM- Copper	
SEM_NI	SEM- Nickel	
SEM_PB	SEM- Lead	
SEM_ZN	SEM- Zinc	

SI	Silicon	7440213
SN	Tin	7440315
TBT	Tributyltin	1461229
TETBT	Tetrabutyltin	1461252
TL	Thallium	7440280
TNONCHL	Trans-Nonachlor	39765805
TOC	Total organic carbon	14762744
TRIMETH	2,3,5-trimethylnaphthalene	2245387
ZN	Zinc	7440666

Fish Taxonomy Code List- this list provides Scientific Names and Common Names and ITIS Taxonomic Serial Numbers for fish species from the Mid-Atlantic region. Records for more species will be added as fish catches for the Coastal 2000 project are reported. States reporting fish species not on this list should provide these data.

Scientific Name	Common Name	TSN
ALOSA PSEUDOHARENGUS	ALEWIFE	161706
ANCHOA HEPSETUS	STRIPED ANCHOVY	161838
ANCHOA MITCHILLI	BAY ANCHOVY	161839
BAIRDIELLA CHRYSOURA	SILVER PERCH	169259
BREVOORTIA TYRANNUS	ATLANTIC MENHADEN	161732
CENTROPRISTIS STRIATA	BLACK SEA BASS	167687
CHAETODIPTERUS FABER	ATLANTIC SPADEFISH	169539
CHILOMYCTERUS SCHOEPPFI	STRIPED BURRFISH	173384
CHLOROSCOMBRUS CHRYSURUS	ATLANTIC BUMPER	168670
CYNOSCION REGALIS	WEAKFISH	169241
CYPRINUS	CARP	163344
CYPRINUS CARPIO	COMMON CARP	163343
DASYATIS AMERICANA	SOUTHERN STINGRAY	160951
DASYATIS SAY	BLUNTNOSE STINGRAY	160954
DOROSOMA CEPEDIANUM	GIZZARD SHAD	161737
ETROPUS MICROSTOMUS	SMALLMOUTH FLOUNDER	172730
EUCINOSTOMUS ARGENTEUS	SPOTFIN MOJARRA	169015
GYMNURA ALTAVELA	SPINY BUTTERFLY RAY	160961
GYMNURA MICRURA	SMOOTH BUTTERFLY RAY	160962
ICTALURUS CATUS	WHITE CATFISH	164037
ICTALURUS FURCATUS	BLUE CATFISH	163997
ICTALURUS NEBULOSUS	BROWN BULLHEAD	164043
ICTALURUS PUNCTATUS	CHANNEL CATFISH	163998
LEIOSTOMUS XANTHURUS	SPOT	169267
LEPOMIS GIBBOSUS	PUMPKINSEED	168144
MENTICIRRHUS AMERICANUS	SOUTHERN KINGFISH	169274
MENTICIRRHUS SAXATILIS	NORTHERN KINGFISH	169276
MICROPOGONIAS UNDULATUS	ATLANTIC CROAKER	169283
MORONE AMERICANA	WHITE PERCH	167678
MORONE SAXATILIS	STRIPED BASS	167680
OPSANUS TAU	OYSTER TOADFISH	164423
ORTHOPRISTIS CHRYSOPTERA	PIGFISH	169077

PARALICHTHYS DENTATUS	SUMMER FLOUNDER	172735
PEPRILUS ALEPIDOTUS	HARVESTFISH	172570
PEPRILUS TRIACANTHUS	BUTTERFISH	172567
PERCA FLAVESCENS	YELLOW PERCH	168469
POGONIAS CROMIS	BLACK DRUM	169288
PRIONOTUS CAROLINUS	NORTHERN SEAROBIN	166974
PRIONOTUS EVOLANS	STRIPED SEAROBIN	166975
RAJA EGLANTERIA	CLEARNOSE SKATE	160855
SERIOLA FASCIATA	LESSER AMBERJACK	168690
SPHOEROIDES MACULATUS	NORTHERN PUFFER	173290
STENOTOMUS CHRYSOPS	SCUP	169182
SYMPHURUS PLAGIUSA	BLACKCHEEK TONGUEFISH	173062
SYNGNATHUS FUSCUS	NORTHERN PIPEFISH	166451
SYNODUS FOETENS	INSHORE LIZARDFISH	162376
TAUTOGA ONITIS	TAUTOG	170479
TRINECTES MACULATUS	HOGCHOKER	172982

Benthic Taxonomic Code List- this list provides Scientific Names and ITIS Taxonomic Serial Numbers (TSN) for benthic organisms from the Mid-Atlantic region. The database table contains additional taxonomic information (Phylum, Class, Order, Family, Genus, Species). Name More records will be added to this table as Coastal 2000 benthic abundance data are reported. States reporting benthic taxa not on this list should provide these data.

TAXNAME	TSN
Ablabesmyia	128079
Ablabesmyia annulata	128081
Ablabesmyia peleensis	128113
Ablabesmyia rhamphe	128121
Acanthohaustorius intermedius	93981
Acanthohaustorius millsi	93982
Acanthohaustorius similis	93985
Acteocina canaliculata	76117
Acteon punctostriatus	76083
Actiniaria	52485
Aeginellidae	95474
Aglaophamus verrilli	66052
Aligena elevata	80685
Almyracuma proximoculi	90979
Alpheus heterochaelis	96602
Alpheus normanni	96606
Amakusanthura magnifica	542886
Amnicola	70747
Ampelisca	93321
Ampelisca abdita	93329
Ampelisca vadorum	93330
Ampelisca verrilli	93331
Ampharetidae	67718

<i>Amphiodia atra</i>	157649
Amphipoda	93294
<i>Amphiporus bioculatus</i>	57533
<i>Amphitrite ornata</i>	67902
Ampithoe	93409
<i>Amygdalum papyrium</i>	79529
Anachis	73616
<i>Anachis obesa</i>	73622
<i>Anadara transversa</i>	79340
<i>Ancinus depressus</i>	92334
<i>Ancistrosyllis</i>	65541
<i>Ancistrosyllis hartmanae</i>	65543
<i>Ancistrosyllis jonesi</i>	65544
Ancylidae	76568
<i>Anguinella palmata</i>	155542
<i>Anomia simplex</i>	79798
<i>Anoplodactylus petiolatus</i>	83646
Anthozoa	51938
Anthuridae	92144
Aoridae	93440
<i>Apoprionospio pygmaea</i>	66846
<i>Arabella iricolor</i>	66441
Arabellidae	555681
Araneae	82732
Arcidae	79326
<i>Arcteonais lomondi</i>	68976
Aricidea	66666
<i>Aricidea catherinae</i>	66764
<i>Aricidea cerrutii</i>	204494
<i>Aricidea fragilis</i>	66678
<i>Aricidea suecica</i>	66667
<i>Aricidea taylora</i>	66684
<i>Aricidea wassi</i>	66673
<i>Asabellides oculata</i>	67786
Ascidacea	158854
<i>Astarte castanea</i>	80801
<i>Astyris lunata</i>	73542
<i>Aulodrilus limnobius</i>	68682
<i>Aulodrilus pigueti</i>	68680
<i>Aulodrilus plurisetia</i>	68684
Autolytus	65588
Balanus	89600
<i>Balanus improvisus</i>	89622
<i>Barnea truncata</i>	81798
<i>Batea catharinensis</i>	93528
Bathyporeia	93990
<i>Bathyporeia parkeri</i>	193514
<i>Bathyporeia quoddyensis</i>	93991

Bezzia	127778
Bhawania heteroseta	65159
Bittium alternatum	71989
Bivalvia	79118
Boccardiella hamata	67011
Boccardiella ligerica	67012
Bodotriidae	91027
Boonea impressa	75989
Boonea seminuda	75991
Bowmaniella	90259
Bowmaniella floridana	90262
Brachyura	98276
Branchiostoma	159681
Branchiostoma caribaeum	159682
Branchiostoma virginiae	206924
Branchiura sowerbyi	68621
Brania	65759
Brania clavata	65761
Brania swedmarki	193502
Brania wellfleetensis	65762
Bratislavia unidentata	69023
Busycon canaliculatum	74096
Cabira incerta	65565
Caecum regulare	71380
Callianassidae	97732
Callinectes sapidus	98696
Callipallene brevirostris	83670
Capitella capitata	67415
Capitellidae	67413
Caprella penantis	95419
Caprellidae	95375
Carazziella hobsonae	67003
Cardiidae	80865
Carinoma tremaphoros	57429
Cassidinidea lunifrons	92347
Cassidinidea ovalis	92348
Caulleriella	67126
Caulleriella killariensis	67131
Cerapus	93585
Cerapus tubularis	93587
Ceratonereis irritabilis	65874
Ceratopogonidae	127076
Cerebratulus lacteus	57455
Chaetopteridae	67095
Chaetopterus variopedatus	67097
Chaoborus	125904
Chaoborus albatrus	125905
Chaoborus punctipennis	125923

Chiridotea	92637
Chiridotea almyra	92638
Chiridotea arenicola	92639
Chiridotea coeca	92640
Chiridotea nigrescens	92642
Chiridotea tuftsi	92643
Chironomidae	127917
Chironomus	129254
Chironomus stigmaterus	129322
Cirratulidae	67116
Cirrophorus	66708
Cladopelma	129350
Cladotanytarsus	129873
Cladotanytarsus mancus	129881
Climacia	115086
Clinotanypus	127996
Clinotanypus pinguis	127998
Clymenella torquata	67528
Coelotanypus	128010
Coenagrionidae	102077
Columbellidae	73532
Corbicula fluminea	81386
Corbicula manilensis	81386
Corduliidae	102020
Corophium	93589
Corophium acherusicum	93590
Corophium lacustre	93594
Corophium simile	93595
Corophium tuberculatum	93596
Cossura longocirrata	67207
Crangon septemspinosa	97110
Crassispira ostrearum	74901
Crassostrea virginica	79872
Cratena pilata	78713
Crepidula	72619
Crepidula convexa	72624
Crepidula fornicata	72623
Crepidula maculosa	72632
Crepidula plana	72627
Cricotopus bicinctus	128583
Cryptochironomus	129368
Cryptochironomus fulvus	129376
Cryptotendipes	129394
Culicoides	127340
Curculionidae	114666
Cyathura	92148
Cyathura burbancki	92150
Cyathura polita	92149

<i>Cyclaspis varians</i>	91033
<i>Cylichnella bidentata</i>	76140
<i>Cymadusa compta</i>	93430
<i>Cyrenellus fraternus</i>	117092
Decapoda	95599
<i>Demicryptochironomus</i>	129421
<i>Demonax microphthalmus</i>	68222
<i>Dero</i>	68898
<i>Dero obtusa</i>	68907
<i>Diadumene leucolena</i>	52749
<i>Dicrotendipes</i>	129428
<i>Dicrotendipes modestus</i>	129448
<i>Dicrotendipes nervosus</i>	129452
<i>Dicrotendipes simpsoni</i>	193743
<i>Diopatra cuprea</i>	66180
Diptera	118831
<i>Dispio uncinata</i>	66941
<i>Donax fossor</i>	81248
<i>Donax variabilis</i>	81248
<i>Doridella obscura</i>	78439
<i>Dorvillea rudolphi</i>	66523
Dorvilleidae	66478
<i>Drilonereis longa</i>	66426
<i>Dubiraphia</i>	114126
<i>Dyspanopeus sayi</i>	98901
<i>Echinarachnius parma</i>	158016
Echinoidea	157821
<i>Edotea montosa</i>	92624
<i>Edotia triloba</i>	544186
<i>Edwardsia elegans</i>	52489
<i>Elasmopus laevis</i>	93761
Elmidae	114093
Enchytraeidae	68510
<i>Endochironomus</i>	129470
<i>Enoplobranchus sanguineus</i>	68018
<i>Ensis directus</i>	81022
<i>Eobrolgus spinosus</i>	94755
Epitonium	72233
<i>Epitonium greenlandicum</i>	72237
<i>Epitonium rupicola</i>	72249
<i>Epoicocladus</i>	128682
<i>Erichsonella</i>	92617
<i>Erichsonella attenuata</i>	92618
<i>Erichsonella filiformis</i>	92619
<i>Ericthonius brasiliensis</i>	93613
<i>Eteone foliosa</i>	65270
<i>Euceramus praelongus</i>	98081
Eunicidae	66260

Eupleura caudata	73300
Eurypanopeus depressus	98759
Eurythoe	65195
Eusarsiella	84300
Eusarsiella zostericola	205822
Exogone dispar	65722
Fargoa bushiana	75997
Ferrissia	76569
Gammarus	93773
Gammarus daiberi	93779
Gammarus fasciatus	93780
Gammarus palustris	93782
Gammarus tigrinus	93781
Gastropoda	69459
Gemma gemma	81511
Geukensia demissa	79555
Gillia altilis	70665
Gitanopsis	93397
Glycera	66102
Glycera americana	66106
Glycera dibranchiata	66107
Glyceridae	66101
Glycinde solitaria	66132
Glyptotendipes	129483
Gobiidae	171746
Gomphidae	101664
Goniadidae	66126
Gyptis brevipalpa	65532
Haber speciosus	68746
Haminoea solitaria	76258
Hargeria rapax	92067
Harmothoe extenuata	64509
Harmothoe imbricata	64513
Harnischia	129516
Harpacticoida	86110
Haustoriidae	93959
Havelockia scabra	158181
Hemichordata	158616
Heptageniidae	100504
Heteromastus filiformis	67420
Hexagenia	101537
Hexagenia bilineata	101538
Hexagenia limbata	101552
Hippolyte pleuracantha	96750
Hirudinea	69290
Hobsonia florida	97314
Holothuroidea	158140
Hutchinsoniella macracantha	83682

Hydrobia	70494
Hydrobia truncata	70495
Hydrobiidae	70493
Hydroides	68281
Hydroides dianthus	68282
Hydroides protulicola	68283
Hypereteone heteropoda	65266
Ilyanassa obsoleta	74169
Ilyanassa trivittata	74170
Ilyodrilus templetoni	68662
Ischadium recurvum	79561
Ischyroceridae	94143
Isochaetides freyi	68810
Jassa falcata	94171
Jassa marmorata	94172
Kiefferulus	129522
Kurtziella cerina	74806
Kurtziella limonitella	74807
Laeonereis culveri	65965
Laevapex fuscus	76577
Leitoscoloplos	66653
Leitoscoloplos fragilis	66656
Leitoscoloplos robustus	66600
Lembos smithi	93458
Lepidactylus dytiscus	93998
Lepidametria commensalis	64702
Lepidonotus sublevis	64610
Leptocheirus	93485
Leptocheirus plumulosus	93486
Leptosynapta tenuis	158432
Leucon americanus	90790
Libinia dubia	98454
Limnodrilus	68638
Limnodrilus cervix	68652
Limnodrilus hoffmeisteri	68639
Limnodrilus udekemianus	68644
Limulus polyphemus	82703
Lineidae	57443
Lipinella	129531
Listriella	94212
Listriella barnardi	94213
Listriella clymenellae	94214
Littoridinops tenuipes	70528
Loimia medusa	68015
Luconacia incerta	95440
Lumbrineridae	66335
Lumbrineris tenuis	66351
Lyonsia hyalina	81926

<i>Lysianopsis alba</i>	94466
<i>Macoma</i>	81033
<i>Macoma baltica</i>	81052
<i>Macoma mitchelli</i>	81054
<i>Macoma tenta</i>	81055
<i>Macroclymene zonalis</i>	67632
Mactridae	80942
<i>Magelona</i>	67043
Majidae	98417
Maldanidae	67515
<i>Malmgrenia lunulata</i>	64518
<i>Manayunkia aestuarina</i>	68171
<i>Mancocuma stellifera</i>	91030
<i>Marginella apicina</i>	74399
<i>Marphysa sanguinea</i>	66301
Mediomastus	67438
<i>Mediomastus ambiseta</i>	67439
<i>Mediomastus californiensis</i>	67440
<i>Melanella</i>	72440
<i>Melinna</i>	67762
<i>Melinna maculata</i>	67766
<i>Melita</i>	93806
<i>Melita nitida</i>	93812
<i>Mercenaria mercenaria</i>	81496
<i>Microdeutopus gryllotalpa</i>	93477
<i>Microphiopholis atra</i>	-1063
<i>Microphthalmus</i>	65476
<i>Microphthalmus szcelkowi</i>	65477
<i>Microprotopus raneyi</i>	94122
<i>Microtendipes pedellus</i>	129541
<i>Micrura leidy</i>	57477
<i>Mitrella lunata</i>	73552
<i>Molanna</i>	116474
<i>Molgula arenata</i>	159576
<i>Molgula manhattensis</i>	159557
<i>Monoculodes</i>	94519
<i>Monoculodes edwardsi</i>	94539
<i>Monopylephorus</i>	68722
<i>Monopylephorus rubroniveus</i>	68728
Montacutidae	80650
<i>Monticellina dorsobranchialis</i>	67152
<i>Mucrogammarus mucronatus</i>	-1069
<i>Mulinia lateralis</i>	80959
<i>Musculium</i>	81427
<i>Musculium transversum</i>	81428
<i>Mya arenaria</i>	81692
<i>Mysella planulata</i>	80661
Mysidacea	89807

Mysidae	89856
Mysidopsis	90138
Mysidopsis almyra	90141
Mysidopsis bigelowi	90139
Mysidopsis furca	90143
Mytilopsis leucophaeata	81335
Nais simplex	68957
Nais variabilis	68959
Nanocladius	128844
Nanocladius balticus	128866
Nanocladius crassicornus	128852
Nassarius trivittatus	74109
Nassarius vibex	74107
Naticidae	72878
Neanthes arenaceodentata	65895
Neanthes succinea	65918
Nematoda	563956
Nemertea	57411
Neomysis americana	90062
Neopanope sayi	98775
Nephtyidae	66010
Nephtys	66011
Nephtys bucera	66027
Nephtys incisa	66028
Nephtys picta	66030
Nereididae	65870
Nereiphylla fragilis	65336
Nereis	65902
Nereis acuminata	65926
Nereis riisei	65927
Nereis succinea	65918
Neumania	83103
Neverita duplicata	72961
Nilothauma	129548
No organisms present	-100
Notocirrus spiniferus	66450
Notomastus	67423
Nucula proxima	79132
Nudibranchia	78156
Odonata	101593
Odostomia	75447
Odostomia engonia	75504
Oecetis	116607
Oecetis inconspicua	116613
Oedicerotidae	94489
Ogyrides alphaerostris	96737
Oligochaeta	68422
Onuphidae	66157

Ophiuroidea	157325
Orbiniidae	66570
Ostracoda	84195
Ovalipes	98710
Ovalipes ocellatus	98714
Owenia fusiformis	67647
Oxyurostylis smithi	90923
Pagastiella	129561
Paguridae	97774
Pagurus	97775
Pagurus longicarpus	97807
Pagurus pollicaris	97809
Paleanotus heteroseta	65152
Panopeus herbstii	98778
Paracaprella pusilla	95435
Paracaprella tenuis	95434
Paracereis caudata	92290
Parachironomus	129564
Parachironomus directus	129573
Paracladopelma	129597
Parahaustorius	94003
Parahaustorius attenuatus	94004
Parahaustorius longimerus	94006
Parahesione luteola	65493
Paralauterborniella	129616
Parametopella cypris	94927
Paranais	68862
Paranais litoralis	68863
Paraonidae	66659
Paraonis fulgens	66697
Parapionosyllis longicirrata	65824
Paraprionospio pinnata	66937
Parasterope pollex	84233
Parougia caeca	66553
Parvilucina multilineata	80388
Pectinaria gouldi	67709
Pentamera pulcherrima	158230
Periploma fragile	81943
Perophora viridis	159138
Petricola pholadiformis	81627
Phascolion strombi	154734
Pherusa affinis	67247
Phoronis	155462
Phoronis architecta	155466
Phoronis psammophila	155467
Photis	94061
Photis pugnator	94077
Photis reinhardi	94063

Phoxocephalidae	94633
Phyllodoce	65359
Phyllodoce arenae	65366
Phyllodocidae	65228
Phylocentropus	115361
Piguetiella michiganensis	68994
Pilargidae	65540
Pinnixa	98993
Pinnixa chaetopterana	98998
Piromis roberti	67272
Pisidium	81400
Pisidium compressum	81406
Pista	67940
Pista palmata	67947
Pista quadrilobata	67951
Placobdella papillifera	69364
Pleustidae	94768
Pleusymtes glaber	94797
Podarke obscura	65517
Podarkeopsis levifuscina	555698
Polinices duplicatus	72918
Polycirrus	67959
Polycirrus eximius	67963
Polydora	66789
Polydora colonia	66808
Polydora commensalis	66800
Polydora cornuta	204501
Polydora ligni	66801
Polydora socialis	66791
Polydora websteri	66802
Polygordius	68419
Polynoidae	64397
Polypedilum	129657
Polypedilum convictum	129671
Polypedilum halterale	129684
Polypedilum illinoense	129686
Polypedilum scalaenum	129708
Polypedilum tritum	129719
Potamilla reniformis	68136
Prionospio	66838
Prionospio heterobranchia	66843
Prionospio perkinsi	66854
Pristinella jenkiniae	69030
Pristinella osborni	69026
Probezzia	127729
Proceraea cornuta	65590
Procladius	128277
Procladius bellus	128285

<i>Procladius sublettei</i>	128316
<i>Protohaustorius deichmannae</i>	94009
<i>Protohaustorius wigleyi</i>	94010
<i>Pseudeurythoe ambigua</i>	65177
<i>Pseudeurythoe paucibranchiata</i>	65175
<i>Pseudochironomus</i>	129851
<i>Pseudohaustorius</i>	94013
<i>Pseudohaustorius caroliniensis</i>	94015
<i>Pseudunciola obliquua</i>	93640
<i>Ptilanthura tenuis</i>	92155
<i>Ptilanthura tricarina</i>	92156
<i>Pyramidella</i>	75947
<i>Pyramidella candida</i>	75948
<i>Pyramidella crenulata</i>	75950
<i>Pyramidellidae</i>	75446
<i>Quistradrilus multisetosus</i>	68794
<i>Rangia cuneata</i>	80962
<i>Rheotanytarsus</i>	129952
<i>Rhepoxynius</i>	94727
<i>Rhepoxynius epistomus</i>	94728
<i>Rhepoxynius hudsoni</i>	94730
<i>Rhithropanopeus harrisii</i>	98790
<i>Rhynchocoela</i>	57411
<i>Rictaxis punctostriatus</i>	76083
<i>Sabellaria vulgaris</i>	67671
<i>Sabellidae</i>	68076
<i>Saccoglossus kowalevskii</i>	158626
<i>Sayella chesapeakea</i>	70946
<i>Scolecopides viridis</i>	66861
<i>Scolelepis</i>	66942
<i>Scolelepis bousfieldi</i>	66944
<i>Scolelepis texana</i>	66949
<i>Scoletoma tenuis</i>	66351
<i>Scoloplos rubra</i>	66603
<i>Serpulidae</i>	68232
<i>Sialis</i>	115002
<i>Sialis mohri</i>	115010
<i>Sigalion arenicola</i>	65094
<i>Sigambra bassi</i>	65554
<i>Sigambra tentaculata</i>	65552
<i>Sipuncula</i>	154520
<i>Solemya velum</i>	79316
<i>Sphaeriidae</i>	112737
<i>Sphaerium</i>	81391
<i>Sphaerosyllis taylori</i>	65747
<i>Spio pettiboneae</i>	66870
<i>Spiochaetopterus costarum</i>	67107
<i>Spiochaetopterus costarum oculus</i>	67108

<i>Spiochaetopterus oculatus</i>	67110
Spionidae	66781
<i>Spiophanes bombyx</i>	66897
Spirorbis	68248
<i>Spisula solidissima</i>	80944
<i>Squilla empusa</i>	99143
Stenelmis	114095
<i>Stenothoe georgiana</i>	94937
<i>Stenothoe minuta</i>	94936
Sthenelais	65082
<i>Sthenelais boa</i>	65084
Stictochironomus	129785
<i>Stictochironomus devinctus</i>	129790
<i>Streblospio benedicti</i>	66939
<i>Streptosyllis arenae</i>	65818
<i>Streptosyllis pettiboneae</i>	65822
<i>Stylochus ellipticus</i>	54089
Syllidae	65587
<i>Syllides verrilli</i>	65812
Synaptidae	158425
<i>Synchelidium americanum</i>	94567
Tagelus	81271
<i>Tagelus divisus</i>	81274
<i>Tagelus plebeius</i>	81272
Tanaidacea	91061
<i>Tanaissus psammophilus</i>	91573
Tanypus	128324
<i>Tanypus neopunctipennis</i>	128329
<i>Tanystylum orbiculare</i>	83618
Tanytarsini	129872
<i>Tanytarsus</i>	129978
Tellina	81074
<i>Tellina agilis</i>	81088
Tellinidae	81032
Terebellidae	67899
Thalassinidea	96105
<i>Tharyx acutus</i>	67147
<i>Tharyx annulosus</i>	67148
<i>Travisia parva</i>	67372
<i>Tribelos jucundus</i>	129827
Trichoptera	115095
Tubificidae	68585
Tubificoides	68687
<i>Tubificoides heterochaetus</i>	68594
<i>Tubificoides wasselli</i>	68692
Tubulanus	57416
Turbellaria	53964
Turbonilla	75676

Turbonilla interrupta	75687
Unciola	93629
Unciola serrata	93633
Unionicola	83073
Unionidae	79913
Upogebia affinis	98209
Urosalpinx cinerea	73264
Valvata	70346
Valvata tricarinata	70354
Xanthidae	98748
Xenanthura brevitelson	92162
Yoldia limatula	79273

Appendix B: Metadata Questionnaires

Coastal 2000 Northeast Region Metadata Questionnaire.

States supplying data to Coastal 2000 are requested to supply the following information pertaining to data collected for the program

1 Field Sampling Methods.

V: Personnel Information. Names of captains, field coordinators

1.2 Platform and gear used. Describe the following

1.2.1 Boat (type and length)

1.2.2 Navigational Instruments (make, models, calibration technique)

1.2.3 Water Monitoring instruments (make, models, calibration techniques)

Describe deployment methods and QA protocols.

1.2.4 Nutrients Sample Processing- describe filtration and sample handling.

1.2.5 Sediment “Grab” Sampler (make, model, area sampled)

Describe deployment methods and QA protocols.

1.2.6 Fishing Gear. Describe trawling methods and QA protocols.

1.3 References. Provide references to relevant field methods manuals.

2 Laboratory Methods. For lab results data, the following information is requested. If national Coastal 2000 contract lab was used to analyze samples, no information is needed from the state.

2.1 Nutrients Analysis

2.1.1 Personnel Information. Names chief lab technicians responsible for samples

2.1.2 Instruments used. List all analytes measured, with a brief description of the instrument and lab procedures used.

2.1.3 Detection Limits. Provide for all analytes.

2.1.3 References. Provide references to lab methods manuals or SOP's.

2.2 Sediment Grain Size

2.2.1 Personnel Information. Names chief lab technicians responsible for samples

2.2.2 Instruments used.

2.2.3 References. Provide references to lab methods manuals or SOP's.

2.3 Sediment Toxicity.

2.3.1 Personnel Information. Names chief lab technicians responsible for samples

2.3.2 Ampellisca test. Describe test protocol briefly

2.3.3 Microtox Test. Describe test protocol briefly

2.3.4 References. Provide references to lab methods manuals or SOP's.

2.4 Sediment Chemistry

2.4.1 Personnel Information. Names chief lab technicians responsible for samples

2.4.2 Instruments used. List all analytes measured, with a brief description of the instrument and lab procedures used.

2.4.3 Detection Limits. Provide for all analytes.

2.4.4 References. Provide references to lab methods manuals or SOP's.

2.5 Tissue Chemistry

2.5.1 Personnel Information. Names chief lab technicians responsible for samples

2.5.2 Composite preparation. Describe how tissue composite samples were created.

2.5.3 Lab Instruments. List all analytes measured, with a brief description of the instrument and lab procedures used.

2.5.4 Detection Limits. Provide for all analytes.

2.5.3 References. Provide references to lab methods manuals or SOP's.

Appendix C: Field Data Sheets

Final copies of datasheets will be attached.